

# CD4<sup>+</sup>CD25<sup>+</sup>Foxp3<sup>+</sup> regulatory T cells: from basic research to potential therapeutic use

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## Summary

Regulatory T cells control immune responses to self- and foreign-antigens and play a major role in maintaining the balance between immunity and tolerance. This article reviews recent key developments in the field of CD4<sup>+</sup>CD25<sup>+</sup>Foxp3<sup>+</sup> regulatory T (T<sub>REG</sub>) cells. It presents their characteristics and describes their range of activity and mechanisms of action. Some models of diseases

triggered by the imbalance between T<sub>REG</sub> cells and effector pathogenic T cells are described and their potential therapeutic applications in humans are outlined.

*Key words:* CD4<sup>+</sup>CD25<sup>+</sup> Foxp3<sup>+</sup> regulatory T cells; tolerance; review

## Introduction

The existence of cell-subsets specialised in suppressing immune responses was postulated in the early seventies by Gershon and colleagues [1]. Nevertheless, convincing evidence came only in the mid eighties from data obtained by Sakaguchi et al. [2] and by Don Mason and colleagues [3, 4] using experimental autoimmune disease mouse or rat models. Progress was hampered until the mid-nineties by the lack of reliable phenotypical markers. The breakthrough in the modern era of regulatory T cells is almost certainly due to the work of Sakaguchi and colleagues, who in 1995 described a subpopulation of CD4<sup>+</sup> T cells with high cell surface expression of the interleukin-2 receptor (IL-2R)  $\alpha$ -chain, also called CD25, which was essential for the prevention of autoimmunity [5].

This review focuses on naturally occurring CD4<sup>+</sup>CD25<sup>+</sup> Foxp3<sup>+</sup> T (T<sub>REG</sub>) cells. It stresses recent key progress in defining their characteristics and focuses on the transcription factor Foxp3, IL-2 and the components of the high-affinity IL-2 receptor, as these molecules are essential for the development, function and survival of T<sub>REG</sub> cells. It also describes the T<sub>REG</sub> cells' mechanisms of action and their potential therapeutic applications either in enhancing their regulatory activity in inflammatory diseases such as autoimmunity, allograft rejection, graft versus host disease (GVHD) and allergic diseases, or in blocking their suppressive activity in tumour immunity or vaccine development.

## A definition according to function

Immunoregulation is an active process in which one population of cells controls the activity of another cell population. So far, various populations of T cells with regulatory properties have been characterised *in vitro* and *in vivo* and have

### Abbreviations

APC	Antigen-presenting cells
BMT	Bone marrow transplantation
CTLA-4	Cytotoxic T lymphocyte-associated antigen-4
DC	Dendritic cells
EAE	Experimental autoimmune encephalomyelitis
GITR	Glucocorticoid induced tumour necrosis factor
GVHD	Graft versus host disease
IBD	Inflammatory bowel disease
IPEX	Immunodysregulation, Polyendocrinopathy, Enteropathy, X-linked recessive
IL-	Interleukin
mAb	Monoclonal antibody
MHC	Major histocompatibility complex
T1D	Type 1 diabetes
TCR	T cell receptor
TGF- $\beta$	Transforming growth factor- $\beta$
TLR	Toll like receptor
T <sub>REG</sub>	CD4 <sup>+</sup> CD25 <sup>+</sup> Foxp3 <sup>+</sup> T
XLAAD	X-linked autoimmunity-allergic dysregulation

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been implicated in the control of autoimmune diseases, allotransplantation tolerance and anti-tumour immune responses. Naturally occurring regulatory T cells have been identified in non-manipulated rodents and humans, and comprise cells of the adaptive immune system (CD4<sup>+</sup>CD25<sup>+</sup>Foxp3<sup>+</sup> T cells) and of the innate immune system (natural killer [NK] T cells) [6-8]. Apart from these spontaneously arising regulatory T cells, it has been reported that an uncommitted T cell can be skewed towards a regulatory function by *in vitro* or *in vivo* manipulation. Repetitive *in vitro* stimulation with immature dendritic cells (DC) or in the presence of suppressive cytokines such as IL-10 or TGF- $\beta$  can induce regulatory T cells. *In vivo*, these cells have been identified in the course of tolerance induction protocols. Regulatory T cells of the CD4<sup>+</sup> subset, such as Tr1 and Th3 cells [9-11], CD8<sup>+</sup> (CD8<sup>+</sup>CD28<sup>-</sup> T cells) [12], double negative CD3<sup>+</sup>CD4<sup>-</sup>CD8<sup>-</sup> T cells [13], and  $\gamma\delta$  T cell [14] subsets have been functionally characterised in *in vitro* and *in vivo* settings. We will not discuss these cells further and will focus on the naturally occurring CD4<sup>+</sup>CD25<sup>+</sup>Foxp3<sup>+</sup> T cells (T<sub>REG</sub>).

T<sub>REG</sub> cells are defined operationally by their functional ability to regulate/suppress immune responses. Per se, T<sub>REG</sub> cells are hyporesponsive to T cell receptor (TCR) stimulation *in vitro*, but exogenous IL-2, strong co-stimulation with increasing concentrations of anti-CD28 monoclonal antibody (mAb) or stimulation bypassing the TCR (such as mitogens) can overcome their anergic state [7]. On polyclonal or antigen-specific TCR stimulation, T<sub>REG</sub> cells potently suppress the proliferation and cytokine production of

effector CD4<sup>+</sup> and CD8<sup>+</sup> T cells by inhibiting IL-2 gene transcription [15-18]; they also inhibit the proliferation and antibody production of B cells [19]. The induction of the suppressive function of T<sub>REG</sub> is antigen-specific and T<sub>REG</sub> cells appear to have a higher avidity for specific antigens as compared to their CD4<sup>+</sup>CD25<sup>-</sup> counterparts, as their suppressive activity is elicited at a 10-100-fold lower concentration of a specific peptide [18]. Several studies have implied that the regulation mediated by T<sub>REG</sub> cells was dependent on a continuous supply of allo-antigens [20] or tissue-specific target auto-antigens [21, 22], as removal of the source of tissue antigens may lead to rapid contraction of the pool of tissue-specific T<sub>REG</sub> cells.

In contrast to their *in vitro* resistance to proliferation upon TCR stimulation, T<sub>REG</sub> cells show active proliferation *in vivo* following antigenic stimulation [23, 24]. Naturally arising T<sub>REG</sub> cells play a key role in the maintenance of peripheral dominant tolerance to self- and alloantigens *in vivo* (reviewed in [7, 25]). Depletion of CD4<sup>+</sup>CD25<sup>+</sup> T cells induces effective tumour immunity [26-29], enhances immune responses to invading microbes, triggers allergic responses to innocuous environmental substances and breaks foeto-maternal tolerance during pregnancy [30]. On the other hand, T<sub>REG</sub> cells have been found to suppress a number of T cell-mediated immune pathologies, including allergic responses and autoimmune diseases such as type 1 diabetes (T1D) [31], experimental autoimmune encephalomyelitis (EAE) [32-34], gastritis, colitis [35], glomerulonephritis, and polyarthritis [5, 36] as well as allograft rejection [37-40] and GVHD [41-43].

## Phenotypic characterisation

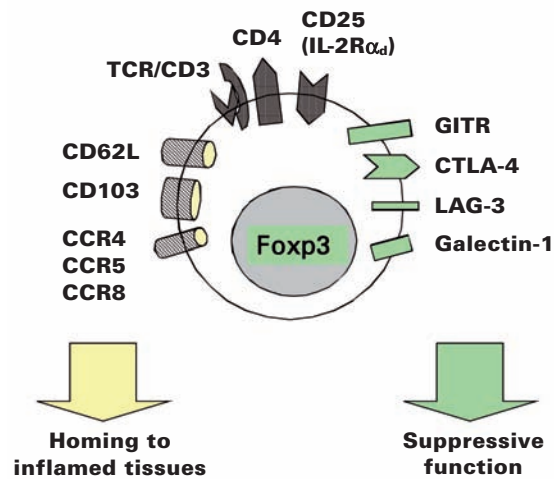
Naturally occurring CD4<sup>+</sup>CD25<sup>+</sup> T cells develop in the thymus [44] and constitute some 5-10% of peripheral CD4<sup>+</sup> T cells in non-manipulated normal mice and humans [45-47], but in humans only the CD4<sup>+</sup>CD25<sup>high</sup> cells which constitute 2-3 % of the CD4<sup>+</sup> T cells are really regulatory [48]. To date the best surface marker correlating with suppressive activity is CD25 (Figure 1). However, this marker is not specific as its up-regulation following cell activation does not confer regulatory properties, and it has been demonstrated that cells with regulatory properties can also be found in the CD4<sup>+</sup>CD25<sup>-</sup> T cell pool [49, 50]. The naturally occurring CD4<sup>+</sup>CD25<sup>+</sup> T cell population also expresses the cytotoxic T lymphocyte-associated antigen-4 (CTLA-4) [35, 51] and the glucocorticoid-induced tumour necrosis factor (GITR) [52, 53], but, like CD25, these molecules are also up-regulated following activation of naïve CD4<sup>+</sup>CD25<sup>-</sup> T cells. When compared to the CD4<sup>+</sup>CD25<sup>-</sup> population, the T<sub>REG</sub> population expresses low levels of CD45RB and contains in-

creased frequencies of CD62L<sup>high</sup> T, indicating an activated/primed or memory state.

The  $\alpha_E\beta_7$ -integrin (CD103), an integrin which recognises epithelial cadherin responsible for tissue-specific retention of lymphocytes, was shown to identify a subpopulation of T<sub>REG</sub> cells [54] and to be important in the homing and retention of T<sub>REG</sub> cells in inflamed tissues [55]. CD103 is expressed in more than 90% of intestinal and extraintestinal (bronchi, inflammatory skin/breast/salivary glands, tumour epithelium) intraepithelial T lymphocytes, while it is only expressed by 0.5-5% of lymphocytes in peripheral blood and lymphoid organs. T<sub>REG</sub> cells also express specific chemokine receptors such as CCR4, CCR5 and CCR8. Mature DC producing CCL17 and CCL22 were found to preferentially attract T<sub>REG</sub> cells to sites of antigen presentation in secondary lymphoid organs and peripheral inflamed areas [56]. T<sub>REG</sub> cells have also been shown to be attracted by CCL4, a chemokine expressed by activated B cells, DC and macrophages [19], and their

**Figure 1**

Phenotypic markers characteristic of naturally occurring T<sub>REG</sub> cells.



homing to allografts was dependent on the CCR4 chemokine receptor pathway [57].

More recently, LAG-3 (CD223) a cell surface CD4-related molecule that binds the major histocompatibility complex (MHC) class II was shown to be selectively expressed on T<sub>REG</sub> cells up on activation and to be involved in their suppressive function [58]. Lechler's group has also reported that a member of the  $\beta$ -galactoside-binding proteins, galectin-1, was constitutively overexpressed in T<sub>REG</sub> cells and was an important effector of regulation mediated by these cells [59]. Also, two groups have recently described a critical role for the Wiskott-Aldrich Syndrome Protein (WASP) in the activation and suppressor function of T<sub>REG</sub> cells which implicates T<sub>REG</sub> cell dysfunction in the autoimmunity associated with the Wiskott-Aldrich syndrome [60, 61].

### Key roles of Foxp3 and of the IL-2/IL-2 receptor components

In addition to CD25 expressed on the cell surface, the transcription factor Foxp3 was shown to be a highly specific intracellular marker for T<sub>REG</sub> cells [62-64]. Foxp3, the cytokine IL-2, and CD25 as a component of the IL-2 receptor, are

essential for the development, function and survival of T<sub>REG</sub> cells because mutation or polymorphisms in the genes encoding these molecules are causative of, or predispose to, autoimmune diseases in both rodents and humans. The *Foxp3* gene was identified as the defective gene in the scurfy mouse strain, whose phenotype is an X-linked recessive mutant with lethality in males within a month after birth due to excessive activation of CD4<sup>+</sup> T cells and overproduction of pro-inflammatory cytokines [65]. Mutations of the human ortholog *Foxp3* gene were subsequently found to be the cause of the immune dysregulation, polyendocrinopathy, and enteropathy, X-linked recessive (IPEX) and the X-linked autoimmunity-allergic dysregulation (XLAAD) syndromes. Patients carrying these mutations develop organ-specific autoimmune diseases such as T1D, thyroiditis, inflammatory bowel disease (IBD), allergic dermatitis, food allergy, haematological disorders, and serious infections [66-68]. The specific role of Foxp3 in the function of T<sub>REG</sub> cells was highlighted by the fact that retroviral transduction of *Foxp3* into CD4<sup>+</sup>CD25<sup>-</sup> T cells can convert them to functional T<sub>REG</sub> cells able to suppress proliferation of other T cells *in vitro* and inhibit the development of autoimmune diseases, such as colitis or T1D, mediated by pathogenic effector T cells in *in vivo* experimental models [7, 63].

IL-2 has been shown to be essential for the generation of T<sub>REG</sub> cells in the thymus and their survival, expansion and suppressive function in the periphery [69]. IL-2-, and IL-2R-deficient mice develop T-cell lympho-proliferation and lethal autoimmunity, very probably due to lack of activation-induced cell death (AICD) and lack of T<sub>REG</sub> cells [70, 71]. Furthermore, *in vivo* IL-2 neutralisation by use of an IL-2 blocking antibody also induces autoimmune diseases in mice [72]. The detailed molecular mechanisms of the effects of IL-2 in the homeostasis and suppressive function of T<sub>REG</sub> cells have still to be clarified.

## Mechanisms of regulation

### Contact-dependent versus cytokine-mediated effect

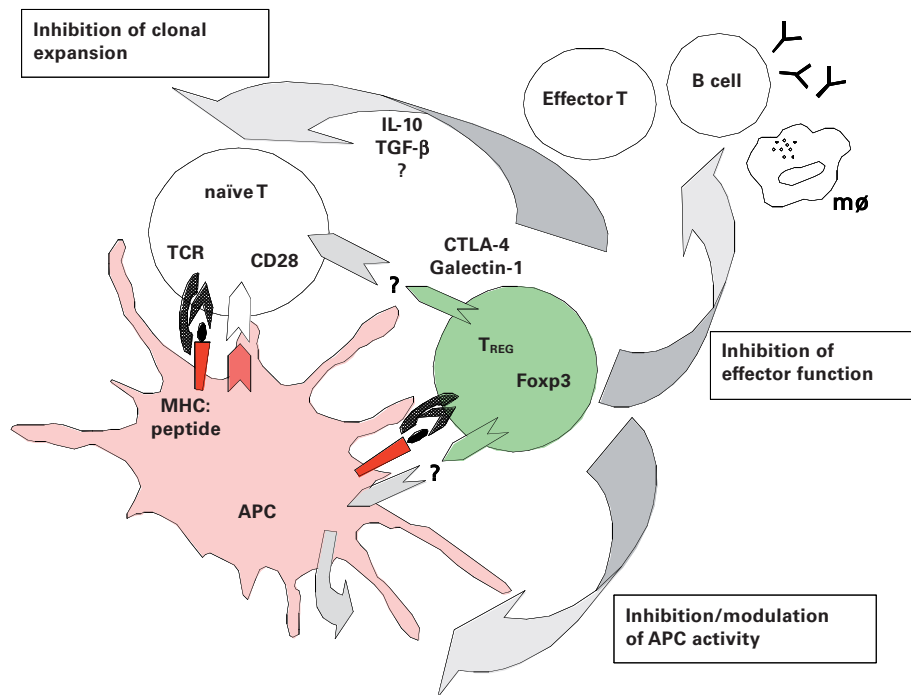
T<sub>REG</sub> cells' precise mechanisms of action are still unclear, with divergent conclusions regarding the importance of cell-cell contact versus cytokines in their suppressive function (Figure 2). The discrepant results might be explained by differences in the experimental systems (*in vitro* versus *in vivo*), the various disease models studied, the pathogenic effector mechanisms and target organs involved, and the contribution of the genetic background of the mouse strains used.

### Role of cell surface molecules

*In vitro*, T<sub>REG</sub> cells have been shown to inhibit the activation of effector CD4<sup>+</sup>CD25<sup>-</sup> T cells by a mechanism which depends on cell-cell contact rather than on soluble mediators [16]. CTLA-4 [35, 51], membrane-bound transforming growth factor- $\beta$  (TGF- $\beta$ ) [73], GITR [52, 53], and galectin-1 [59] appear to be functionally important molecules since in various models monoclonal antibodies (mAbs) against these molecules have been shown to overcome the suppression mediated by T<sub>REG</sub> cells *in vitro* and/or *in vivo*. Moreover, after activation, human T<sub>REG</sub> cells were shown to be able to directly kill activated CD4<sup>+</sup>

**Figure 2**

Mechanisms of regulation of immune responses mediated by T<sub>REG</sub> cells. T<sub>REG</sub> cells can suppress the activation, clonal expansion and/or the effector function of auto- or alloreactive pathogenic T cells, B cells and macrophages (mφ), as well as modulate the immunogenicity of APC. The precise mechanisms of action of T<sub>REG</sub> cells are still unclear, with divergent conclusions regarding the importance of cell-cell contact (via surface-bound molecules) versus secreted cytokines in their suppressive function.



and CD8<sup>+</sup> T cells in a perforin or granzyme-dependent way [74].

Direct T-T interactions via surface molecules may mediate suppression *in vitro*, since activated T<sub>REG</sub> cells have been shown to suppress the proliferation of effector T cells in response to TCR stimulation in the complete absence of antigen-presenting cells (APC) [7]. T<sub>REG</sub> cells may also down-modulate the function of APC and render them unable to activate effector T cells. It was shown that co-culture with activated T<sub>REG</sub> cells leads to reduced amounts of costimulatory molecules on DC and B cells [75], inhibiting their immunogenic properties. “Tolerogenic” DCs may in turn induce other regulatory cells, thus contributing to the maintenance of tolerance. *In vivo*, however, it is likely that more complex interactions between T<sub>REG</sub> cells, effector T cells and APC/DC determine the resulting immune response (“ménage à trois”). Another way in which T<sub>REG</sub> cells could mediate their immune suppressive properties is to induce differentiation of naive T cells into cells with a regulatory function rather than into pathogenic effector T cells. This phenomenon has been termed “infectious tolerance”, and obvious candidate molecules mediating this effect may be regulatory cytokines such as IL-10 or TGF-β [76–78].

#### CTLA-4

A large proportion of T<sub>REG</sub> cells express CTLA-4 in both mouse and human. CTLA-4 is known to be a negative regulator of T-cell activation but also to play a key role in immunological self-tolerance. Non-activating anti-CTLA-4 mAbs have been shown to block the suppressor activity of T<sub>REG</sub> cells *in vitro* and *in vivo* [35, 51], and mice deficient in CTLA-4 develop a fatal lymphopro-

liferative disease and multiorgan inflammation [79]. The role of CTLA-4 expression in the homeostasis and function of T<sub>REG</sub> cells remains however controversial [80].

#### Role of soluble cytokines (IL-10, TGF-β)

##### IL-10

*In vivo* models of colitis [81–83] and infection [84] show that IL-10 is often crucial for the maintenance of immune homeostasis and regulation mediated by T<sub>REG</sub> cells. In some transplantation models anti-IL-10 or anti-IL-10R mAbs abrogated the induction of peripheral tolerance by otherwise previously proven robust tolerogenic protocols [39]. IL-10 is produced by a large number of immune cells such as T cells, monocytes, macrophages and epithelial cells, and has pleiotropic effects on B, T and NK cells, DC and mast cells. Signalling through the IL-10/IL-10R results in potent inhibition of cell proliferation, of pro-inflammatory cytokine production and of the maturation and antigen presentation of DC [78]. IL-10 also plays a pivotal role in T<sub>REG</sub> cell function, CD4<sup>+</sup>CD25<sup>+</sup> T cells from IL-10<sup>-/-</sup> mice having been shown to be significantly less potent than their wild type counterparts. Using a T cell-mediated model of murine colitis, Powrie’s group has shown that intestinal inflammation mediated by the transfer of naive CD4<sup>+</sup>CD25<sup>-</sup> T cells can be cured by the transfer of T<sub>REG</sub> cells. In this model, during the cure of colitis, IL-10-secreting Foxp3<sup>+</sup> T cells selectively enriched within the colonic lamina propria. The administration of an anti-IL-10R mAb to mice treated with T<sub>REG</sub> cells completely abolished the cure. These findings were further confirmed by analysis of human colonic samples, where an accumulation of

Foxp3<sup>+</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells was found in the lamina propria of patients with IBD, diverticulitis, pseudomembranous and CMV colitis [85, 86].

### TGF- $\beta$

TGF- $\beta$  is the prototype of a family of polypeptides involved in growth control, extracellular matrix production and development. Three isoforms of TGF- $\beta$  exist in mammals (TGF- $\beta$ 1, - $\beta$ 2, and - $\beta$ 3) and TGF- $\beta$ 1 has been identified as an immunoregulatory molecule with both immunogenic and immunosuppressive properties depending on the cellular environment [77]. Most models of immune regulation showing a role of IL-10 also implicate TGF- $\beta$ , the production and

action of these two cytokines being interrelated and likely to involve positive feedback loops in which IL-10 enhances expression of TGF- $\beta$  and vice versa. IL-10 may act locally at the site of inflammation, while TGF- $\beta$  seems to have a more systemic effect on the immune response. The suppressive activity of TGF- $\beta$  is best highlighted by the fact that TGF- $\beta$ -deficient mice develop a lethal lymphoproliferative disease [87]. The results concerning the requirement for TGF- $\beta$  expression by T<sub>REG</sub> *in vitro* and *in vivo* are controversial [73, 88, 89], and it appears that regulation is dictated primarily by the responsiveness of the effector T cell to TGF- $\beta$  [90, 91].

## Models of diseases triggered by imbalance between T<sub>REG</sub> and effector T cells

### Autoimmune and allergic diseases

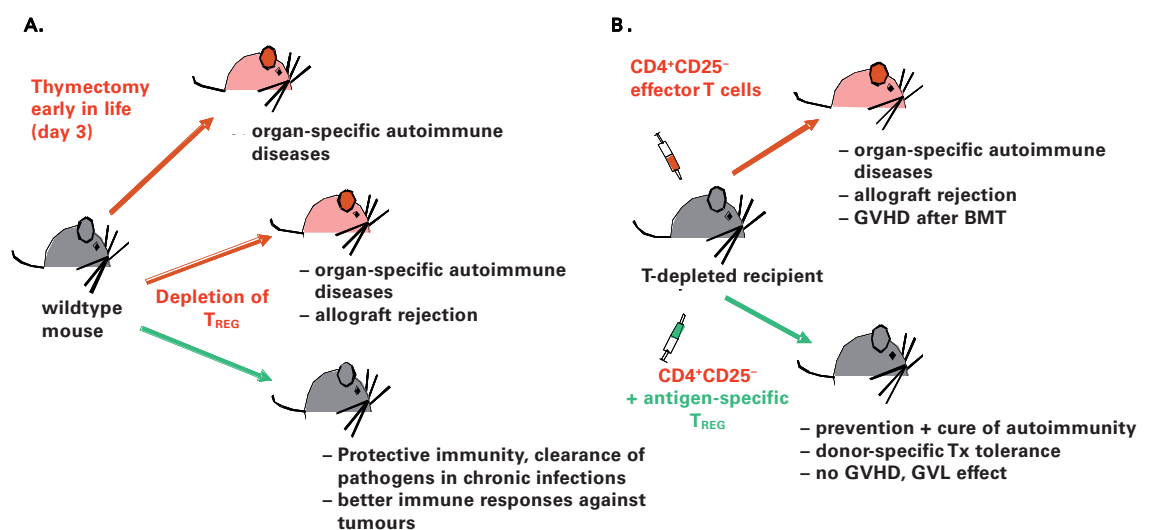
There is now a mass of evidence that T<sub>REG</sub> actively suppress the activation and expansion of self-reactive T cells *in vivo* and thereby the development of autoimmune diseases. Neonatal thymectomy or depletion of T<sub>REG</sub> cells from normal animals result in spontaneous development of various organ-specific autoimmune diseases (such as gastritis, IBD, T1D, EAE, myocarditis, depending on the genetic background of the strains of mice used) and increase alloresponses to skin allografts [5, 7] (Figure 3). In these cases, autoimmunity is prevented by reconstitution of the mice with wild type T<sub>REG</sub> cells. In the regulation of allergy T<sub>REG</sub> cells have been shown to be effective in suppressing the production of antigen-specific IgE in TCR transgenic murine models [92].

In humans, a study by Danke *et al.* illustrates the fact that T cells reactive to target self-antigens, such as glutamic acid decarboxylase, which is involved in T1D, could be easily expanded *in vitro* from peripheral blood of normal individuals only after depleting the CD25<sup>+</sup> subset [93]. These results indicate that self-reactive T cells are present in most individuals but their pathogenic potential is kept in check by coexisting T<sub>REG</sub> cells. IPEX as a prototype of disease induced by an intrinsic defect of T<sub>REG</sub> cells is thus far the clearest example showing that an abnormality in the pool of naturally arising T<sub>REG</sub> cells is a primary cause of autoimmune diseases in humans, and that T<sub>REG</sub> cells are crucial in the maintenance of dominant self-tolerance. Alterations in the numbers or functional activity of T<sub>REG</sub> cells, in peripheral blood or

**Figure 3**

### T<sub>REG</sub> cells regulate immune pathology *in vivo*.

A. Neonatal thymectomy or depletion of T<sub>REG</sub> cells from normal animals, results in spontaneous development of autoimmune diseases and increased alloresponses to solid organ allografts, while it enhances immune responses against tumours. B. In experimental models the co-transfer of T<sub>REG</sub> cells in lymphopenic hosts protects against immune pathologies induced by effector CD4<sup>+</sup>CD25<sup>+</sup> T cells. GVL: graft versus leukaemia. Tx: transplantation.



target organs, have also been associated with several human chronic inflammatory and autoimmune diseases such as multiple sclerosis [94], rheumatoid arthritis [95–97], T1D [98], IBD [99], systemic lupus erythematosus [100], polyglandular syndrome type II [101], myasthenia gravis [102], allergic asthma [103], cow's milk allergy [104], nickel allergy [105] and atopic dermatitis [106].

### Infectious diseases

T<sub>REG</sub> cells probably participate in the immune response to all infectious agents, since apart from regulating adaptive immune responses they have been shown to be also capable of directly suppressing innate immune responses in a model of murine pathogen-mediated colitis [84]. How T<sub>REG</sub> cell activity is regulated to allow effective immune responses towards pathogens without pathological anti-self reactivity has baffled immunologists [107].

Activation of DCs through Toll like Receptor (TLR) signalling can overcome dominant T<sub>REG</sub> cell suppression *in vitro* and enhance effector T cell responses [108]. T<sub>REG</sub> cells appear however to restrain too vigorous immune reactivity, which in many chronic infections will benefit the host by limiting tissue damage. Current evidence suggests that in the course of an infection TLR signalling would directly regulate the suppressive function of T<sub>REG</sub> cells by augmenting T<sub>REG</sub> cell proliferation with a temporal loss of their suppressive activity. The T<sub>REG</sub> cells would recover their suppressive activity when the infection has subsided, in time to limit potential autoimmunity that could result from overactivated effector mechanisms. It therefore appears that the balance between T<sub>REG</sub> cells and effector T cells depends on the activation status of the innate immune system and particularly the DC.

In many chronic infections T<sub>REG</sub> cells appear to restrain immune reactivity in order to limit host tissue damage, but this may handicap the efficacy of protective immunity and clearance of pathogens [107]. Studies on persistent chronic infections, such as herpes simplex virus (HSV), hepatitis C virus (HCV) and HIV, have shown that the presence of T<sub>REG</sub> cells at the time of infection may affect the magnitude of protective immunity and the outcome of infection [109–113]. When T<sub>REG</sub> cells were depleted, animals developed memory responses and could subsequently mount better recall responses. T<sub>REG</sub> cells were increased in number and activity in chronic HCV patients compared to controls who resolved their infections. Such T<sub>REG</sub> cells isolated from peripheral blood, and in one instance from the liver itself, modulated peptide-specific proliferative responses and maturation of HCV-specific CD8<sup>+</sup> T cells. Similarly, reduction of tissue-damaging immunopathology by T<sub>REG</sub> cell function has been observed in some parasitic infections such as *Pneumocystis carinii*, *Leishmania major* and *Schistosoma masoni* [114, 115].

### Transplantation

In the transplant setting, circulating alloreactive T cells are crucial in the initiation and the coordination of the rejection response and, to promote tolerance, it is important to deplete or minimise the alloreactive effector T cell pool while enhancing regulatory mechanisms. Various strategies, targeting T cell activation, expansion and/or effector function, have been described as a means of achieving robust peripheral transplantation tolerance in experimental protocols [116]. More recent studies have shown that in many of these protocols immunoregulatory mechanisms dependent on T<sub>REG</sub> cells were critical in the induction and maintenance of peripheral tolerance. Thus, after a short course of immunomodulatory drugs, such as non-depleting anti-T cell mAbs or costimulatory blockade, donor alloantigen-specific T<sub>REG</sub> cells are generated which are capable, on adoptive transfer, of suppressing rejection of donor allografts mediated by naïve recipient CD4<sup>+</sup> or CD8<sup>+</sup> T cells [37–40]. We have also reported that *in vivo* allo-responses can be harnessed by donor alloantigen-specific T<sub>REG</sub> cells selected and expanded *in vitro* [117].

In murine models of allogeneic bone marrow transplantation (BMT), it has similarly been shown that freshly isolated or *ex-vivo* expanded donor-derived T<sub>REG</sub> cells can delay or even prevent GVHD, and that the selective depletion of T<sub>REG</sub> cells in the transplant results in increased severity of acute GVHD [41–43]. Patients who developed acute or chronic GVHD after allogeneic BMT had a decreased number of peripheral and tissue infiltrating T<sub>REG</sub> cells [118, 119]. Intestinal mucosal specimens without histological signs of GVHD showed increased ratios of FOXP3<sup>+</sup>/CD8<sup>+</sup> T cells compared to samples with histological signs of acute and chronic GVHD, providing further evidence that GVHD lesions are associated with lack of regulation by T<sub>REG</sub> cells.

### Tumour immunity

Depletion of T<sub>REG</sub> cells using CD25-specific mAbs has been shown to promote rejection of several transplantable murine tumour cell lines, including melanoma, fibrosarcoma, leukaemia and colorectal carcinoma. These studies imply that T<sub>REG</sub> cells normally inhibit the generation of effective T cell-dependent anti-tumour immune responses [27–29]. These findings have been confirmed in the clinical setting, where the prevalence of T<sub>REG</sub> cells was found to be increased in the peripheral blood and tumour microenvironment of cancer patients [26, 120–123].

## Therapeutic potential of T<sub>REG</sub> cells

Approaches to prevention or treatment of T cell-mediated diseases, such as autoimmunity and transplant rejection, have focused historically on potent immunosuppressive drugs non-specifically targeting T cell responses. However, the improved survival rates of allografts and the better outcome of autoimmune diseases have come at a cost, with increased frequencies of drug-related adverse effects. A better understanding of the role of naturally occurring T<sub>REG</sub> cells in the maintenance of immune homeostasis has prompted researchers to investigate their therapeutic potential. T<sub>REG</sub> cells could be used as an immunotherapeutic tool, either by enhancing their activity in inflammatory diseases such as autoimmunity, allograft rejection or GVHD, or by blocking their suppressive activity in tumour immunity or vaccine development.

If T<sub>REG</sub> cells are to be used in immunotherapy aiming at the prevention or treatment of autoimmune diseases or allograft rejection, antigen-specific cells are needed which can specifically control immune responses to the relevant auto- or allo-antigens, while allowing protective immune responses to pathogens. Furthermore, because of the low precursor frequency of alloantigen cross-reactive T<sub>REG</sub> cells expected in a normal individual without prior antigen exposure, this specific population needs to be expanded *in vivo* or gener-

ated in large numbers *in vitro* for adoptive transfer. New experimental evidence has shown, in various experimental settings, that antigen-specific T<sub>REG</sub> cells can be generated and expanded in sufficient quantities *in vitro* without loss of their characteristic phenotype and regulatory properties [117, 124–126]. Importantly, we have shown that *in vitro* manipulations did not modify their *in vivo* homeostasis, migration patterns or suppressive functions, as demonstrated by adoptive transfer experiments [117]. Alternatively, immunomodulatory drugs, such as rapamycin, could be used to selectively expand T<sub>REG</sub> cells *in vivo* while controlling the effector T cell pool [127].

The other side of the coin is that there is now substantial evidence to show that the presence of T<sub>REG</sub> cells is deleterious in cancer patients and contributes to the unsuccessful immune responses in some chronic persistent infections. Therefore, depletion of T<sub>REG</sub> cells in combination with other anti-tumour therapies could optimise eradication of malignancies. Depletion of T<sub>REG</sub> cells can also be used to boost immune responses in vaccine development or to enhance protective immune responses against invading microbes. This may, however, prevent the induction of long-term infectious immunity or favour the development of autoimmune diseases.

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## Conclusion

These novel therapeutic approaches are not without risks. The key to success will be to establish the correct balance between T<sub>REG</sub> cells and effector cells. These exciting new therapeutic concepts will undoubtedly act as a spur to the translational research community over the next few years. Indeed, steady progress in our understanding of T<sub>REG</sub> cell function *in vivo* and their dynamics with pathogenic T cells provides the rationale for a novel form of individualised “tailored” medicine using T<sub>REG</sub> cell-based therapies in the prevention and treatment of autoimmune diseases and in transplantation.

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## References

- Gershon RK. A disquisition on suppressor T cells. *Transplant Rev.* 1975;26:170–85.
- Sakaguchi S, Fukuma K, Kuribayashi K, Masuda T. Organ-specific autoimmune diseases induced in mice by elimination of T cell subset. I. Evidence for the active participation of T cells in natural self-tolerance; deficit of a T cell subset as a possible cause of autoimmune disease. *J Exp Med.* 1985;161:72–87.
- Fowell D, Mason D. Evidence that the T cell repertoire of normal rats contains cells with the potential to cause diabetes. Characterization of the CD4+ T cell subset that inhibits this autoimmune potential. *J Exp Med.* 1993;177:627–36.
- Powrie F, Mason D. OX-22high CD4+ T cells induce wasting disease with multiple organ pathology: prevention by the OX-22low subset. *J Exp Med.* 1990;172:1701–8.
- Sakaguchi S, Sakaguchi N, Asano M, Itoh M, Toda M. Immunologic self-tolerance maintained by activated T cells expressing IL-2 receptor alpha-chains (CD25). Breakdown of a single mechanism of self-tolerance causes various autoimmune diseases. *J Immunol.* 1995;155:1151–64.
- Bendelac A, Rivera MN, Park SH, Roark JH. Mouse CD1-specific NK1 T cells: development, specificity, and function. *Annu Rev Immunol.* 1997;15:535–62.

- 7 Sakaguchi S. Naturally arising CD4<sup>+</sup> regulatory T cells for immunologic self-tolerance and negative control of immune responses. *Annu Rev Immunol.* 2004;22:531–62.
- 8 Shevach EM. CD4<sup>+</sup> CD25<sup>+</sup> suppressor T cells: more questions than answers. *Nat Rev Immunol.* 2002;2:389–400.
- 9 Groux H, O'Garra A, Bigler M, Rouleau M, Antonenko S, De Vries JE, et al. A CD4<sup>+</sup> T-cell subset inhibits antigen-specific T-cell responses and prevents colitis. *Nature.* 1997;389:737–42.
- 10 Roncarolo MG, Gregori S, Levings M. Type 1 T regulatory cells and their relationship with CD4<sup>+</sup>CD25<sup>+</sup> T regulatory cells. *Novartis Found Symp* 2003;252:115–27; discussion 127–131, 203–110.
- 11 Weiner HL. Induction and mechanism of action of transforming growth factor-beta-secreting Th3 regulatory cells. *Immunol Rev.* 2001;182:207–14.
- 12 Liu Z, Tugulea S, Cortesini R, Suci-Foca N. Specific suppression of T helper alloreactivity by allo-MHC class I-restricted CD8<sup>+</sup>CD28<sup>-</sup> T cells. *Int Immunol.* 1998;10:775–83.
- 13 Zhang ZX, Yang L, Young KJ, DuTemple B, Zhang L. Identification of a previously unknown antigen-specific regulatory T cell and its mechanism of suppression. *Nat Med.* 2000;6:782–9.
- 14 Hayday A, Tigelaar R. Immunoregulation in the tissues by gammadelta T cells. *Nat Rev Immunol.* 2003;3:233–42.
- 15 Itoh M, Takahashi T, Sakaguchi N, Kuniyasu Y, Shimizu J, Otsuka F, et al. Thymus and autoimmunity: production of CD25<sup>+</sup>CD4<sup>+</sup> naturally anergic and suppressive T cells as a key function of the thymus in maintaining immunologic self-tolerance. *J Immunol.* 1999;162:5317–26.
- 16 Thornton AM, Shevach EM. CD4<sup>+</sup>CD25<sup>+</sup> immunoregulatory T cells suppress polyclonal T cell activation in vitro by inhibiting interleukin 2 production. *J Exp Med.* 1998;188:287–96.
- 17 Piccirillo CA, Shevach EM. Cutting edge: control of CD8<sup>+</sup> T cell activation by CD4<sup>+</sup>CD25<sup>+</sup> immunoregulatory cells. *J Immunol.* 2001;167:1137–40.
- 18 Thornton AM, Shevach EM. Suppressor effector function of CD4<sup>+</sup>CD25<sup>+</sup> immunoregulatory T cells is antigen nonspecific. *J Immunol.* 2000;164:183–90.
- 19 Bystry RS, Aluvihare V, Welch KA, Kallikourdis M, Betz AG. B cells and professional APCs recruit regulatory T cells via CCL4. *Nat Immunol.* 2001;2:1126–32.
- 20 Scully R, Qin S, Cobbold S, Waldmann H. Mechanisms in CD4 antibody-mediated transplantation tolerance: kinetics of induction, antigen dependency and role of regulatory T cells. *Eur J Immunol.* 1994;24:2383–92.
- 21 Green EA, Choi Y, Flavell RA. Pancreatic lymph node-derived CD4<sup>+</sup>CD25<sup>+</sup> Treg cells: highly potent regulators of diabetes that require TRANCE-RANK signals. *Immunity.* 2002;16:183–91.
- 22 Seddon B, Mason D. Peripheral autoantigen induces regulatory T cells that prevent autoimmunity. *J Exp Med.* 1999;189:877–82.
- 23 Fisson S, Darrasse-Jeze G, Litvinova E, Septier F, Klatzmann D, Liblau R, et al. Continuous activation of autoreactive CD4<sup>+</sup>CD25<sup>+</sup> regulatory T cells in the steady state. *J Exp Med.* 2003;198:737–46.
- 24 Walker LS, Chodos A, Eggena M, Dooms H, Abbas AK. Antigen-dependent proliferation of CD4<sup>+</sup> CD25<sup>+</sup> regulatory T cells in vivo. *J Exp Med.* 2003;198:249–58.
- 25 Maloy KJ, Powrie F. Regulatory T cells in the control of immune pathology. *Nat Immunol.* 2001;2:816–22.
- 26 Curiel TJ, Coukos G, Zou L, Alvarez X, Cheng P, Mottram P, et al. Specific recruitment of regulatory T cells in ovarian carcinoma fosters immune privilege and predicts reduced survival. *Nat Med.* 2004;10:942–9.
- 27 Gallimore A, Sakaguchi S. Regulation of tumour immunity by CD25<sup>+</sup> T cells. *Immunology.* 2002;107:5–9.
- 28 Onizuka S, Tawara I, Shimizu J, Sakaguchi S, Fujita T, Nakayama E. Tumor rejection by in vivo administration of anti-CD25 (interleukin-2 receptor alpha) monoclonal antibody. *Cancer Res.* 1999;59:3128–33.
- 29 Shimizu J, Yamazaki S, Sakaguchi S. Induction of tumor immunity by removing CD25<sup>+</sup>CD4<sup>+</sup> T cells: a common basis between tumor immunity and autoimmunity. *J Immunol.* 1999;163:5211–8.
- 30 Zenclussen AC. CD4<sup>+</sup>CD25<sup>+</sup> T regulatory cells in murine pregnancy. *J Reprod Immunol.* 2005;65:101–10.
- 31 Salomon B, Lenschow DJ, Rhee L, Ashourian N, Singh B, Sharpe A, et al. B7/CD28 costimulation is essential for the homeostasis of the CD4<sup>+</sup>CD25<sup>+</sup> immunoregulatory T cells that control autoimmune diabetes. *Immunity.* 2000;12:431–40.
- 32 Furtado GC, Olivares-Villagomez D, Curotto de Lafaille MA, Wensky AK, Latkowski JA, Lafaille JJ. Regulatory T cells in spontaneous autoimmune encephalomyelitis. *Immunol Rev.* 2001;182:122–34.
- 33 Hori S, Haury M, Lafaille JJ, Demengeot J, Coutinho A. Peripheral expansion of thymus-derived regulatory cells in anti-myelin basic protein T cell receptor transgenic mice. *Eur J Immunol.* 2002;32:3729–35.
- 34 Kohm AP, Carpentier PA, Anger HA, Miller SD. Cutting edge: CD4<sup>+</sup>CD25<sup>+</sup> regulatory T cells suppress antigen-specific autoreactive immune responses and central nervous system inflammation during active experimental autoimmune encephalomyelitis. *J Immunol.* 2002;169:4712–6.
- 35 Read S, Malmstrom V, Powrie F. Cytotoxic T lymphocyte-associated antigen 4 plays an essential role in the function of CD25<sup>+</sup>CD4<sup>+</sup> regulatory cells that control intestinal inflammation. *J Exp Med.* 2000;192:295–302.
- 36 Suri-Payer E, Amar AZ, Thornton AM, Shevach EM. CD4<sup>+</sup>CD25<sup>+</sup> T cells inhibit both the induction and effector function of autoreactive T cells and represent a unique lineage of immunoregulatory cells. *J Immunol.* 1998;160:1212–8.
- 37 Cobbold SP, Graca L, Lin CY, Adams E, Waldmann H. Regulatory T cells in the induction and maintenance of peripheral transplantation tolerance. *Transpl Int.* 2003;16:66–75.
- 38 Graca L, Cobbold SP, Waldmann H. Identification of regulatory T cells in tolerated allografts. *J Exp Med.* 2002;195:1641–6.
- 39 Kingsley CI, Karim M, Bushell AR, Wood KJ. CD25<sup>+</sup>CD4<sup>+</sup> regulatory T cells prevent graft rejection: CTLA-4- and IL-10-dependent immunoregulation of alloresponses. *J Immunol.* 2002;168:1080–6.
- 40 Wood KJ, Sakaguchi S. Regulatory T cells in transplantation tolerance. *Nat Rev Immunol.* 2003;3:199–210.
- 41 Cohen JL, Trenado A, Vasey D, Klatzmann D, Salomon BL. CD4<sup>+</sup>CD25<sup>+</sup> immunoregulatory T Cells: new therapeutics for graft-versus-host disease. *J Exp Med.* 2002;196:401–6.
- 42 Etinger M, Hoffmann P, Ermann J, Drago K, Fathman CG, Strober S, et al. CD4<sup>+</sup>CD25<sup>+</sup> regulatory T cells preserve graft-versus-tumor activity while inhibiting graft-versus-host disease after bone marrow transplantation. *Nat Med.* 2003;9:1144–50.
- 43 Taylor PA, Lees CJ, Blazar BR. The infusion of ex vivo activated and expanded CD4<sup>+</sup>CD25<sup>+</sup> immune regulatory cells inhibits graft-versus-host disease lethality. *Blood.* 2002;99:3493–9.
- 44 Fontenot JD, Dooley JL, Farr AG, Rudensky AY. Developmental regulation of Foxp3 expression during ontogeny. *J Exp Med.* 2005;202(7):901–6.
- 45 Jonuleit H, Schmitt E, Stassen M, Tuettenberg A, Knop J, Enk AH. Identification and functional characterization of human CD4<sup>+</sup>CD25<sup>+</sup> T cells with regulatory properties isolated from peripheral blood. *J Exp Med.* 2001;193:1285–94.
- 46 Ng WF, Duggan PJ, Ponchel F, Matarese G, Lombardi G, Edwards AD, et al. Human CD4<sup>+</sup>CD25<sup>+</sup> cells: a naturally occurring population of regulatory T cells. *Blood.* 2001;98:2736–44.
- 47 Stephens LA, Mottet C, Mason D, Powrie F. Human CD4<sup>+</sup>CD25<sup>+</sup> thymocytes and peripheral T cells have immune suppressive activity in vitro. *Eur J Immunol.* 2001;31:1247–54.
- 48 Baecher-Allan C, Brown JA, Freeman GJ, Hafler DA. CD4<sup>+</sup>CD25<sup>high</sup> regulatory cells in human peripheral blood. *J Immunol.* 2001;167:1245–53.
- 49 Graca L, Thompson S, Lin CY, Adams E, Cobbold SP, Waldmann H. Both CD4<sup>+</sup>CD25<sup>+</sup> and CD4<sup>+</sup>CD25<sup>-</sup> regulatory cells mediate dominant transplantation tolerance. *J Immunol.* 2002;168:5558–65.
- 50 Stephens LA, Mason D. CD25 is a marker for CD4<sup>+</sup> thymocytes that prevent autoimmune diabetes in rats, but peripheral T cells with this function are found in both CD25<sup>+</sup> and CD25<sup>-</sup> subpopulations [In Process Citation]. *J Immunol.* 2000;165:3105–10.
- 51 Takahashi T, Tagami T, Yamazaki S, Uede T, Shimizu J, Sakaguchi N, et al. Immunologic self-tolerance maintained by CD25<sup>+</sup>CD4<sup>+</sup> regulatory T cells constitutively expressing cytotoxic T lymphocyte-associated antigen 4. *J Exp Med.* 2000;192:303–10.
- 52 McHugh RS, Whitters MJ, Piccirillo CA, Young DA, Shevach EM, Collins M, et al. CD4<sup>+</sup>CD25<sup>+</sup> immunoregulatory T cells: gene expression analysis reveals a functional role for the glucocorticoid-induced TNF receptor. *Immunity.* 2002;16:311–23.



- 53 Shimizu J, Yamazaki S, Takahashi T, Ishida Y, Sakaguchi S. Stimulation of CD25(+)CD4(+) regulatory T cells through GITR breaks immunological self-tolerance. *Nat Immunol.* 2002;3:135-42.
- 54 Lehmann J, Huehn J, de la Rosa M, Maszyna F, Kretschmer U, Krenn V, et al. Expression of the integrin alpha Ebeta 7 identifies unique subsets of CD25+ as well as CD25- regulatory T cells. *Proc Natl Acad Sci U S A.* 2002;99:13031-6.
- 55 Suffia I, Reckling SK, Salay G, Belkaid Y. A role for CD103 in the retention of CD4+CD25+ Treg and control of Leishmania major infection. *J Immunol.* 2005;174:5444-55.
- 56 Tellem A, Mariani M, Lang R, Recalde H, Panina-Bordignon P, Sinigaglia F, et al. Unique chemotactic response profile and specific expression of chemokine receptors CCR4 and CCR8 by CD4(+)CD25(+) regulatory T cells. *J Exp Med.* 2001;194:847-53.
- 57 Lee I, Wang L, Wells AD, Dorf ME, Ozkaynak E, Hancock WW. Recruitment of Foxp3+ T regulatory cells mediating allograft tolerance depends on the CCR4 chemokine receptor. *J Exp Med.* 2005;201:1037-44.
- 58 Huang CT, Workman CJ, Flies D, Pan X, Marson AL, Zhou G, et al. Role of LAG-3 in regulatory T cells. *Immunity.* 2004;21:503-13.
- 59 Garin MI, Chu CC, Golshayan D, Cernuda-Morollon E, Wait R, Lechler RI. Galectin-1: a key effector of regulation mediated by CD4+CD25+ T cells. *Blood.* 2007;109:2058-65.
- 60 Maillard MH, Cotta-de-Almeida V, Takeshima F, Nguyen DD, Michetti P, Nagler C, et al. The Wiskott-Aldrich syndrome protein is required for the function of CD4+CD25+Foxp3+ regulatory T cells. *J Exp Med.* 2007;204:381-91.
- 61 Marangoni F, Trifari S, Scaramuzza S, Panaroni C, Martino S, Notarangelo LD, et al. WASP regulates suppressor activity of human and murine CD4+CD25+FOXP3+ natural regulatory T cells. *J Exp Med.* 2007;204:369-80.
- 62 Fontenot JD, Gavin MA, Rudensky AY. Foxp3 programs the development and function of CD4+CD25+ regulatory T cells. *Nat Immunol.* 2003;4:330-6.
- 63 Hori S, Nomura T, Sakaguchi S. Control of regulatory T cell development by the transcription factor Foxp3. *Science.* 2003;299:1057-61.
- 64 Khattri R, Cox T, Yasayko SA, Ramsdell F. An essential role for Scurfin in CD4+CD25+ T regulatory cells. *Nat Immunol.* 2003;4:337-42.
- 65 Brunkow ME, Jeffery EW, Hjerrild KA, Paepers B, Clark LB, Yasayko SA, et al. Disruption of a new forkhead/winged-helix protein, scurf1, results in the fatal lymphoproliferative disorder of the scurfy mouse. *Nat Genet.* 2001;27:68-73.
- 66 Bennett CL, Christie J, Ramsdell F, Brunkow ME, Ferguson PJ, Whitesell L, et al. The immune dysregulation, polyendocrinopathy, enteropathy, X-linked syndrome (IPEX) is caused by mutations of FOXP3. *Nat Genet.* 2001;27:20-1.
- 67 Chatila TA, Blaeser F, Ho N, Lederman HM, Voulgaropoulos C, Helms C, et al. JM2, encoding a fork head-related protein, is mutated in X-linked autoimmunity-allergic dysregulation syndrome. *J Clin Invest.* 2000;106:R75-81.
- 68 Wildin RS, Ramsdell F, Peake J, Faravelli F, Casanova JL, Buist N, et al. X-linked neonatal diabetes mellitus, enteropathy and endocrinopathy syndrome is the human equivalent of mouse scurfy. *Nat Genet.* 2001;27:18-20.
- 69 Malek TR, Bayer AL. Tolerance, not immunity, crucially depends on IL-2. *Nat Rev Immunol.* 2004;4:665-74.
- 70 Sadlack B, Lohler J, Schorle H, Klebb G, Haber H, Sickel E, et al. Generalized autoimmune disease in interleukin-2-deficient mice is triggered by an uncontrolled activation and proliferation of CD4+ T cells. *Eur J Immunol.* 1995;25:3053-9.
- 71 Suzuki H, Kundig TM, Furlonger C, Wakeham A, Timms E, Matsuyama T, et al. Deregulated T cell activation and autoimmunity in mice lacking interleukin-2 receptor beta. *Science.* 1995;268:1472-6.
- 72 Setoguchi R, Hori S, Takahashi T, Sakaguchi S. Homeostatic maintenance of natural Foxp3+ CD25+ CD4+ regulatory T cells by interleukin (IL)-2 and induction of autoimmune disease by IL-2 neutralization. *J Exp Med.* 2005;201:723-35.
- 73 Nakamura K, Kitani A, Strober W. Cell contact-dependent immunosuppression by CD4(+)CD25(+) regulatory T cells is mediated by cell surface-bound transforming growth factor beta. *J Exp Med.* 2001;194:629-44.
- 74 Grossman WJ, Verbsky JW, Barchet W, Colonna M, Atkinson JP, Ley TJ. Human T regulatory cells can use the perforin pathway to cause autologous target cell death. *Immunity.* 2004;21:589-601.
- 75 Cederbom L, Hall H, Ivars F. CD4+CD25+ regulatory T cells down-regulate co-stimulatory molecules on antigen-presenting cells. *Eur J Immunol.* 2000;30:1538-43.
- 76 Jonuleit H, Schmitt E, Kakirman H, Stassen M, Knop J, Enk AH. Infectious tolerance: human CD25(+) regulatory T cells convey suppressor activity to conventional CD4(+) T helper cells. *J Exp Med.* 2002;196:255-60.
- 77 Letterio JJ, Roberts AB. Regulation of immune responses by TGF-beta. *Annu Rev Immunol.* 1998;16:137-61.
- 78 Moore KW, de Waal Malefyt R, Coffman RL, O'Garra A. Interleukin-10 and the interleukin-10 receptor. *Annu Rev Immunol.* 2001;19:683-765.
- 79 Tivol EA, Borriello F, Schweitzer AN, Lynch WP, Bluestone JA, Sharpe AH. Loss of CTLA-4 leads to massive lymphoproliferation and fatal multiorgan tissue destruction, revealing a critical negative regulatory role of CTLA-4. *Immunity.* 1995;3:541-7.
- 80 Boden E, Tang Q, Bour-Jordan H, Bluestone JA. The role of CD28 and CTLA4 in the function and homeostasis of CD4+CD25+ regulatory T cells. *Novartis Found Symp.* 2003;252:55-63; discussion 63-56, 106-114.
- 81 Annacker O, Pimenta-Araujo R, Burlen-Defranoux O, Barbosa TC, Cumano A, Bandeira A. CD25+ CD4+ T cells regulate the expansion of peripheral CD4 T cells through the production of IL-10. *J Immunol.* 2001;166:3008-3018.
- 82 Asseman C, Fowler S, Powrie F. Control of experimental inflammatory bowel disease by regulatory T cells. *Am J Respir Crit Care Med.* 2000;162:S185-9.
- 83 Kuhn R, Lohler J, Rennick D, Rajewsky K, Muller W. Interleukin-10-deficient mice develop chronic enterocolitis. *Cell.* 1993;75:263-74.
- 84 Maloy KJ, Salaun L, Cahill R, Dougan G, Saunders NJ, Powrie F. CD4+CD25+ T(R) cells suppress innate immune pathology through cytokine-dependent mechanisms. *J Exp Med.* 2003;197:111-9.
- 85 Mottet C, Uhlig HH, Powrie F. Cutting edge: cure of colitis by CD4+CD25+ regulatory T cells. *J Immunol.* 2003;170:3939-43.
- 86 Uhlig HH, Coombes J, Mottet C, Izcue A, Thompson C, Fanger A, et al. Characterization of Foxp3+CD4+CD25+ and IL-10-secreting CD4+CD25+ T cells during cure of colitis. *J Immunol.* 2006;177:5852-60.
- 87 Christ M, McCartney-Francis NL, Kulkarni AB, Ward JM, Mizel DE, Mackall CL, et al. Immune dysregulation in TGF-beta 1-deficient mice. *J Immunol.* 1994;153:1936-46.
- 88 Piccirillo CA, Letterio JJ, Thornton AM, McHugh RS, Mamura M, Mizuhara H, et al. CD4(+)CD25(+) regulatory T cells can mediate suppressor function in the absence of transforming growth factor beta1 production and responsiveness. *J Exp Med.* 2002;196:237-46.
- 89 Powrie F, Carlino J, Leach MW, Mauze S, Coffman RL. A critical role for transforming growth factor-beta but not interleukin 4 in the suppression of T helper type 1-mediated colitis by CD45RB(low) CD4+ T cells. *J Exp Med.* 1996;183:2669-74.
- 90 Fahlen L, Read S, Gorelik L, Hurst SD, Coffman RL, Flavell RA, et al. T cells that cannot respond to TGF-beta escape control by CD4+CD25+ regulatory T cells. *J Exp Med.* 2005;201:737-46.
- 91 Gorelik L, Flavell RA. Abrogation of TGFbeta signaling in T cells leads to spontaneous T cell differentiation and autoimmune disease. *Immunity.* 2000;12:171-81.
- 92 Curotto de Lafaille MA, Muriglan S, Sunshine MJ, Lei Y, Kutchukhidze N, Furtado GC, et al. Hyperimmunoglobulin E response in mice with monoclonal populations of B and T lymphocytes. *J Exp Med.* 2001;194:1349-59.
- 93 Danke NA, Koelle DM, Yee C, Beheray S, Kwok WW. Autoreactive T cells in healthy individuals. *J Immunol.* 2004;172:5967-72.
- 94 Viglietta V, Baecher-Allan C, Weiner HL, Hafler DA. Loss of functional suppression by CD4+CD25+ regulatory T cells in patients with multiple sclerosis. *J Exp Med.* 2004;199:971-9.
- 95 Cao D, van Vollenhoven R, Klareskog L, Trollmo C, Malmstrom V. CD25brightCD4+ regulatory T cells are enriched in inflamed joints of patients with chronic rheumatic disease. *Arthritis Res Ther.* 2004;6:R335-46.
- 96 de Kleer IM, Wedderburn LR, Taams LS, Patel A, Varsani H, Klein M, et al. CD4+CD25bright regulatory T cells actively regulate inflammation in the joints of patients with the remitting form of juvenile idiopathic arthritis. *J Immunol.* 2004;172:6435-43.

- 97 Ehrenstein MR, Evans JG, Singh A, Moore S, Warnes G, Isenberg DA, Mauri C. Compromised function of regulatory T cells in rheumatoid arthritis and reversal by anti-TNF $\alpha$  therapy. *J Exp Med.* 2004;200:277–85.
- 98 Kukreja A, Cost G, Marker J, Zhang C, Sun Z, Lin-Su K, et al. Multiple immuno-regulatory defects in type-1 diabetes. *J Clin Invest.* 2002;109:131–40.
- 99 Makita S, Kanai T, Oshima S, Uraushihara K, Totsuka T, Sawada T, et al. CD4<sup>+</sup>CD25<sup>bright</sup> T cells in human intestinal lamina propria as regulatory cells. *J Immunol.* 2004;173:3119–30.
- 100 Crispin JC, Martinez A, Alcocer-Varela J. Quantification of regulatory T cells in patients with systemic lupus erythematosus. *J Autoimmun.* 2003;21:273–6.
- 101 Kriegel MA, Lohmann T, Gabler C, Blank N, Kalden JR, Lorenz HM. Defective Suppressor Function of Human CD4<sup>+</sup>CD25<sup>+</sup> Regulatory T Cells in Autoimmune Polyglandular Syndrome Type II. *J Exp Med.* 2004;199:1285–91.
- 102 Balandina A, Lecart S, Darteville P, Saoudi A, Berrih-Aknin S. Functional defect of regulatory CD4<sup>(+)</sup>CD25<sup>+</sup> T cells in the thymus of patients with autoimmune myasthenia gravis. *Blood.* 2005;105:735–41.
- 103 Ling EM, Smith T, Nguyen XD, Pridgeon C, Dallman M, Arbery J, et al. Relation of CD4<sup>+</sup>CD25<sup>+</sup> regulatory T-cell suppression of allergen-driven T-cell activation to atopic status and expression of allergic disease. *Lancet.* 2004;363:608–15.
- 104 Karlsson MR, Rugtveit J, Brandtzaeg P. Allergen-responsive CD4<sup>+</sup>CD25<sup>+</sup> regulatory T cells in children who have outgrown cow's milk allergy. *J Exp Med.* 2004;199:1679–88.
- 105 Cavani A, Nasorri F, Ottaviani C, Sebastiani S, De Pita O, Girolomoni G. Human CD25<sup>+</sup> regulatory T cells maintain immune tolerance to nickel in healthy, nonallergic individuals. *J Immunol.* 2003;171:5760–8.
- 106 Ou LS, Goleva E, Hall C, Leung DY. T regulatory cells in atopic dermatitis and subversion of their activity by superantigens. *J Allergy Clin Immunol.* 2004;113:756–63.
- 107 Mills KH. Regulatory T cells: friend or foe in immunity to infection? *Nat Rev Immunol.* 2004;4:841–55.
- 108 Pasare C, Medzhitov R. Toll pathway-dependent blockade of CD4<sup>+</sup>CD25<sup>+</sup> T cell-mediated suppression by dendritic cells. *Science.* 2003;299:1033–6.
- 109 Aandahl EM, Michaelsson J, Moretto WJ, Hecht FM, Nixon DF. Human CD4<sup>+</sup> CD25<sup>+</sup> regulatory T cells control T-cell responses to human immunodeficiency virus and cytomegalovirus antigens. *J Virol.* 2004;78:2454–9.
- 110 Cabrera R, Tu Z, Xu Y, Firpi RJ, Rosen HR, Liu C, Nelson DR. An immunomodulatory role for CD4<sup>(+)</sup>CD25<sup>(+)</sup> regulatory T lymphocytes in hepatitis C virus infection. *Hepatology.* 2004;40:1062–71.
- 111 Kinter AL, Hennessey M, Bell A, Kern S, Lin Y, Daucher M, et al. CD25<sup>(+)</sup>CD4<sup>(+)</sup> regulatory T cells from the peripheral blood of asymptomatic HIV-infected individuals regulate CD4<sup>(+)</sup> and CD8<sup>(+)</sup> HIV-specific T cell immune responses in vitro and are associated with favorable clinical markers of disease status. *J Exp Med.* 2004;200:331–43.
- 112 Suvas S, Kumaraguru U, Pack CD, Lee S, Rouse BT. CD4<sup>+</sup>CD25<sup>+</sup> T cells regulate virus-specific primary and memory CD8<sup>+</sup> T cell responses. *J Exp Med.* 2003;198:889–901.
- 113 Toka FN, Suvas S, Rouse BT. CD4<sup>+</sup> CD25<sup>+</sup> T cells regulate vaccine-generated primary and memory CD8<sup>+</sup> T-cell responses against herpes simplex virus type 1. *J Virol.* 2004;78:13082–9.
- 114 Belkaid Y, Piccirillo CA, Mendez S, Shevach EM, Sacks DL. CD4<sup>+</sup>CD25<sup>+</sup> regulatory T cells control Leishmania major persistence and immunity. *Nature.* 2002;420:502–7.
- 115 Belkaid Y, Rouse BT. Natural regulatory T cells in infectious disease. *Nat Immunol.* 2005;6:353–60.
- 116 Golshayan D, Buhler L, Lechler RI, Pascual M. From current immunosuppressive strategies to clinical tolerance of allografts. *Transpl Int.* 2007;20:12–24.
- 117 Golshayan D, Jiang S, Tsang J, Garin MI, Mottet C, Lechler RI. In vitro-expanded donor alloantigen-specific CD4<sup>+</sup>CD25<sup>+</sup> regulatory T cells promote experimental transplantation tolerance. *Blood.* 2007;109:827–35.
- 118 Rieger K, Loddenkemper C, Maul J, Fietz T, Wolff D, Terpe H, et al. Mucosal FOXP3<sup>+</sup> regulatory T cells are numerically deficient in acute and chronic GvHD. *Blood.* 2006;107:1717–23.
- 119 Zorn E, Kim HT, Lee SJ, Floyd BH, Litsa D, Arumugarajah S, et al. Reduced frequency of FOXP3<sup>+</sup> CD4<sup>+</sup>CD25<sup>+</sup> regulatory T cells in patients with chronic graft-versus-host disease. *Blood.* 2005;106:2903–11.
- 120 Ichihara F, Kono K, Takahashi A, Kawaida H, Sugai H, Fujii H. Increased populations of regulatory T cells in peripheral blood and tumor-infiltrating lymphocytes in patients with gastric and esophageal cancers. *Clin Cancer Res.* 2003;9:4404–8.
- 121 Liyanage UK, Moore TT, Joo HG, Tanaka Y, Herrmann V, Doherty G, et al. Prevalence of regulatory T cells is increased in peripheral blood and tumor microenvironment of patients with pancreas or breast adenocarcinoma. *J Immunol.* 2002;169:2756–61.
- 122 Viguier M, Lemaitre F, Verola O, Cho MS, Gorochov G, Dubertret L, et al. Foxp3 expressing CD4<sup>+</sup>CD25<sup>(high)</sup> regulatory T cells are overrepresented in human metastatic melanoma lymph nodes and inhibit the function of infiltrating T cells. *J Immunol.* 2004;173:1444–53.
- 123 Woo EY, Yeh H, Chu CS, Schlienger K, Carroll RG, Riley JL, et al. Cutting edge: Regulatory T cells from lung cancer patients directly inhibit autologous T cell proliferation. *J Immunol.* 2002;168:4272–6.
- 124 Jiang S, Camara N, Lombardi G, Lechler RI. Induction of allopeptide-specific human CD4<sup>+</sup>CD25<sup>+</sup> regulatory T cells ex vivo. *Blood.* 2003;102:2180–6.
- 125 Tang Q, Henriksen KJ, Bi M, Finger EB, Szot G, Ye J, et al. In vitro-expanded antigen-specific regulatory T cells suppress autoimmune diabetes. *J Exp Med.* 2004;199:1455–65.
- 126 Tarbell KV, Yamazaki S, Olson K, Toy P, Steinman RM. CD25<sup>+</sup> CD4<sup>+</sup> T cells, expanded with dendritic cells presenting a single autoantigenic peptide, suppress autoimmune diabetes. *J Exp Med.* 2004;199:1467–77.
- 127 Battaglia M, Stabellini A, Roncarolo MG. Rapamycin selectively expands CD4<sup>+</sup>CD25<sup>+</sup>FoxP3<sup>+</sup> regulatory T cells. *Blood.* 2005;105:4743–8.

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