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1 **Molecular Epidemiology of *Pseudomonas aeruginosa* in Intensive Care Units (ICUs)**
2 **over a 10-Year Period (1998-2007)**

3

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14

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Abstract

Pseudomonas aeruginosa is one of the leading nosocomial pathogens in ICUs. The source of this microorganism can be either endogenous or exogenous. The proportion of cases due to transmission is still debated, and its elucidation is important for implementing appropriate control measures. In order to understand the relative importance of exogenous *versus* endogenous sources of *P. aeruginosa*, molecular typing was performed on all available *P. aeruginosa* isolated from ICU clinical and environmental specimens in 1998, 2000, 2003, 2004 and 2007. Patient samples were classified according to their *P. aeruginosa* genotypes in 3 categories: A) identical to faucet, B) identical to at least one other patient sample and not found in faucet, and C) unique genotype. Cases in categories A and B were considered as possibly exogenous, and cases in category C as possibly endogenous.

A mean number of 34 cases /1000 admissions per year were found to be colonized or infected by *P. aeruginosa*. Higher levels of faucet contamination were correlated with a higher number of cases in category A. The number of cases in category B varied from 1.9 to 20 cases/1000 admissions. This number exceeded 10/1000 admissions on 3 occasions and was correlated with an outbreak in one occasion. Number of cases considered as endogenous (category C) was stable and independent from the number of cases in categories A and B.

Repeated molecular typing over time allowed to document variations in the epidemiology of *P. aeruginosa* in ICU patients susceptible to require continuous adaptation of infection control measures.

45 **Introduction**

46 *Pseudomonas aeruginosa* is a ubiquitous environmental bacterium with minimal requirements
47 for survival and a remarkable ability to adapt to a variety of environments. Although humans
48 are frequently in contact with *P. aeruginosa*, the species only colonizes the normal human
49 host intermittently. In contrast, it was found to be part of the intestinal flora of 2,6 to 24,0 %
50 of hospitalized patients [1]. The importance of *P. aeruginosa* as an opportunistic pathogen
51 stems from its ability to activate well-adapted phenotypes under environmental stress and to
52 persist in adverse conditions such as in the presence of antibiotic or antiseptic substances. *P.*
53 *aeruginosa* infections occur mostly in immunocompromised hosts and intensive care units
54 (ICUs) patients. In one study, *P. aeruginosa* was responsible for 15.6 % of all nosocomial
55 pneumonia cases in medical-surgical ICUs in the USA [2]. It was also found to be the most
56 frequent Gram-negative bacterium recovered from patients with nosocomial pneumonia
57 during ICU surveillance over the last two decades [3]. Furthermore, the overall rate of *P.*
58 *aeruginosa* infections increased rapidly in some patients populations, even if the resistance
59 profiles of *P. aeruginosa* from nosocomial infections remained stable [4].

60 As *P. aeruginosa* is ubiquitous in the environment and is also part of the endogenous flora of
61 hospitalized patients, only studies using powerful molecular typing methods can explore the
62 routes of colonization and/or infection. The identification of the source is only possible
63 through comprehensive screening of the environment and the characterisation of the
64 endogenous flora of patients, which requires significant resources. Molecular typing makes it
65 possible to track the dissemination of specific strains and may facilitate the breaking down of
66 endemic transmission to the level of micro-epidemics [5].

67 The question we wished to address is whether *P. aeruginosa* infections in ICUs patients are
68 mainly due to endogenous or exogenous sources. We previously found that 42 % of ICU
69 patients with *P. aeruginosa* harboured isolates identical to those found in the faucets [6].

70 Following control measures, the number of these patients decreased significantly [7]. These
71 results showed that the epidemiology of *P. aeruginosa* is likely to vary over time. On the one
72 hand, several recent studies using molecular typing in a non-epidemic ICU setting suggested
73 that the major reservoir of *P. aeruginosa* was the endogenous flora of the patients [8,9,10,11].
74 At the same time, other investigators emphasized the role of the environment as a source of
75 colonization and infection in ICUs [12,13]. It is still unclear whether the environment is
76 mainly a passive site where strains are deposited, or whether it plays an active role in patient
77 colonization [14]. In addition, the importance of other patients as exogenous sources should
78 also be taken into consideration, even in periods free of recognized outbreak.
79 The aim of our study was to investigate over a 10-year period the respective roles of
80 exogenous (faucets or other patients) versus endogenous flora as sources of colonization or
81 infection by *P. aeruginosa* in ICU's patients, and to document epidemiological variations
82 over time.

83

84 **Materials and Methods**

85 **Setting.** The University Hospital of Lausanne is a 850-bed tertiary care hospital with a 32-bed
86 adult (medical and surgical) ICUs and a 9-bed pediatric ICU. The ICUs have two distinct
87 water distribution networks: i) medical ICUs (network I) and ii) surgical and pediatric ICUs
88 (network II). Between 2006 and 2007, the medical ICUs were rebuilt and, consequently, all
89 faucets and the water network were changed.

90 **Study population.** Patients were identified through the laboratory database. They were
91 included in the study if they were hospitalized in one of the ICUs and had at least one culture
92 of a clinical specimen positive for *P. aeruginosa*. No routine screening of *P. aeruginosa*
93 carriage was performed. No clinical information was collected in order to distinguish between
94 colonization and infection by *P. aeruginosa*. Epidemiological data (unit and room of

95 hospitalization, dates of admission and discharge) were retrieved from the hospital
96 information system.

97 **Study period.** The study covered a period of ten years, from 1998 through 2007. Molecular
98 and epidemiological analyses were conducted in 1998, 2000, 2003, 2004 and 2007.

99 **Environment sampling.** From 1998 to 2007, 18-56 ICU's faucets were swabbed once to
100 three times a year (most usually twice). To identify the presence of *P. aeruginosa*, swabs were
101 plated on cetrimide agar. All isolates from faucets were saved for molecular typing.

102 **Molecular typing** was performed in 1998, 2000, 2003 and 2004 on clinical and
103 environmental isolates using pulsed-field-gel electrophoresis (PFGE), as described previously
104 [6,7,15]. For the year 2007 samples, we used the recently developed double digest selective
105 label (DDSL) typing technique, which is similar to PFGE (DNA restriction with the same rare
106 cutting enzyme *SpeI*), but based on a much faster and easier simultaneous double
107 digestion/labeling reaction procedure [16]. Indistinguishable isolates (identical bands) and
108 closely related isolates (1-6 band differences) were considered to be of the same genotype.

109 **Epidemiological definitions and classification of cases.** A case was defined as the isolation
110 of a given genotype of *P. aeruginosa* from a clinical specimen obtained during a patient's stay
111 in an ICU. Thus, recovery of isolates having two different genotypes in a single patient
112 yielded two distinct cases. Cases were classified into four categories (A-D) according to
113 molecular typing data. Category A included cases with a *P. aeruginosa* genotype identical to
114 that of an isolate recovered from a faucet during the same year or previously. Category B
115 comprised cases where the genotype was identical to that of an isolate recovered from at least
116 one other patient, but absent in isolates from the faucets. Cases from categories A and B were
117 considered of possible exogenous origin. Category C comprised cases with a unique
118 genotype; these cases were considered of possible endogenous origin. Finally, category D
119 comprised cases for which isolates were not available for typing.

120 Cases which shared the same PFGE/DDSL genotype as isolates from faucets or other patients
121 were defined as belonging to the same "genetic" cluster. Within a genetic cluster, if
122 epidemiological links (see definition thereafter) were found between cases, they were
123 considered as members of an "epidemiological" cluster. Epidemiological links were
124 considered if i) a patient was hospitalized in the same ICU where a faucet was found to be
125 contaminated with the same genotype during the same year, or ii) patients with identical
126 genotypes were hospitalized during overlapping periods in the same ICU.

127

128 **Results**

129 **Study population.** During the 10-year period of the study, the number of admissions in ICUs
130 remained stable (c.a. 2500 admissions per year, Table 1). A mean number of 86 ICU patients
131 per year (34 per 1000 admissions) were observed to be colonized or infected by *P.*
132 *aeruginosa*. The highest incidences were observed in 1998 and 1999 (57.1 and 42.2/1000
133 admissions, respectively), and then decreased below 40/1000 admissions. These incidences
134 were not correlated with the number of clinical specimens received by the laboratory of
135 bacteriology (Table 1; Pearson's test of correlation, $p=0.1995$), excluding a sampling bias.

136 **Environmental contamination.** To measure the colonization rate with *P.aeruginosa* over the
137 10 years, only faucets that were regularly swabbed during the 10 years of the study (12 in the
138 medical ICUs, and 17 in the surgical and pediatric ICUs) were included (Table 1). Results
139 showed a difference of contamination over the years between the two water networks.
140 Whereas <10% of the faucets were contaminated in the surgical and pediatric ICUs, the level
141 of contamination was high in the medical network from 1998 to 2002, but dropped below
142 10% between 2003 and 2005 (no sampling was performed during renovation in 2006 and
143 2007; and an investigation done in 2008 revealed no contamination of the new faucets).

144 **Faucets as a major environmental exogenous source of contamination.** During the first
145 year of investigation (1998), a high number of patients were found to be colonized with a *P.*
146 *aeruginosa* strain identical to a strain found in the faucets (category A, Table 1, Figure). The
147 higher contamination rates of faucets in the medical ICU (25%), compared to the surgical-
148 pediatric ICUs (6.4%), was correlated with a higher number of cases of category A in this
149 ICU (32.6 cases/1000 admissions), compared to the other ICUs (16.8 cases/1000 admissions)
150 (Table 1, Figure). This correlation was also observed in 2000. The number of cases in
151 category A in the medical unit only became similar to that of the surgical and pediatric ICUs
152 in 2003, when the percentage of contaminated faucet dropped below 10%.

153 **Other patients as exogenous source of contamination.** Transmission was suspected when a
154 patient shared a *P. aeruginosa* genotype with at least one other patient. The number of cases
155 of this category varied over the 10 years of surveillance from 1,9 to 20 cases per 1000
156 admissions (category B, Table 1). This number exceeded 10 per 1000 admissions on 3
157 occasions: i) in both units in 1998, ii) in the surgical unit in 2003-4, and iii) in the same unit in
158 2007. In 1998, two large clusters of patients carrying the same *P aeruginosa* genotype (11 and
159 6 patients, respectively; Table 2) were identified, and one outbreak was recognized
160 (epidemiological links between 6 patients in one cluster). Introduction of alcohol-based hand
161 rub seemed to have played a role for the reduction of cases in this category [7]. In 2003, two
162 large clusters of patients carrying the same *P aeruginosa* genotype (13 and 8 patients,
163 respectively) were observed in the surgical unit, reflecting one outbreak due to the emergence
164 and spread of two multi-resistant *P. aeruginosa* strains (susceptible to ≤ 1 of the 4 antibiotic
165 classes used for therapy). This outbreak lasted until 2004. In addition, the present
166 investigation revealed that several patients harbored non multi-resistant strains that were
167 genetically identical to the epidemic multi-resistant strains. During the outbreak of 2003,

168 environmental sources of contamination (transoesophageal probe, siphons) were identified
169 that may explain the high level of cases.

170 **Endogenous source of contamination.** Over the 10 years of the study, the number of cases
171 with a unique *P. aeruginosa* genotype (category C) remained stable and independent from the
172 variations in the number of cases in categories A and B (Table 1 and figure).

173 **Concordance between molecular typing and epidemiological data.** Within category A, 22
174 genetic clusters including 1 to 30 cases were observed. An epidemiological link could not be
175 found for 2 cases in 2 genetic clusters (one patient in each of the 2 genetic clusters was
176 hospitalized in another ICU than the one in the unit with a positive faucet) (Table 2). In 2007,
177 we found 4 patients sharing the same genotype as one found in faucets during previous years,
178 but not recovered from incriminated faucets in 2007. Within category B, 40 genetic clusters
179 comprised of 2 to 13 cases were observed. Epidemiological links were found between all
180 cases in only 8 of these clusters, between some of the cases for 6 other clusters, but not in the
181 remaining 26 clusters (Table 2). During the years 2004 and 2007, when no outbreak was
182 recognized, we observed many small genetic clusters, however without any concordance with
183 epidemiological data (Table 2).

184

185 **Discussion**

186 Over the 10-year period under study, we observed several variations in the epidemiology of *P.*
187 *aeruginosa* in our ICU setting. If endogenous sources appeared to remains stable over time,
188 exogenous sources varied depending on environmental contamination and outbreak
189 occurrence.

190 The relative importance of faucets as an environmental source of *P. aeruginosa* was high in
191 1998 but then decreased as a result of additional infection control measures. Taps
192 contaminated with *P. aeruginosa* had already been shown to serve as a continuous source for

193 transmission [12,17]. However, our study is the first to investigate this contamination over a
194 long period of time. A possible explanation for the observed decrease of contamination is a
195 late effect of the increase in temperature of the water network from 50 to 65°C, a change that
196 occurred in March 2000 [18]. Some experts recommend the use of microfilters in each faucet
197 of the ICU (end-line filtration) [17,19]. Based on our surveillance data showing that cases
198 linked to contaminated faucets have been very rare since 2003, we did not introduce such
199 filters.

200 Although a contaminated faucet can be the source of a cluster of *P. aeruginosa* colonization
201 or infection, alternative hypotheses should be taken into consideration to explain the cases in
202 category A that could not be distinguished in our study. Firstly, the patient him- or herself
203 may be the source of the contamination of a faucet. We have previously shown that such
204 events are probably infrequent [6]. Secondly, a mixed scenario can also occur when a patient
205 contaminated from a faucet becomes the source of patient-to-patient transmission.

206 In our study, patient-to-patient transmission was mainly measured as the number of patients
207 sharing identical *P. aeruginosa* genotypes. To understand this epidemiology, we should not
208 only consider the number of cases, but also the size and the number of genetic clusters. Small
209 clusters probably reflect sporadic patient-to-patient transmissions, whereas larger clusters
210 suggest the contribution of a persisting source, most probably from the environment. A large
211 outbreak due to the emergence and spread of multi-resistant *P. aeruginosa* was observed in
212 2003 and 2004. Molecular investigation enabled us to understand this outbreak, which
213 involved two strains and multiple exogenous sources of contamination (transoesophageal
214 probe, siphons, and other patients). The duration of the outbreak was probably due to the
215 persistence of the epidemic strains in the environment, and to the lack of additional contact
216 measures of ICU patients harboring the epidemic strains that did not show a multi-resistant
217 phenotype. This episode suggests that environmental sources other than faucets should also be

218 suspected when a *P. aeruginosa* outbreak involves several patients. This is in accordance with
219 reports of *P. aeruginosa* outbreaks in the literature always showing the implication of the
220 environment [20,21,22,23,24].

221 Molecular typing results of patients' isolates should disclose unsuspected clusters. Indeed, the
222 axiom of molecular epidemiology is that when two isolates are genetically identical, they
223 should originate from the same chain of transmission [25]. However, even if the population
224 diversity of *P. aeruginosa* is very high, presence of predominant genotypes in patients
225 without epidemiological links should also be taken into consideration. Therefore, to
226 investigate transmission, it is important to analyze the concordance of molecular results with
227 epidemiological data. A low concordance suggests that either the epidemiological link was
228 missing, or that a predominant *P. aeruginosa* genotype was found in epidemiologically
229 unrelated patients. Our definition of epidemiological link relies mainly on the existence of
230 overlapping periods of hospitalization in the same ICU or hospitalizations in the same ICU
231 where a faucet was contaminated with the same genotype. In 2003, the outbreak involving
232 two multi-resistant strains was not restricted to the ICUs (operating theatre, other wards),
233 which strongly suggests exogenous origin of contamination for all patients harboring these
234 genotypes. Similarly, we might have been able to find epidemiological links during the other
235 years, if deeper investigations had been conducted.

236 In conclusion, the epidemiology of *P. aeruginosa* in the ICU setting shows that the
237 contribution of endogenous versus exogenous reservoirs to the colonization and infection of
238 patients varies over time. The major factors contributing to such variations are probably the
239 contamination of the environment, the compliance of health care workers with infection
240 control measures, and the genetics of the pathogen itself. As *P. aeruginosa* is ubiquitous in
241 humid environments and colonizes up to 15% of hospitalized patients, eradication of the
242 reservoir may prove quite difficult. We have demonstrated that, in our setting, this eradication

243 was possible in faucets by increasing the temperature of hot water. In addition, when a multi-
244 resistant strain is repeatedly recovered from patients and from the environment, efforts should
245 be undertaken to achieve eradication of this strain from the environment. However, patient-to-
246 patient transmission was also found to be an important and recurrent cause of exogenous
247 origin. Therefore, additional efforts should be deployed to reinforce standard precautions and
248 specific infection control measures and thus limit the transmission of *P. aeruginosa*.

249

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254

255 **Transparency declaration**

256 Potential conflict of interest. All authors : none

257

258 **References**

- 259 1. Morrison AJ and Wenzel RP. Epidemiology of Infections Due to *Pseudomonas*
260 *aeruginosa*. *Rev Infect Dis* 1984; **6**: S627-S642.
- 261 2. Richards MJ, Edwards JR, Culver DH et al. Nosocomial infections in combined
262 medical-surgical intensive care units in the United States. *Infect Control Hosp*
263 *Epidemiol* 2000; **21**: 510-515.
- 264 3. Gaynes R and Edwards JR. Overview of nosocomial infections caused by gram-negative
265 bacilli. *Clin Infect Dis* 15-9-2005; **41**: 848-854.
- 266 4. Slama TG. Gram-negative antibiotic resistance: there is a price to pay. *Crit Care* 2008;
267 **12 Suppl 4**: S4-

- 268 5. Blanc DS. The use of molecular typing for the epidemiological surveillance and
 269 investigation of endemic nosocomial infections. *Infection, Genetics and Evolution* 2004;
 270 **4**: 193-197.
- 271 6. Blanc DS, Nahimana I, Petignat C et al. Faucet as a reservoir of endemic *Pseudomonas*
 272 *aeruginosa* colonization/infection in intensive care units. *Intensive Care Med* 2004; **30**:
 273 1964-1968.
- 274 7. Petignat C, Francioli P, Nahimana I et al. Exogenous sources of *Pseudomonas*
 275 *aeruginosa* in intensive care unit patients: implementation of infection control measures
 276 and follow-up with molecular typing. *Infect Control Hosp Epidemiol* 2006; **27**: 953-957.
- 277 8. Berthelot P, Grattard F, Mahul P et al. Prospective study of nosocomial colonization and
 278 infection due to *Pseudomonas aeruginosa* in mechanically ventilated patients. *Intensive*
 279 *Care Med* 2001; **27**: 503-512.
- 280 9. Kropec A, Huebner J, Riffel M et al. Exogenous or endogenous reservoirs of
 281 nosocomial *Pseudomonas aeruginosa* and *Staphylococcus aureus* infections in a
 282 surgical intensive care unit. *Int C Med* 1993; **19**: 161-165.
- 283 10. Bonten MJ, Bergmans DC, Speijer H et al. Characteristics of polyclonal endemicity of
 284 *Pseudomonas aeruginosa* colonization in intensive care units. Implications for infection
 285 control. *Am J Respir Crit Care Med* 1999; **160**: 1212-1219.
- 286 11. Valles J, Mariscal D, Cortes P et al. Patterns of colonization by *Pseudomonas*
 287 *aeruginosa* in intubated patients: a 3-year prospective study of 1,607 isolates using
 288 pulsed-field gel electrophoresis with implications for prevention of ventilator-associated
 289 pneumonia. *Intensive Care Med* 2004; **30**: 1768-1775.
- 290 12. Reuter S, Sigge A, Wiedeck H et al. Analysis of transmission pathways of *Pseudomonas*
 291 *aeruginosa* between patients and tap water outlets. *Crit Care Med* 2002; **30**: 2222-2228.

- 292 13. Agodi A, Barchitta M, Cipresso R et al. *Pseudomonas aeruginosa* carriage,
293 colonization, and infection in ICU patients. *Intensive Care Med* 2007; **33**: 1155-1161.
- 294 14. Bonten MJ and Weinstein RA. Transmission pathways of *Pseudomonas aeruginosa* in
295 intensive care units: don't go near the water. *Crit Care Med* 2002; **30**: 2384-2385.
- 296 15. Struelens MJ, Schwam V, Deplano A et al. Genome macrorestriction analysis of
297 diversity and variability of *Pseudomonas aeruginosa* strains infecting cystic fibrosis
298 patients. *J Clin Microbiol* 1993; **31**: 2320-2326.
- 299 16. Terletskiy V, Kuhn G, Francioli P et al. Application and evaluation of double digest
300 selective label (DDSL) typing technique for *Pseudomonas aeruginosa* hospital isolates.
301 *J Microbiol Methods* 2008; **72**: 283-287.
- 302 17. Trautmann M, Bauer C, Schumann C et al. Common RAPD pattern of *Pseudomonas*
303 *aeruginosa* from patients and tap water in a medical intensive care unit. *Int J Hyg*
304 *Environ Health* 2006; **209**: 325-331.
- 305 18. Blanc DS, Carrara P, Zanetti G et al. Water disinfection with ozone, copper and silver
306 ions, and temperature increase to control *Legionella*: seven years of experience in a
307 university teaching hospital. *J Hosp Infect* 2005; **60**: 69-72.
- 308 19. Trautmann M, Halder S, Hoegel J et al. Point-of-use water filtration reduces endemic
309 *Pseudomonas aeruginosa* infections on a surgical intensive care unit. *Am J Infect*
310 *Control* 2008; **36**: 421-429.
- 311 20. Bukholm G, Tannaes T, Kjelsberg AB et al. An outbreak of multidrug-resistant
312 *Pseudomonas aeruginosa* associated with increased risk of patient death in an intensive
313 care unit. *Infect Control Hosp Epidemiol* 2002; **23**: 441-446.
- 314 21. Sanchez-Carrillo C, Padilla B, Marin M et al. Contaminated feeding bottles: the source
315 of an outbreak of *Pseudomonas aeruginosa* infections in a neonatal intensive care unit.
316 *Am J Infect Control* 2009; **37**: 150-154.

- 317 22. Hota S, Hirji Z, Stockton K et al. Outbreak of multidrug-resistant *Pseudomonas*
318 *aeruginosa* colonization and infection secondary to imperfect intensive care unit room
319 design. *Infect Control Hosp Epidemiol* 2009; **30**: 25-33.
- 320 23. Eckmanns T, Oppert M, Martin M et al. An outbreak of hospital-acquired *Pseudomonas*
321 *aeruginosa* infection caused by contaminated bottled water in intensive care units. *Clin*
322 *Microbiol Infect* 2008; **14**: 454-458.
- 323 24. Kikuchi T, Nagashima G, Taguchi K et al. Contaminated oral intubation equipment
324 associated with an outbreak of carbapenem-resistant *Pseudomonas* in an intensive care
325 unit. *J Hosp Infect* 2007; **65**: 54-57.
- 326 25. van Belkum A, Tassios PT, Dijkshoorn L et al. Guidelines for the validation and
327 application of typing methods for use in bacterial epidemiology. *Clinical Microbiology*
328 *and Infection* 2007; **13**: 1-46.

329

330

331 **Figure legend**

332

333 **Figure** : Evolution of the annual no. of cases per 1,000 admissions and per category, and of
334 the percentage of faucets contaminated with *P. aeruginosa* in the medical ICU (A), and in the
335 paediatric-surgical ICUs (B)(category A : identical to faucet ; category B : identical to at least
336 one other patient ; category C : unique genotype ; category D : not typed).

Table 1 . Epidemiological data of patients with *Pseudomonas aeruginosa* in the Intensive Care Units over a 10-year period

	1998	1999	2000	2001	2002	2003	2004	2005	2006	2007
No. of admissions in ICUs	2434	2538	2449	2586	2427	2400	2554	2530	2567	2675
No. of patients with <i>P. aeruginosa</i>	139	107	75	52	87	92	82	64	75	83
No. of patients with <i>P. aeruginosa</i> /1,000 admissions	57,1	42,2	30,6	20,1	35,8	38,3	32,1	24,9	29,2	31,0
No. of clinical specimens [#] /1000 patient-days of hospitalization	NA	NA	668	792	786	817	815	727	757	815
Percent of faucets [§] contaminated with <i>P. aeruginosa</i>										
in the medical ICUs (water network I)	25	53,1	38,1	13,3	22,7	8,3	8,7	4,2	ND	ND
in surgical and pediatric ICUs (water network II)	6,4	0	0	4,3	3,1	4,3	2,9	0	0	0
Categories of cases* (no. of cases/1,000 admissions)										
in the medical ICU:										
Category A	32,6	ND	8,5	ND	ND	1,0	1,9	ND	ND	0
Category B	20	ND	7,4	ND	ND	1,9	5,6	ND	ND	8,3
Category C	10,5	ND	8,5	ND	ND	18,4	16,7	ND	ND	16,5
Category D	4,2	ND	1,1	ND	ND	13,6	5,6	ND	ND	0,9
Total	67,3	55,4	25,5	16,2	26,1	34,9	29,8	21,1	22,3	25,7
in the surgical and pediatric ICUs:										
Category A	16,8	ND	4,0	ND	ND	0	1,4	ND	ND	0
Category B	13,5	ND	6,7	ND	ND	18,3	10,2	ND	ND	12,6
Category C	18,2	ND	18,0	ND	ND	16,8	16,9	ND	ND	20,2
Category D	4,0	ND	2,0	ND	ND	10,2	6,1	ND	ND	2,5
Total	52,5	36,5	30,7	23,1	46,7	45,3	34,6	27,5	35,5	35,3

NA, not available

ND, not done (see text)

[#], Clinical specimens received by the laboratory of bacteriology.^{*}, Category A : cases with a *P.aeruginosa* genotype identical to one found in faucets during the same year or previously; category B : identical to at least one other patient; category C : unique genotype ; category D : not typed.[§], the percentage of contaminated faucets was based on data from faucets that were regularly swabbed over the 10 years.

Table 2. Analysis of genetic and epidemiological clusters in cases of category A (patients with a *P. aeruginosa* genotype identical to that found in faucets) and category B (patients with a *P.aeruginosa* genotype identical to at least one other patient and not found in faucets).

	1998	2000	2003	2004	2007
No. of cases in category A	57	17	1	4	0
No. of genetic clusters	10	7	1	4	0
No. of patients in genetic clusters	30, 6*, 5, 4, 4, 3, 2, 1, 1, 1	8, 4, 1, 1, 1, 1, 1	1*	1, 1, 1, 1	0
No. of cases in category B	39	17	27	21	29
No. of genetic clusters	11	4	5	8	12
No. of patients in genetic clusters	11 (6, 2)	8 (3)	13 (9,2)	5 (3)	3 (3)
(no. of patients in epidemiologic clusters)	6 (0)	4 (0)	8 (3,3,2)	3 (0)	3 (0)
	3 (3)	3 (2)	2 (0)	3 (0)	2 (2)
	3 (2)	2 (0)	2 (0)	2 (2)	2 (2)
	3 (0)		2 (0)	2 (0)	2 (0)
	3 (0)			2 (0)	2 (0)
	2 (2)			2 (0)	2 (0)
	2 (2)			2 (0)	2 (0)
	2 (0)				2 (0)
	2 (0)				2 (0)
	2 (0)				2 (0)
					2(0)

* one patient harbored a genotype identical to that found in faucet, but was hospitalized in a different ICU, no probable transmission was thus considered.