



REVIEW ARTICLE

The α_1 -adrenergic receptors: diversity of signaling networks and regulation

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Abstract

The α_1 -adrenergic receptor (AR) subtypes (α_{1a} , α_{1b} , and α_{1d}) mediate several physiological effects of epinephrine and norepinephrine. Despite several studies in recombinant systems and insight from genetically modified mice, our understanding of the physiological relevance and specificity of the α_1 -AR subtypes is still limited. Constitutive activity and receptor oligomerization have emerged as potential features regulating receptor function. Another recent paradigm is that β arrestins and G protein-coupled receptors themselves can act as scaffolds binding a variety of proteins and this can result in growing complexity of the receptor-mediated cellular effects. The aim of this review is to summarize our current knowledge on some recently identified functional paradigms and signaling networks that might help to elucidate the functional diversity of the α_1 -AR subtypes in various organs.

Keywords: Constitutive activity; oligomerization; β -arrestin; protein interactions; knock out mice; transgenic mice

Introduction

Within the large family of G protein-coupled receptors (GPCR), the adrenergic receptors (ARs) mediate the functional effects of catecholamines, like epinephrine and norepinephrine. The AR family includes nine different gene products, three β (β_1 , β_2 , β_3), three α_1 (α_{1a} , α_{1b} , and α_{1d}), and three α_2 (α_{2A} , α_{2B} , and α_{2C}) receptor subtypes.

The α_1 -AR subtypes are expressed in various organs, including brain, heart, blood vessels, liver, kidney, prostate, and spleen, in which they mediate a variety of functional effects such as modulation of neurotransmission, vasoconstriction, cardiac inotropy, and chronotropy, regulation of metabolism (reviewed in ref. 1). Activation of the three α_1 -AR subtypes causes polyphosphoinositide hydrolysis catalyzed by phospholipase C (PLC) via pertussis toxin-insensitive G proteins in most tissues where this effect has been examined (1).

Radioligand binding studies in rat tissues initially demonstrated two classes of α_1 -AR binding sites, "A" and "B" with high and low affinity for the α_1 -AR antagonists

WB4101 and phentolamine, respectively. The first α_1 -AR cloned, was unequivocally assigned to the pharmacological α_{1b} subtype and hence named α_{1b} -AR. The pharmacological α_{1A} subtype, today identified as α_{1a} -AR, was initially cloned from a bovine brain library and inappropriately named α_{1C} -AR or $\alpha_{1A/C}$ -AR. Finally, the cloned α_{1d} -AR (initially named α_{1A} -AR or $\alpha_{1A/D}$ -AR) was a novel receptor subtype not clearly identified by previous pharmacological studies (reviewed in ref. 2,3).

Studies aiming to assess the specific functional responses mediated by distinct α_1 -AR subtypes have been hampered by the fact that the subtype-selective drugs are only moderately selective. Recently, studies on genetically modified mice lacking or overexpressing one or more α_1 -AR subtypes have provided some important insight into the functional roles played by distinct receptors. However, our understanding on the functional implications of α_1 -AR heterogeneity in physiological systems is still quite limited.

Extensive mutational analysis performed by our group and other investigators helped to identify the structural

determinants of the α_1 -AR subtypes involved in each of the three main "classical" functional properties of GPCRs: (i) ligand-binding; (ii) receptor activation and coupling to G protein; and (iii) desensitization. These findings have been reviewed elsewhere (4,5). Beyond these "classical" features, a number of novel functional paradigms have been recently described for GPCRs including receptor constitutive activity (6), oligomerization (7) and interaction with a variety of signaling proteins (8). These functional features imply a growing complexity of signaling and regulation of the α_1 -AR subtypes which might represent the mechanistic basis of their functional specificity in various tissues.

The aim of this review is to summarize our current knowledge on some recently identified functional paradigms and signaling networks that might help to elucidate the functional diversity of the α_1 -AR subtypes in various organs.

Constitutive activity of the α_1 -AR subtypes

For both the α_{1a} and α_{1b} -AR mutation-induced and spontaneous constitutive activity have been reported (9,10). Interestingly, most of the known α -blockers behave as inverse agonists both at the wild type and constitutively active mutants of the two receptors (10). Studies on constitutively activating mutations of the α_{1b} -AR provided important insight into the potential molecular mechanisms of GPCR activation (11). In particular, they highlighted the highly conserved E/DRY sequence at the cytosolic end of helix 3 as an important switch of receptor activation.

Interestingly, activating mutations which perturb the helix 3/helix 6 packing of the receptor have been found in both the α_{1a} and α_{1b} -AR subtypes suggesting common mechanisms of receptor activation (12). These include: (i) mutations of A293^(6,34) and of A271^(6,34) in the cytosolic extension of helix 6 in the α_{1b} -AR and α_{1a} -AR, respectively (9,10); (ii) mutations of D142^(3,49) and D123^(3,49) of the E/DRY motif in the α_{1b} -AR and α_{1a} -AR, respectively (10,11).

However, some differences in the activation properties can be observed between the α_{1a} and α_{1b} -AR in recombinant systems measuring the inositol phosphate response. The agonist-independent activity of both the wild type α_{1b} -AR and its constitutively active mutants was significantly higher than that of the wild type α_{1a} -AR or its mutant. In contrast, the epinephrine-induced inositol phosphate accumulation above basal at the α_{1a} -AR was significantly higher than that at the α_{1b} -AR or its constitutively active mutants expressed at comparable levels (10,13). This suggests that in recombinant systems the agonist-occupied α_{1a} -AR has greater efficacy in activating PLC than the α_{1b} -AR whereas its spontaneous or

mutation-induced isomerization toward the active states is lower. Only one study reported the opposite showing that in rat neonatal cardiomyocytes a different constitutively active mutant of the α_{1a} -AR displayed higher basal activity than the analogous mutant of the α_{1b} -AR (14). This finding is intriguing and should be further explored.

The properties of the α_{1d} -AR subtype have been investigated very little because its expression as well as the agonist-induced inositol phosphate response mediated by this receptor were often found to be much smaller than those of the other two subtypes (15,16). Constitutively activating mutations of the α_{1d} -AR have not been reported so far. However, an interesting study reported that the α_{1d} -AR expressed in rat fibroblasts is constitutively active and internalized (17). In fact, the basal activity of the α_{1d} -AR was 2-fold greater than that of the α_{1b} -AR and was increased following the treatment with the inverse agonist prazosin which caused its redistribution from the intracellular compartments to the plasma membrane. The constitutive activity of the α_{1d} -AR was also observed in physiological systems like in aorta and mesenteric arteries where it could be inhibited by inverse agonists (18). For the α_{1a} or α_{1b} -AR constitutive activity in physiological systems has not been investigated.

Altogether, these findings indicate that there might be important differences in the constitutive activity of the α_1 -AR subtypes which could have consequences in their signaling and regulatory properties *in vivo*. Such differences should be further explored and the elucidation of their physiological implications might represent an important area of investigation.

Oligomerization of the α_1 -AR subtypes

Findings in the last decade challenged the widely held view of GPCRs functioning as monomeric units. Co-immunoprecipitation of differentially tagged GPCRs or functional complementation of pairs of co-expressed inactive receptor mutants provided strong evidence that GPCR oligomers do exist. The widespread use of biophysical techniques such as fluorescence resonance energy transfer (FRET) or bioluminescence resonance energy transfer (BRET) between GPCRs carrying the appropriate pair of fluorescent/bioluminescent labels suggested oligomerization of a variety of GPCRs. Each technique employed has its own shortcomings: whereas co-immunoprecipitation cannot rule out indirect interaction, energy transfer techniques can only certify that the two partners are in close proximity, not necessarily in immediate contact. However, convergent results obtained through independent methods eventually led to the widespread acknowledgment of the existence of GPCR oligomers (7).

Both homo- and hetero-oligomerization have been demonstrated for the three α_1 -AR subtypes in recombinant systems (Table 1) (15,16,19). FRET measurements as well as co-immunoprecipitation experiments provided evidence that both the α_{1a} and α_{1b} -AR can form homo-oligomers (19). Oligomerization of the α_{1b} -AR did not require the integrity of its C-tail, of two glycoporphin motifs or of the N-linked glycosylation sites at its N-terminus. Constitutively active or non-functional α_{1b} -AR mutants displayed the same propensity to oligomerize as the wild-type receptor, indicating that the activation state of the receptor is irrelevant for this process. Receptor oligomerization was not influenced by the agonist epinephrine or by the inverse agonist prazosin. Thus, whether homo-oligomerization of the α_{1a} or α_{1b} -AR has any functional relevance is unknown.

Hetero-oligomers were observed between the α_{1a} and the α_{1b} -AR subtypes, but not between the α_{1b} -AR and other GPCRs. Interestingly, hetero-oligomerization was found to have an impact on receptor endocytosis (19). Whereas the α_{1b} -AR undergoes agonist-induced internalization, the α_{1a} -AR does not. However, when the two AR subtypes were co-expressed forming heterodimers, the endocytosis of each monomer could be triggered by stimulation of the other. Colocalization of the two monomers could be seen in endocytic vesicles suggesting that the α_{1a}/α_{1b} dimers remained stable throughout the endocytosis process.

An important effect of hetero-oligomerization has been reported for the α_{1d} -AR subtype. In fact, co-expression of the α_{1d} -AR with the α_{1b} -AR (15) or the β_2 -AR (19) was able to rescue surface expression of the α_{1d} -AR, the majority of which is intracellular when expressed alone in various cell lines. Interestingly, the interaction with the α_{1b} -AR modified the pharmacological profile of the α_{1d} -AR which loses its affinity for its selective ligand BMY7378 when it is co-expressed with the α_{1b} -AR. The α_{1b}/α_{1d} dimer behaves as a single functional entity with increased response to norepinephrine relative to either monomer alone. The α_{1d} -AR receptor was long supposed to be little expressed in the heart, as its selective ligand BMY7378 could detect only minimal levels of the receptor. However, these findings should be considered in a new light, given that the α_{1b} and α_{1d} -AR subtypes co-exist in various tissues and the pharmacological profile of the α_{1d} -AR might be different than expected because of oligomerization.

Oligomerization of α_1 -AR subtypes in physiological systems has not been explored so far for lack of appropriate experimental tools. Therefore, the functional relevance of α_1 -AR oligomerization *in vivo* remains elusive. However, oligomerization might represent an additional mechanism regulating the physiological responses mediated by the α_1 -AR subtypes which are often co-expressed in the same cells. Further exploring the functional correlates of receptor oligomerization and assessing if it

occurs in physiological systems might provide interesting information about cross-talk effects at the level of α_1 -AR signaling or regulation.

Signaling pathways of the α_1 -AR subtypes

It has become increasingly evident that the variety of functional effects mediated by the α_1 -ARs in different organs must imply the activation of multiple signaling pathways beyond activation of PLC via Gq/11. Therefore, several studies have attempted to investigate whether each α_1 -AR subtype may activate distinct signaling pathways, but our knowledge on this issue is still limited.

It has been reported that stimulation of the α_{1b} and α_{1d} -AR can result in the activation of phospholipase A2 in COS-1 cells (20); the α_{1a} -AR was not explored. In NIH3T3 cells, the activation of the α_{1a} and α_{1b} -AR, but not that of the α_{1d} , resulted in the stimulation of p21-ras, PI3-kinase and mitogen-activated protein kinase (MAPK) (21). However, the steps leading to the activation of these pathways seem to differ between the two receptor subtypes.

In hepatocyte derived cells, stimulation of the α_{1b} -AR subtype inhibits interleukin 6 signaling by a MAPK mechanism (22). An interesting microarray study indicated that the α_1 -AR subtypes expressed in Rat fibroblasts have a differential effect on cell cycle genes with the α_{1b} mediating cell-cycle progression, and the α_{1a} and α_{1d} -AR mediating G1-S cell cycle arrest (23).

Most of the work investigating α_1 -AR signaling has been performed in cardiomyocytes. In fact, hearts of most species express both α_{1a} and α_{1b} -AR at protein level whereas the expression of α_{1d} -AR is very low. The α_{1a} -AR predominates in humans, whereas the α_{1b} -AR in rodents. Some seminal studies (24,25) demonstrated that stimulation of the α_1 -ARs in cardiomyocytes induces a hypertrophic response accompanied by the activation of early genes (*c-fos*, *c-jun*, *egr-1*) upregulation of contractile proteins (myosin light chain-2) and reactivation of embryonic genes (atrial natriuretic factor (ANF), β -myosin heavy chain, skeletal α -actin).

Various studies provided clear evidence for the involvement of both the PLC-MAPK pathway (26) and

Table 1. Oligomerization of the α_1 -adrenergic receptor subtypes.

Receptors	Trafficking	Pharmacology	Signaling	Ref.
α_{1a}/α_{1b}	Co-endocytosis	No change	—	19
α_{1b}/α_{1d}	↑ α_{1d} Surface expression	↓ α_{1d} Affinity for selective ligands	↑ Signaling	15
α_{1d}/β_2	↑ α_{1d} Surface expression co-endocytosis	—	—	16
Homo-oligomers	—	—	—	15,19
$\alpha_{1a}, \alpha_{1b}, \alpha_{1d}$				

Rho-signaling (27) in the α_1 -AR-induced hypertrophic response in cardiomyocytes. A recent study supports these earlier findings suggesting that α_1 -AR-induced cardiac hypertrophy is mediated by three parallel pathways: G12/13-Rho-JNK, Gq-JNK (Rho-independent) and G $\beta\gamma$ (JNK independent) (28). Recent findings have demonstrated that the α_1 -ARs endogenously expressed in rat neonatal cardiomyocytes promote RhoA-activation via a mechanism that requires G12 and the Rho-guanine nucleotide exchange factor AKAP-Lbc and this pathway mediates hypertrophy (29).

The respective role in stimulating cardiac hypertrophy of the two α_1 -AR subtypes expressed in heart, the α_{1a} and α_{1b} -AR, does not emerge clearly from the *in vitro* studies published so far probably because of the limited selectivity of the pharmacological tools available. In one study on rat neonatal cardiomyocytes, a constitutively active form of the α_{1a} -AR activated gene expression of the ANF, whereas the analogous constitutively active mutant of the α_{1b} -AR stimulated gene expression of c-fos, but not of ANF (14). However, these findings are intriguing considering that other studies reported the opposite and that overexpression of the α_{1b} -AR in transgenic mice resulted in a marked increase in ANF (see below). In the future, it would be interesting to carry on a systematic investigation of different signaling pathways comparing the α_1 -AR subtypes expressed in the same cellular systems and to correlate these findings with the growing information provided by *in vivo* studies on genetically modified mice (see below).

Regulatory mechanisms and β arrestin interaction at the α_1 -AR subtypes

The α_1 -AR subtypes display quite divergent regulatory properties. In fact, the α_{1b} -AR in recombinant systems undergoes rapid phosphorylation, desensitization and endocytosis upon exposure to the agonist (30–32). Desensitization involves phosphorylation of residues in the C-tail of the receptor mediated by G protein-coupled receptor kinases (GRKs) (31). The endocytosis of the α_{1b} -AR occurs via clathrin-coated vesicles and seems to involve β arrestins (32).

In contrast, the α_{1a} -AR expressed in rat-1 fibroblasts is poorly phosphorylated and desensitized compared to the α_{1b} -AR (33). In addition, it undergoes very modest agonist-induced endocytosis (32).

Fewer studies have investigated the desensitization of the α_{1d} -AR probably because of its poor expression in recombinant systems. It has been reported that noradrenaline and direct activation of protein kinase C induce phosphorylation of the α_{1d} -AR and this correlates with desensitization of the receptor (34). However,

desensitization of the α_{1d} -AR was not compared with that of the other two subtypes in this study.

Overall, the impact of α_1 -AR desensitization in physiological systems where the receptors are endogenously expressed has been poorly investigated, as it is the case for most GPCRs. Therefore, what is the impact of different regulatory properties of the α_1 -AR subtypes on complex functions like vasoconstriction, metabolic response, and others, is unknown.

Interestingly, the different regulatory features of the α_{1a} and α_{1b} -AR seem to correlate with their pattern of interaction with β arrestins. In fact, the results from both co-immunoprecipitation experiments and β arrestin translocation assays indicated that the agonist-induced interaction of the α_{1a} -AR with β arrestin was much weaker than that of the α_{1b} -AR (32). The interaction of β arrestin with the α_{1d} -AR has not been directly explored so far.

These differences in receptor/ β arrestin interaction might have broader implications in α_1 -AR mediated signaling because of the well established role played by β arrestins in coordinating a variety of signaling networks (35). In particular, it is well established that β arrestins are scaffolds for components of the MAPK cascade thus mediating MAPK activation induced by various GPCRs. Investigation of β arrestin-mediated signaling at the α_1 -AR subtypes is an interesting area of investigation which has been poorly explored so far and might represent one of the mechanisms contributing to the variety of the receptor-mediated-responses.

Proteins interacting with different α_1 -AR subtypes

One of the most recent paradigms is that GPCRs can bind a variety of proteins and this can promote multiple signaling events which results in growing complexity of the receptor-mediated cellular effects (8)

A number of approaches have been followed to identify novel proteins interacting with the α_1 -ARs, including yeast two-hybrid screen using cytosolic portions of the receptors as bait, pull-down or *in vitro* overlay assays using purified proteins, co-immunoprecipitation of receptor-protein complexes from recombinant or native cells, FRET or BRET technology in cells. These studies resulted in the identification of a variety of proteins interacting with the α_1 -AR subtypes, several of them in a receptor subtype selective pattern (Table 2).

The α_{1a} -AR subtype contains a PDZ binding sequence G-E-E-V at its C-terminus that can be expected to give rise to PDZ-domain mediated interactions. An early report, at the issue of a yeast two-hybrid screen, identified the type III PDZ domain of nNOS (neuronal nitric oxide synthase) as a potential α_{1a} -AR interacting protein (36) However,

Table 2. Proteins interacting with the α_1 -adrenergic receptor subtypes.

Receptor	Partner	Binding site	Functional role	Ref.
α_{1a} α_{1b} α_{1d}	nNOS	Unknown	Unknown	36
α_{1a}	Tolloid	C-tail	↓ Surface expression	37
α_{1a}	RGS2	i3 loop(K219-S220-R238)	↓ Gq signaling	38
α_{1b}	AP50	C-tail (8 Arg)	↑ Endocytosis	40
α_{1b}	Ezrin	C-tail (8 Arg)	↑ Recycling	41
α_{1b}	Spinophilin	i3 loop	↓ Ca ²⁺ signaling induced by RGS2	43
α_{1d}	Syntrophins	C-term (ETDI)	Stabilization of receptor at cell surface	44
α_{1b} α_{1d}	gC1qR	C-tail (Arg)	Unknown	42

co-immunoprecipitation studies, while confirming this interaction, failed to highlight selectivity for the α_{1a} -AR subtype since all three α_1 -AR subtypes could be co-immunoprecipitated with nNOS and this even when they were lacking their C-terminus. This interaction appeared to be without apparent physiological implications in spite of the known role of NO in the regulation of blood pressure and of nNOS as local metabolic inhibitor of α_1 -AR-mediated vasoconstriction.

Another study reported that the CUB5 domain of mammalian tolloid (mTLD), a zinc-finger matrix metalloprotease of the astacin family, interacted with α_{1a} -AR C-tail in a yeast two hybrid screen (37). Overexpression of mTLD reduced the number of cell surface receptors without affecting total receptor level or affinity when transiently expressed in HEK293 cells. No mechanism was proposed to account for the observed phenomena.

Interesting prospects were opened by the report of the direct interaction between RGS2 (regulator of G protein signaling 2) and the third intracellular loop of the α_{1a} -AR (38). RGS proteins are well characterized inhibitors of heterotrimeric G protein function, acting as GAPs (GTPase activating proteins) to increase the rate of GTP hydrolysis at G α subunits and thus terminate signaling. More than 30 RGS proteins have been identified so far, but many RGS proteins can non-selectively bind to and inhibit G $\alpha i/o$ and G $\alpha q11$ in reconstituted systems, suggesting that other factors may regulate their specificity for a particular signaling pathway. RGS2 was found to interact with the α_{1a} -AR third intracellular loop confirming what previously shown for other Gq-coupled receptors, namely the cholinergic muscarinic M1, M3, and M5 receptors (39) and it inhibited agonist-induced inositol phosphate responses without affecting ligand binding.

Two main interacting partners were pulled out of a yeast two-hybrid screen for the α_{1b} -AR: the $\mu 2$ (or AP50) subunit of the clathrin adaptor complex AP2 (40) and ezrin, a member of the ezrin-radixin-moesin (ERM) protein family (41). The AP2 complex is part of the endocytic machinery mediating clathrin-dependent

endocytosis of membrane proteins and it is recruited to agonist-activated GPCRs through the intermission of β arrestins. Binding of AP50 relied on a basic stretch of eight arginines in the proximal C-tail of the receptor. Direct association of the α_{1b} -AR to AP50 contributed to the agonist-induced internalization of the receptor as demonstrated by the fact that a receptor mutant lacking the AP50 binding motif was delayed in internalization. The presence of the eight arginine motif in the C-tail of a GPCR is not common, which rules out the hypothesis that direct AP50 interaction is a common mechanism for clathrin-mediated endocytosis. Interestingly, this feature is shared by the α_{1d} -AR, which contains a stretch of seven positive charges in its C-tail, but no studies were undertaken using this receptor subtype.

In addition to AP50, the same yeast two-hybrid screen identified ezrin as a potentially direct binding partner of the α_{1b} -AR (41). Ezrin belongs to the ERM family of proteins, primarily described as linkers between membrane proteins and cortical actin. Ezrin interactions with polytopic membrane proteins generally occur through the adaptor proteins EBP50 (NHERF1) and E3KARP (NHERF2). So far, a role for the ERM proteins in GPCR trafficking was inferred from the finding that NHERF1 binding to some GPCRs promoted their recycling, depending on its binding to ERM proteins. The α_{1b} -AR is the first GPCR for which a direct interaction with ezrin has been found. Disruption of this interaction by overexpression of a dominant negative mutant of ezrin inhibited receptor recycling after internalization, as did actin depolymerization. However, ezrin was also shown to be involved in the remodelling of the actin cytoskeleton, in the modulation of Rho-signaling (by binding to Rho-GTP dissociation inhibitor and through direct association to several Rho-GTP/GDP exchange factors) as well as in anchoring of protein kinase A. Therefore, it would be interesting to test whether ezrin is also involved in ρ -signaling mediated by the α_{1b} -AR.

Another protein, the receptor for globular "Heads" of c1q (gC1qR), was reported to interact with the same arginine-rich sequence in the α_{1b} and the α_{1d} -AR (42). gC1qR is a glycoprotein mainly displaying intracellular localization, but also present on the surface of macrophages and T cells through anchoring to β -integrin, where it is part of a complement receptor. No functional relevance was demonstrated for its interaction with the α_1 -ARs.

An interesting protein found to interact with the α_{1b} -AR is spinophilin which interacts with other GPCRs, including the α_2 -AR, as well as with the N-terminal domain of RGS proteins (RGS1, 2, 4, and 16) which participates in GPCR recognition (43). Thus spinophilin might represent an interesting functional bridge between RGS and α_1 AR subtypes that don't bind RGS, like the α_{1b} AR. In fact, it has been found that spinophilin increases the RGS2-induced

inhibition of the α_{1b} -AR calcium response. These data offer a glimpse into a potentially more general regulatory mechanisms of GPCR function by spinophilin.

The α_{1d} -AR was for a long time a “poor relative” to the other α_1 -AR subtypes, the α_{1a} and α_{1b} because poorly expressed at the cell surface in heterologous systems, probably because of its long N-terminus. This peculiarity hampered the investigation of its potential interactions with other proteins. Apart from the above mentioned interaction with gC1qR, whose functional implications are unknown (42), another interacting partner of the α_{1d} -AR was α -syntrophin (44). α -syntrophin, a protein containing one PDZ domain and two PH (pleckstrin homology) domains, specifically recognized the C-tail of the α_{1d} -AR, but not that of the α_{1a} or α_{1b} , in the yeast two-hybrid assay. The PDZ domains of syntrophin isoforms α , $\beta 1$, and $\beta 2$, but not $\gamma 1$ or $\gamma 2$, could interact with the α_{1d} -AR C-tail. The α_{1d} -AR possesses the C-terminal sequence E-T-D-I, whose mutation impaired syntrophin binding to the receptor and markedly decreased norepinephrine-induced inositol phosphate accumulation. This mutation also dramatically decreased receptor expression levels. Taken altogether these results suggested that syntrophins act to maintain the stability of the α_{1d} -AR through a PDZ-mediated interaction.

Altogether these findings indicate a rather complex and heterogeneous pattern of receptor/protein interactions whose physiological implications are far from being fully elucidated. The direct interaction of α_1 -AR subtypes with selected partners identified in recombinant systems might result in new mechanisms of receptor signaling and regulation. Since these mechanisms might be specific for distinct receptors or cell types, the study of these interactions is an interesting approach to better understand the functional specificity of the receptors. However, this would require a systematic proteomic approach in different tissues expressing the α_1 -AR subtypes as well as good experimental tools to investigate its functional implications.

Insights from genetically modified mice

Recently, mouse lines carrying genetic modifications of the α_1 -AR subtypes have provided interesting information on the *in vivo* functions of the receptors giving some insight into the specificity of their role. The α_{1b} -AR knock out (KO) mouse was the first model to be created (45) and it was characterized for a number of functional parameters. The α_{1b} -KO mice displayed: (i) decreased blood pressure response to phenylephrine with normal resting pressure (45); (ii) hyperinsulinemia, insulin resistance and high fat diet-induced obesity (46); and (iii) behavioral changes including blunted locomotor response to drugs of abuse and markedly decreased sensitivity to morphine and cocaine (47). Other mice carrying genetic modifications

of the α_1 -AR subtypes have been mainly characterized for their cardiovascular phenotype (Table 3) thus allowing to build a more comprehensive picture of the functional role of each receptor in the cardiovascular system.

Both the α_{1a} and α_{1d} -AR KO mice displayed decreased resting blood pressure as well as phenylephrine stimulated pressure response (48,49). The fact that the acute response to phenylephrine is decreased in all three KO mice indicates that the α_{1a} , α_{1b} and α_{1d} -AR all contribute to the regulation of the vascular tone. However, the contribution of the α_{1a} and α_{1d} -AR subtypes is prominent because deletion of either one of the two receptors leads also to decreased resting blood pressure. This can be explained by the fact that the α_{1a} -AR prevails in distributing arteries (mesenteric, renal) (48) and the α_{1d} -AR in large conducting arteries (aorta and carotid) (49), whereas the expression of the α_{1b} -AR is minor in all arteries.

Studies on genetically modified mice have also provided interesting insight into the role of the α_1 -AR in cardiac function and hypertrophy. As mentioned above, the α_{1a} and α_{1b} -AR subtypes are both expressed in cardiomyocytes with the α_{1a} predominating in humans and the α_{1b} in rodents. Transgenic mice overexpressing a constitutively active α_{1b} -AR mutant specifically in the heart display cardiac hypertrophy without any change in blood pressure (50). This supports previous evidence that stimulation of α_1 -ARs in cardiomyocytes *in vitro* leads to a hypertrophic response (24). This finding is also consistent with the role played by the Gq/PLC pathway in heart as demonstrated by the fact that transgenic mice overexpressing a constitutively active G α_q develop cardiac hypertrophy (51).

Interestingly, another transgenic mouse overexpressing a different constitutively active α_{1b} -AR mutant, under the control of the receptor own promoter, displayed a more complex phenotype characterized by cardiac hypertrophy as well as autonomic failure (52). This confirms a direct role of the α_{1b} -AR in cardiac hypertrophy, but indicates that broader effects occur when the receptor is generally overexpressed.

Mice overexpressing constitutively active mutant of the α_{1a} -AR subtype have not been generated. However, the role of the α_{1a} -AR in heart growth *in vivo* has been demonstrated by studies on double KO mice carrying deletions of both the α_{1a} and α_{1b} -AR (53) which displayed several abnormalities including: (i) reduced growth of the heart after birth; (ii) reduced cardiac output; and (iii) increased mortality after pressure overload. These findings demonstrate that both the α_{1a} and α_{1b} -AR play an important role in heart growth after birth and their integrity is required to maintain correct heart function.

These changes were, however, sex specific since they were observed in males, but not in females. This might be explained by the fact that females have a lower sympathetic tone and the growth of their hearts is less dependent on the α_1 -ARs.

Cardiac hypertrophy was not observed in transgenic mice with cardiac-specific overexpression of the wild type α_{1a} or α_{1b} -AR subtype (54,55) despite the fact that they displayed increased expression of ANF. This is unlike the phenotype of mice overexpressing the constitutively active α_{1b} -AR mutant (50). This difference might be due to the fact that the signaling of a constitutively active mutant is somehow different or has greater efficacy than that of the wild-type receptor.

However, transgenic mice overexpressing either the α_{1a} or α_{1b} -AR subtype in the heart provided a number of novel findings on the functional role of these receptors in heart. In fact, in the heart of the α_{1b} -AR transgenic mice left ventricular contraction in response to β -agonists was depressed (55). Interestingly, it was found that dampening of β -AR signaling through adenylate cyclase was due to activation of a pertussis-sensitive inhibitory G protein. This clearly suggests that when overexpressed α_1 -ARs can couple to inhibitory G proteins.

In conclusion, as summarized in Figure 1, studies on mice carrying genetic modifications of the α_1 -AR genes have provided evidence that: (a) all three α_1 -AR

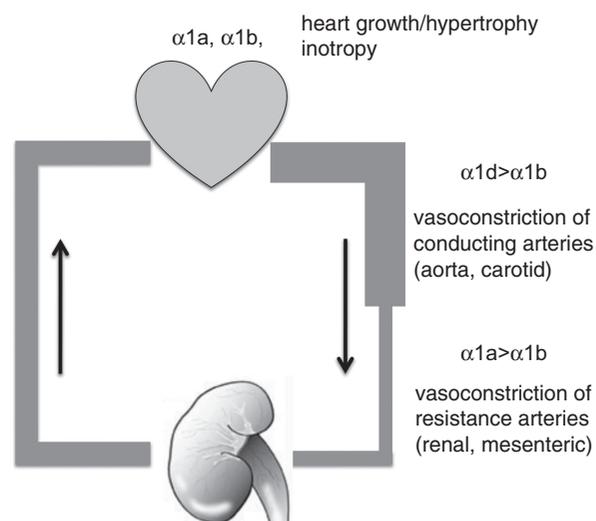


Figure 1. The α_1 -adrenergic receptor subtypes in the cardiovascular system. This figure summarizes the main roles played by distinct α_1 -AR subtypes in the cardiovascular system highlighted by studies on genetically modified mice.

Table 3. Cardiovascular phenotype of mice carrying genetic modifications of different α_1 -adrenergic receptor subtypes.

Receptor	Genetic modification	Phenotype	Ref.
α_{1b}	Gene deletion	↓ Resting blood pressure ↓ Blood pressure response to phenylephrine	48
α_{1a}	Overexpression/heart-specific promoter	↑ Contractile response ↑ Survival ↑ ANF mRNA No hypertrophy ↑ Post-ischemic protection	54
α_{1b}	Gene deletion	Normal resting blood pressure ↓ Blood pressure response to phenylephrine ↓ Vasoconstriction	45
α_{1b}	Overexpression/heart-specific promoter	↑ Phospholipase C activity ↑ ANF mRNA No hypertrophy ↓ Contractile and heart rate response to β -AR	55
CAM α_{1b}	Overexpression/heart-specific promoter	↑ Phospholipase C activity ↑ Hypertrophy ↑ ANF mRNA Normal blood pressure	50
CAM α_{1b}	Overexpression/receptor promoter	↓ Contractile response to β -AR Autonomic failure ↑ Hypertrophy	52
$\alpha_{1a} \alpha_{1b}$	Double gene deletion	<i>In males</i> Normal resting blood pressure ↓ Cardiac growth after birth ↓ Heart rate, ↓ cardiac output ↓ Basal ERK activity ↑ Mortality to pressure overload Contraction abnormalities	53
α_{1d}	Gene deletion	↓ Resting blood pressure ↓ Blood pressure response to phenylephrine ↓ Vasoconstriction	49
$\alpha_{1d} \alpha_{1b}$	Double gene deletion	↓ Resting blood pressure ↓↓ Blood pressure response to phenylephrine ↓↓ Vasoconstriction	57

subtypes contribute to the regulation of blood pressure with a prominent role for the α_{1a} and α_{1d} ; (b) both the α_{1a} and α_{1b} -AR play a role in cardiac pathological hypertrophy (independent from pressure overload) or physiological hypertrophy associated with postnatal growth; and (c) the α_1 -ARs maintain normal heart function as demonstrated by the fact that the double deletion of the α_{1a} and α_{1b} -AR results in some features of failing heart.

Other interesting features of the α_1 -AR subtypes have emerged from studies on the genetically modified mice including their effects on heart contractile function, cardiac rhythm and protection from ischemic injury (56). Additional studies are required to gain a deeper understanding of these complex effects.

Conclusions and perspectives

In the past years, we have gained significant information of some molecular properties and functional implications of the α_1 -AR subtypes both from *in vitro* and *in vivo* studies.

Several studies focused on individual receptor subtypes whereas only few others attempted to compare the behavior of different receptors in similar experimental conditions. This latter approach should be implemented in future studies, both *in vitro* and *in vivo*, to better assess differences and similarities among the three α_1 -AR subtypes.

The elucidation of receptor-mediated signaling events in time and space will depend on a much deeper understanding of the interactions among receptors and signaling molecules which has recently emerged as an important paradigm in the GPCR field. Beyond receptor oligomerization (Table 1), a number of novel proteins have been found to interact with the α_1 -AR subtypes (Table 2), but for most of these interactions the functional implications are elusive. The vast majority of studies on α_1 -AR subtypes have been performed in recombinant systems. A big challenge in the future will be to explore the functional implications of a variety of interactions in different tissues and physiological conditions. The α_1 -AR subtypes are important regulators of several physiological parameters as highlighted by studies in genetically modified mice (Table 3), and further investigation on this receptor system might have new interesting implications in pharmacology and drug development.

Declaration of interest

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References

- Graham RM, Perez DM, Hwa J, Piascik MT. α_1 -adrenergic receptor subtypes. Molecular structure, function, and signaling. *Circ Res* 1996, 78, 737–749.
- Michel MC, Kenny B, Schwinn DA. Classification of α_1 -adrenoceptor subtypes. *Naunyn Schmiedeberg Arch Pharmacol* 1995, 352, 1–10.
- Schwinn DA, Johnston GI, Page SO, Mosley MJ, Wilson KH, Worman NP, Campbell S, Fidock MD, Furness LM, Parry-Smith DJ. Cloning and pharmacological characterization of human α_1 -adrenergic receptors: sequence corrections and direct comparison with other species homologues. *J Pharmacol Exp Ther* 1995, 272, 134–142.
- Cotecchia S, Stanasila L, Diviani D, Björklöf K, Rossier O, Fanelli F. Structural determinants involved in the activation and regulation of G protein-coupled receptors: lessons from the α_1 -adrenergic receptor subtypes. *Biol Cell* 2004, 96, 327–333.
- Cotecchia S, Björklöf K, Rossier O, Stanasila L, Greasley P, Fanelli F. The α_{1b} -adrenergic receptor subtype: molecular properties and physiological implications. *J Recept Signal Transduct Res* 2002, 22, 1–16.
- Costa T, Cotecchia S. Historical review: Negative efficacy and the constitutive activity of G-protein-coupled receptors. *Trends Pharmacol Sci* 2005, 26, 618–624.
- Angers S, Salahpour A, Bouvier M. Dimerization: an emerging concept for G protein-coupled receptor ontogeny and function. *Annu Rev Pharmacol Toxicol* 2002, 42, 409–435.
- Ritter SL, Hall RA. Fine-tuning of GPCR activity by receptor-interacting proteins. *Nat Rev Mol Cell Biol* 2009, 10, 819–830.
- Kjelsberg MA, Cotecchia S, Ostrowski J, Caron MG, Lefkowitz RJ. Constitutive activation of the α_{1B} -adrenergic receptor by all amino acid substitutions at a single site. Evidence for a region which constrains receptor activation. *J Biol Chem* 1992, 267, 1430–1433.
- Rossier O, Abuin L, Fanelli F, Leonardi A, Cotecchia S. Inverse agonism and neutral antagonism at α_{1a} - and α_{1b} -adrenergic receptor subtypes. *Mol Pharmacol* 1999, 56, 858–866.
- Scheer A, Fanelli F, Costa T, De Benedetti PG, Cotecchia S. The activation process of the α_{1B} -adrenergic receptor: potential role of protonation and hydrophobicity of a highly conserved aspartate. *Proc Natl Acad Sci USA* 1997, 94, 808–813.
- Greasley PJ, Fanelli F, Rossier O, Abuin L, Cotecchia S. Mutagenesis and modelling of the α_{1b} -adrenergic receptor highlight the role of the helix 3/helix 6 interface in receptor activation. *Mol Pharmacol* 2002, 61, 1025–1032.
- Theroux TL, Esbenshade TA, Peavy RD, Minneman KP. Coupling efficiencies of human α_1 -adrenergic receptor subtypes: titration of receptor density and responsiveness with inducible and repressible expression vectors. *Mol Pharmacol* 1996, 50, 1376–1387.
- McWhinney C, Wenham D, Kanwal S, Kalman V, Hansen C, Robishaw JD. Constitutively active mutants and the α_{1b} -adrenergic receptor subtypes reveal coupling to different signaling pathways and physiological responses in rat cardiac myocytes. *J Biol Chem* 2000, 275, 2087–2097.
- Hague C, Uberti MA, Chen Z, Hall RA, Minneman KP. Cell surface expression of α_{1D} -adrenergic receptors is controlled by heterodimerization with α_{1B} -adrenergic receptors. *J Biol Chem* 2004, 279, 15541–15549.
- Uberti MA, Hague C, Oller H, Minneman KP, Hall RA. Heterodimerization with β_2 -adrenergic receptors promotes surface expression and functional activity of α_{1D} -adrenergic receptors. *J Pharmacol Exp Ther* 2005, 313, 16–23.
- García-Sáinz JA, Torres-Padilla ME. Modulation of basal intracellular calcium by inverse agonists and phorbol myristate acetate in rat-1 fibroblasts stably expressing α_{1d} -adrenoceptors. *FEBS Lett* 1999, 443, 277–281.
- Gisbert R, Ziani K, Miquel R, Noguera MA, Ivorra MD, Anselmi E, D'Ocon P. Pathological role of a constitutively active population of α_{1D} -adrenoceptors in arteries of spontaneously hypertensive rats. *Br J Pharmacol* 2002, 135, 206–216.

19. Stanasila L, Perez JB, Vogel H, Cotecchia S. Oligomerization of the alpha 1a- and alpha 1b-adrenergic receptor subtypes. Potential implications in receptor internalization. *J Biol Chem* 2003, 278, 40239-40251.
20. Perez DM, DeYoung MB, Graham RM. Coupling of expressed alpha 1B- and alpha 1D-adrenergic receptor to multiple signaling pathways is both G protein and cell type specific. *Mol Pharmacol* 1993, 44, 784-795.
21. Hu ZW, Shi XY, Lin RZ, Hoffman BB. Contrasting signaling pathways of alpha1A- and alpha1B-adrenergic receptor subtype activation of phosphatidylinositol 3-kinase and Ras in transfected NIH3T3 cells. *Mol Endocrinol* 1999, 13, 3-14.
22. Nguyen VA, Gao B. Cross-talk between alpha(1B)-adrenergic receptor (alpha(1B)AR) and interleukin-6 (IL-6) signaling pathways. Activation of alpha(1b)AR inhibits il-6-activated STAT3 in hepatic cells by a p42/44 mitogen-activated protein kinase-dependent mechanism. *J Biol Chem* 1999, 274, 35492-35498.
23. Gonzalez-Cabrera PJ, Shi T, Yun J, McCune DF, Rorabaugh BR, Perez DM. Differential regulation of the cell cycle by alpha1-adrenergic receptor subtypes. *Endocrinology* 2004, 145, 5157-5167.
24. Iwaki K, Sukhatme VP, Shubeita HE, Chien KR. Alpha- and beta-adrenergic stimulation induces distinct patterns of immediate early gene expression in neonatal rat myocardial cells. *fos/jun* expression is associated with sarcomere assembly; *Egr-1* induction is primarily an alpha 1-mediated response. *J Biol Chem* 1990, 265, 13809-13817.
25. Knowlton KU, Michel MC, Itani M, Shubeita HE, Ishihara K, Brown JH, Chien KR. The alpha 1A-adrenergic receptor subtype mediates biochemical, molecular, and morphologic features of cultured myocardial cell hypertrophy. *J Biol Chem* 1993, 268, 15374-15380.
26. Ramirez MT, Sah VP, Zhao XL, Hunter JJ, Chien KR, Brown JH. The MEKK-JNK pathway is stimulated by alpha1-adrenergic receptor and ras activation and is associated with in vitro and in vivo cardiac hypertrophy. *J Biol Chem* 1997, 272, 14057-14061.
27. Sah VP, Hoshijima M, Chien KR, Brown JH. Rho is required for Galphaq and alpha1-adrenergic receptor signaling in cardiomyocytes. Dissociation of Ras and Rho pathways. *J Biol Chem* 1996, 271, 31185-31190.
28. Maruyama Y, Nishida M, Sugimoto Y, Tanabe S, Turner JH, Kozasa T, Wada T, Nagao T, Kurose H. Galpha(12/13) mediates alpha(1)-adrenergic receptor-induced cardiac hypertrophy. *Circ Res* 2002, 91, 961-969.
29. Appert-Collin A, Cotecchia S, Nenniger-Tosato M, Pedrazzini T, Diviani D. The A-kinase anchoring protein (AKAP)-Lbc-signaling complex mediates alpha1 adrenergic receptor-induced cardiomyocyte hypertrophy. *Proc Natl Acad Sci USA* 2007, 104, 10140-10145.
30. Diviani D, Lattion AL, Larbi N, Kunapuli P, Pronin A, Benovic JL, Cotecchia S. Effect of different G protein-coupled receptor kinases on phosphorylation and desensitization of the alpha1B-adrenergic receptor. *J Biol Chem* 1996, 271, 5049-5058.
31. Diviani D, Lattion AL, Cotecchia S. Characterization of the phosphorylation sites involved in G protein-coupled receptor kinase- and protein kinase C-mediated desensitization of the alpha1B-adrenergic receptor. *J Biol Chem* 1997, 272, 28712-28719.
32. Stanasila L, Abuin L, Dey J, Cotecchia S. Different internalization properties of the alpha1a- and alpha1b-adrenergic receptor subtypes: the potential role of receptor interaction with beta-arrestins and AP50. *Mol Pharmacol* 2008, 74, 562-573.
33. Vázquez-Prado J, Medina LC, Romero-Avila MT, González-Espinosa C, García-Sáinz JA. Norepinephrine- and phorbol ester-induced phosphorylation of alpha(1a)-adrenergic receptors. Functional aspects. *J Biol Chem* 2000, 275, 6553-6559.
34. García-Sáinz JA, Rodríguez-Pérez CE, Romero-Avila MT. Human alpha1D-adrenoceptor phosphorylation and desensitization. *Biochem Pharmacol* 2004, 67, 1853-1858.
35. De Wire SM, Seungkirl A, Lefkowitz RJ, Shenoy SK. Beta-arrestins and cell signaling. *Annu Rev Physiol* 2007, 69, 483-510.
36. Pupo AS, Minneman KP. Interaction of neuronal nitric oxide synthase with alpha1-adrenergic receptor subtypes in transfected HEK-293 cells. *BMC Pharmacol* 2002, 2, 17.
37. Xu Q, Xu N, Zhang T, Zhang H, Li Z, Yin F, Lu Z, Han Q, Zhang Y. Mammalian tolloid alters subcellular localization, internalization, and signaling of alpha(1a)-adrenergic receptors. *Mol Pharmacol* 2006, 70, 532-541.
38. Hague C, Bernstein LS, Ramineni S, Chen Z, Minneman KP, Hepler JR. Selective inhibition of alpha1A-adrenergic receptor signaling by RGS2 association with the receptor third intracellular loop. *J Biol Chem* 2005, 280, 27289-27295.
39. Bernstein LS, Ramineni S, Hague C, Cladman W, Chidiac P, Levey AI, Hepler JR. RGS2 binds directly and selectively to the M1 muscarinic acetylcholine receptor third intracellular loop to modulate Gq/11alpha signaling. *J Biol Chem* 2004, 279, 21248-21256.
40. Diviani D, Lattion AL, Abuin L, Staub O, Cotecchia S. The adaptor complex 2 directly interacts with the alpha 1b-adrenergic receptor and plays a role in receptor endocytosis. *J Biol Chem* 2003, 278, 19331-19340.
41. Stanasila L, Abuin L, Diviani D, Cotecchia S. Direct interaction of ezrin with the alpha1b-adrenergic receptor regulates recycling of the internalized receptors. *J Biol Chem* 2006, 281, 4354-4363.
42. Pupo AS, Minneman KP. Specific interactions between gC1qR and alpha1-adrenoceptor subtypes. *J Recept Signal Transduct Res* 2003, 23, 185-195.
43. Wang Q, Zhao J, Brady AE, Feng J, Allen PB, Lefkowitz RJ, Greengard P, Limbird LE. Spinophilin blocks arrestin actions in vitro and in vivo at G protein-coupled receptors. *Science* 2004, 304, 1940-1944.
44. Chen Z, Hague C, Hall RA, Minneman KP. Syntrophins regulate alpha1D-adrenergic receptors through a PDZ domain-mediated interaction. *J Biol Chem* 2006, 281, 12414-12420.
45. Cavalli A, Lattion AL, Hummler E, Nenniger M, Pedrazzini T, Aubert JF, Michel MC, Yang M, Lembo G, Vecchione C, Mostardini M, Schmidt A, Beermann F, Cotecchia S. Decreased blood pressure response in mice deficient of the alpha 1b-adrenergic receptor. *Proc Natl Acad Sci USA* 1997, 94, 11589-11594.
46. Burcelin R, Uldry M, Foretz M, Perrin C, Dacosta A, Nenniger-Tosato M, Seydoux J, Cotecchia S, Thorens B. Impaired glucose homeostasis in mice lacking the alpha1b-adrenergic receptor subtype. *J Biol Chem* 2004, 279, 1108-1115.
47. Auclair A, Drouin C, Cotecchia S, Glowinski J, Tassin JP. 5-HT2A and alpha1b-adrenergic receptors externally mediate dopamine release, locomotor response and behavioural sensitization to opiates and psychostimulants. *Eur J Neurosci* 2004, 20, 3073-3084.
48. Rokosh DG, Simpson PC. Knockout of the alpha 1A/C-adrenergic receptor subtype: the alpha 1A/C is expressed in resistance arteries and is required to maintain arterial blood pressure. *Proc Natl Acad Sci USA* 2002, 99, 9474-9479.
49. Tanoue A, Nasa Y, Koshimizu T, Shinoura H, Oshikawa S, Kawai T, Sunada S, Takeo S, Tsujimoto G. The alpha(1D)-adrenergic receptor directly regulates arterial blood pressure via vasoconstriction. *J Clin Invest* 2002, 109, 765-775.
50. Milano CA, Dolber PC, Rockman HA, Bond RA, Venable ME, Allen LF, Lefkowitz RJ. Myocardial expression of a constitutively active alpha 1B-adrenergic receptor in transgenic mice induces cardiac hypertrophy. *Proc Natl Acad Sci USA* 1994, 91, 10109-10113.
51. Dorn GW 2nd, Brown JH. Gq signaling in cardiac adaptation and maladaptation. *Trends Cardiovasc Med* 1999, 9, 26-34.
52. Zuscik MJ, Chalothorn D, Hellard D, Deighan C, McGee A, Daly CJ, Waugh DJ, Ross SA, Gaivin RJ, Morehead AJ, Thomas JD, Plow EF, McGrath JC, Piascik MT, Perez DM. Hypotension, autonomic failure, and cardiac hypertrophy in transgenic mice overexpressing the alpha 1B-adrenergic receptor. *J Biol Chem* 2001, 276, 13738-13743.
53. O'Connell TD, Ishizaka S, Nakamura A, Swigart PM, Rodrigo MC, Simpson GL, Cotecchia S, Rokosh DG, Grossman W, Foster E, Simpson PC. The alpha(1A/C)- and alpha(1B)-adrenergic receptors are required for physiological cardiac hypertrophy in the double-knockout mouse. *J Clin Invest* 2003, 111, 1783-1791.

54. Lin F, Owens WA, Chen S, Stevens ME, Kesteven S, Arthur JF, Woodcock EA, Feneley MP, Graham RM. Targeted alpha(1A)-adrenergic receptor overexpression induces enhanced cardiac contractility but not hypertrophy. *Circ Res* 2001, 89, 343-350.
55. Akhter SA, Milano CA, Shotwell KF, Cho MC, Rockman HA, Lefkowitz RJ, Koch WJ. Transgenic mice with cardiac overexpression of alpha1B-adrenergic receptors. In vivo alpha1-adrenergic receptor-mediated regulation of beta-adrenergic signaling. *J Biol Chem* 1997, 272, 21253-21259.
56. Woodcock EA. Roles of α 1A and α 1B-adrenoceptors in heart: insights from studies on genetically modified mice. *Clin and Exp Pharmacol and Physiol* 2007 34, 884-888.
57. Hosoda C, Koshimizu TA, Tanoue A, Nasa Y, Oikawa R, Tomabechi T, Fukuda S, Shinoura H, Oshikawa S, Takeo S, Kitamura T, Cotecchia S, Tsujimoto G. Two alpha1-adrenergic receptor subtypes regulating the vasopressor response have differential roles in blood pressure regulation. *Mol Pharmacol* 2005, 67, 912-922.