

# A new low-latitude late Paleocene-early Eocene radiolarian biozonation based on unitary associations: applications for accreted terranes

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**ABSTRACT:** The late Paleocene – early Eocene sequences of DSDP Leg 10 Sites 86, 94, 95, and 96, Leg 43 Site 384 and ODP Leg 171B Hole 1051A have been re-sampled and re-examined for radiolarians. A new late Paleocene to early Eocene low-latitude radiolarian zonation suited for the correlation of accreted terranes is established by using the Unitary Association (UA) method. This method has the property of attributing equal weight to each species occurrence, which has the advantage of not being dependant on a limited set of key datums. Twenty-two UAs have been erected and correlated to the existing age models (given by nannofossils, planktonic foraminifera and radiolarians) for each site. The 22 UAs have been united into seven Unitary Associations Zones (UA Zones) (JP10- JE4) to increase lateral traceability. Herein we present the resulting composite range chart and correlation between the studied cores.

The position of the UA Zones in the Paleogene timescale of Berggren et al. (1995) have been estimated using a general consensus correlation with calcareous microfossil groups and the existing radiolarian zonation. Reproducible radiolarian events identified in the present work are bound to directly tied and compiled absolute ages given by Nigrini et al. (2006) and Sanfilippo and Nigrini (1998a). The RP zones (Sanfilippo and Nigrini 1998a) and the UA Zones are consistent. Unitary Associations permit to distinguish supplementary zonal subdivisions within RP7 and RP6.

Topotypes from DSDP Leg 10 have been illustrated using mainly SEM imaging to facilitate the identification of re-crystallized forms.

## INTRODUCTION

A common problem in biostratigraphic data established from different stratigraphic records is that the sequence of appearances and disappearances of fossil taxa is contradictory from one record to the other. In other words, a direct stratigraphic representation of the correlation lines between these records will show numerous crossovers. This may be due to many different factors such as preservation, incorrect identification and biogeographic constraints. In some cases, diachronous first or last appearances can be estimated when absolute ages for the sections are independently known (Johnson and Nigrini 1985; Moore et al. 1993; Nigrini and Caulet 1992). As a consequence of biogeographically controlled variations in species ranges, various zonations can be found for different realms: Tropics (Sanfilippo and Nigrini 1998a); South Pacific (Hollis 1993, 2002, Hollis et al. 2005); Antarctic (Caulet 1991, Lazarus 1992); Norwegian/Greenland Sea (Goll and Bjorklund 1989); North Pacific (Morley and Nigrini 1995, Shilov 1995); Boreal (Kovlova 1999). Herein we present a new biochronology for the low-latitude Atlantic late Paleocene to early Eocene, constructed by means of Unitary Associations (Guex 1977, 1991). The study is based on all available literature and re-sampled material from DSDP Leg 43 Site 384, (Nishimura 1992), ODP Leg 171B Hole 1051A, (Sanfilippo and Blome 2001) and DSDP Leg 10 Sites 86, 94, 95 and 96 (Foreman 1973 and Sanfilippo and Riedel 1973) (text-fig. 1).

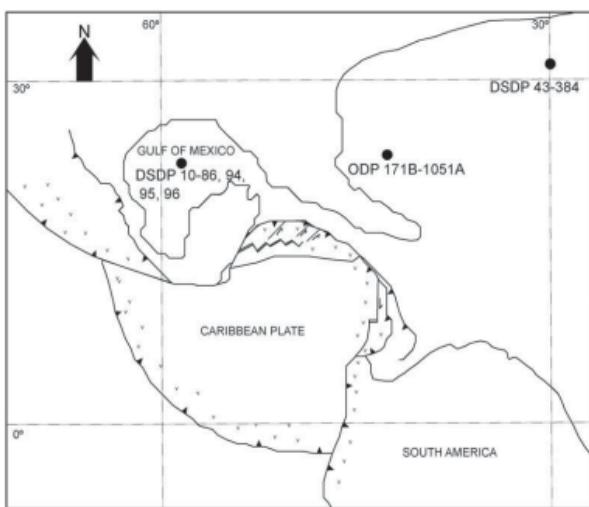
We discuss the biostratigraphic correlation resulting from our new scale with the existing radiolarian (Sanfilippo and Nigrini 1998a), planktonic foraminifera (Berggren et al. 1995) and nannofossil (Bukry 1973, 1975; Okada and Bukry 1980) zonations. The positions of the established UA Zones are estimated in the Paleogene timescale (Berggren et al. 1995).

## Paleogene biochronology and its application in accreted terranes

The tropical Paleocene- early Eocene radiolarian record is discontinuous. Preservation is often variable both overall and can affect individual species differently. Foreman (1973), Riedel and Sanfilippo (1970) and Sanfilippo and Riedel (1973) established a zonation ranging from the latest Paleocene to the early Eocene based on fauna from the Gulf of Mexico (DSDP Leg 4 and DSDP Leg 10). The interval between the Cretaceous/Paleogene boundary and the latest Paleocene zone (*Bekoma bidartensis* Zone (RP7 Sanfilippo and Nigrini 1998a, the lower boundary being defined by the FO of *B. bidartensis*) was referred to as the unzoned interval (Foreman 1973, Sanfilippo et al. 1985). Nishimura (1987, 1992) reduced this interval by describing a new upper Paleocene zone preceding RP7 based on material from DSDP Legs 43 and 93 in the northwest Atlantic (RP6 *Bekoma campechensis* Zone, the lower boundary being defined by the FO of *B. campechensis*). This zone is locally sub-dividable into three sub-units.

Sparse low-latitude (<30°) early Paleocene assemblages have been reported from land sections: California (Foreman 1968), Ecuador (Keller et al. 1997) and Cuba (Florez Abin 1983; Florez Abin and Fernandez 1985). Hollis (1993) proposed a zonation for the early Paleocene based on land sections in New Zealand. Re-examination of these sections in addition to DSDP Sites 208, 277, 1121, resulted in extending this mid-latitude South Pacific zonation into the middle Eocene (Hollis 1997, 2002; Hollis et al. 2005; Strong et al. 1995).

Sanfilippo and Nigrini (1998a) reviewed the tropical zonation, adopted Hollis' zone code system and deduced mean numerical ages for zonal boundary events using the geomagnetic polarity time scale (Cande and Kent 1995) via correlation with calcareous microfossils. The zonation of Sanfilippo and Nigrini (1998a) is



TEXT-FIGURE 1

Paleogeographic reconstruction of the late Paleocene (56 Ma) (simplified from Pindell et al. 2005) showing the location of DSDP and ODP sites sampled for this study (large circles).

constructed using the datum method which is the standard in Cenozoic micropaleontology. The subdivisions are interval zones which are defined as intervals separated by the first occurrence, last occurrence or evolutionary transitions of species.

During our extended work along the convergent margin of Costa Rica we studied Paleocene-Eocene radiolarian cherts and siliceous mudstones. In the field these rocks are often indistinguishable from Mesozoic radiolarites. Radiolarian assemblages are extracted with dilute HF and show preservation characteristics similar to Mesozoic radiolarians that have undergone burial diagenesis. In geologically complex regions where good preservation is biased towards robust taxa, there are difficulties in applying interval zones established in ocean cores with well preserved markers. For example in the Herradura Block, Nicoya Peninsula and Osa-Caño Accretionary Complex (dated as late Paleocene-late Eocene) we rarely find more than the cephalo-thorax of *Bekoma* spp. and consequently, they cannot be used as markers (Bandini et al. this volume; Diserens et al. 2003). In some cases the use of secondary zonal events which are usually based on readily preserved taxa (Sanfilippo and Nigrini 1998a) can overcome this difficulty. The order of these events is although not often regionally reproducible within ocean cores (Sanfilippo and Blome 2001). Given these contradictory occurrences, the consistency of these events as zonal markers needs to be established. The Unitary Association method provides an option for overcoming both of these problems, by 1) giving equal chronologic weight to all considered taxa and 2) establishing maximum co-existences of species (deduced from several sections), that can be used as a reference tool to evaluate the order and reproducibility of various events and their importance as zonal markers (see 2.3). The occurrence data of taxa in six stratigraphic sections is compiled and presented in a composite UA range chart in order to facilitate the correlation of accreted terranes.

## MATERIALS AND METHODS

### Samples

The goal of this study is to produce a low-latitude late Paleocene to early Eocene radiolarian zonation based on Unitary Associa-

tions and to enhance documentation of stratigraphically useful taxa from topotypic material. To do this, we studied all available literature on deep-sea sections in the circum-Caribbean and temperate Atlantic regions. Previously reported preservation, abundance, core recovery, signs of reworking, hiatuses, presences of topotypes and the ages given by calcareous microfossils were taken into consideration. Out of the 195 samples reviewed, 167 samples have been re-studied. Although core recovery of DSDP Leg 10 is relatively poor compared to present day operations, topotype imagery was carried out on 44 samples from Sites 86, 94, 95, 96 (Foreman 1973; Sanfilippo and Riedel 1973). 121 samples were selected from ODP Leg 171B Site 1051A, (partially from previously unstudied intervals) as it contains a well preserved, nearly complete Paleocene –Eocene radiolarian record (Sanfilippo and Blome 2001). Two samples were taken from the well preserved late Paleocene sequence of DSDP 43 Site 384 (Nishimura 1992). Additional data was incorporated from the literature. The full occurrence data set for all samples is available as a supplement to this paper at [Pangaea database at WWDC-Mare].

### Preparation method

Samples were sieved at 40µm following procedures published by Sanfilippo et al. (1985). Two to four strewn slides were made per sample. Residues were then dried and prepared for SEM and transmitted light imaging (Jackett and Baumgartner, in press). SEM imaging of Paleogene topotypes was carried out in order to accurately compare re-crystallized radiolarians from accreted terranes with the literature. Species used for biostratigraphy were recorded as either absent or present. All material is lodged in the Museum of Geology, Lausanne, Switzerland (coll. No. MGL 96,257-96,456).

### Analytic method

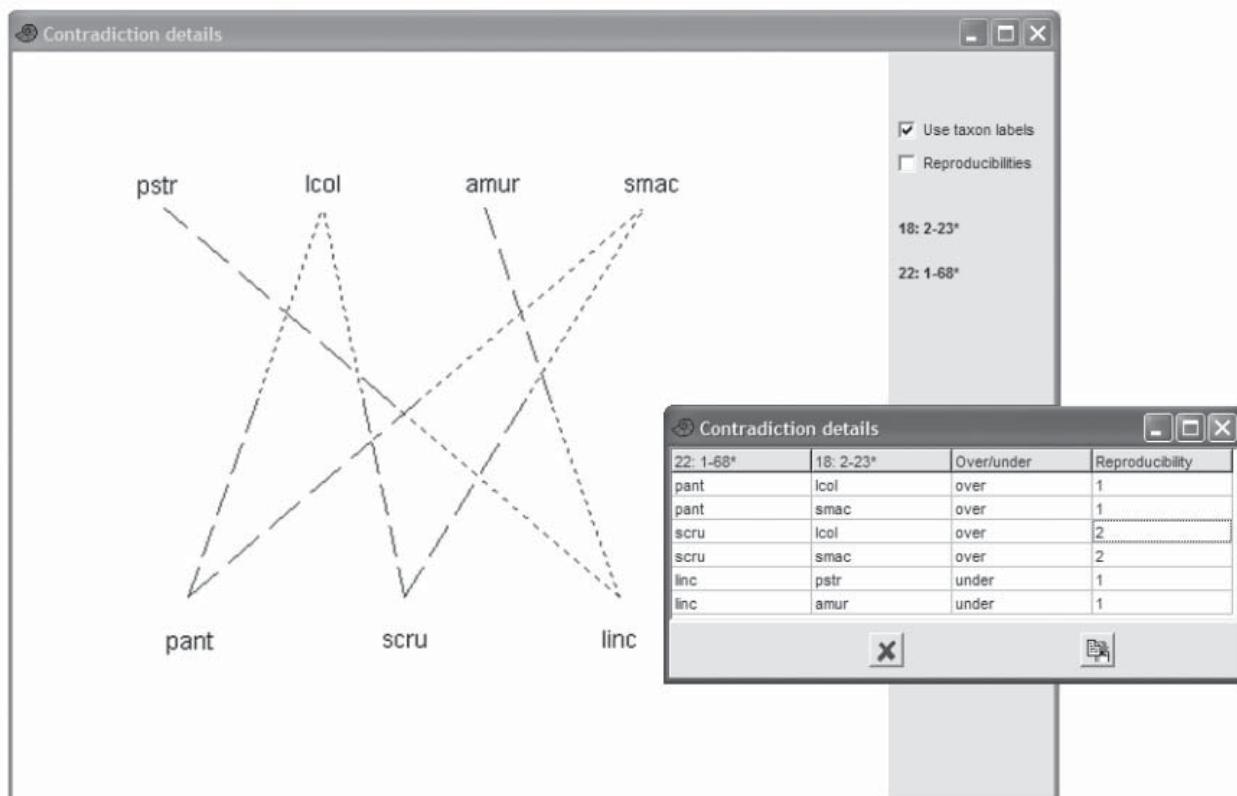
#### General considerations

Over 200 radiolarian species have been identified from 6 stratigraphic sections. Species thought to be long-ranging but only occurring sporadically in exceptionally well-preserved samples, such as *Corythomelissa* and *Saturnalis* were not considered. They cannot be used as markers and their real existence interval is not reflected in raw biostratigraphic data. In a given section a single occurrence of such taxa cannot reasonably be interpreted as a significant distribution and only leads to false superpositional constraints with other taxa, which create unnecessary contradictions. In addition, for the results to be meaningful the taxonomy of each species needs to be uniform. Therefore species with confused concepts have not been included, such as *Pterocodon?* *tenellus* and *Clathrocyclus*. 112 taxa have been retained to construct the zonation.

#### Building unitary associations

Radiolarian biochronology presented in this paper has been established using the Unitary Association (UA) method (Guex 1977, 1991) and the software UGraph (Hammer, Guex and Savary). This method is a deterministic mathematical model designed to construct a discrete sequence of co-existence intervals of species. The program considers two categories of relationships between species:

- 1) Co-existence ('edges', in A terminology): If the ranges of two species overlap in at least one section, they are considered as co-existing in time. Also, the co-existence in time of two species showing no co-occurrence in given sections can be logically as-



TEXT-FIGURE 2

Screen capture from UAgraph showing a contradictory superpositional relationship between maximal cliques 18 and 22 (the concerned species are expressed with their respective codes). The superpositional relationship between species belonging to the respective cliques are shown as differentially dotted lines. The program calculates the relative support for superpositional relationships for taxa pairs in the entire data set. Poorly supported relationships are incrementally converted to co-existences (implied range extensions) until between group superposition conflicts are gone.

sumed if the species display an opposite order of appearance in two or more sections when they are both present. Therefore, co-existence of species in time can be deduced without observed co-occurrences. Such co-existences are said to be 'virtual'.

2) Superpositional relations ('arcs', in UA terminology): If the ranges of two species, A and B, never overlap and if the two species appear systematically in a given order when both present in a section, a superpositional relationship can be established (e.g. A above B). When compiling several sections, collections of mutually co-existing species ('cliques', in UA terminology) can be deduced by examining and arranging a matrix of all species-species co-existences. A clique is maximal if the set of species is not contained in a larger co-existing set.

The UAgraph program (Hammer, Guex and Savary) calculates the relative chronologic constraints of each pair of maximal cliques by considering the existing superpositional relationships between the species of the respective cliques. In certain cases two or more pairs of species indicate contradictory superpositional relationships between two maximal cliques ('inter-clique contradiction'). These contradictions are treated by ignoring the superpositions that are less reproducible in the whole dataset. (e.g. see text-figure 2). A detailed description of the computing process is given in Guex 1991.

Ranges of the species generating superpositional contradictions can be improved by re-examination of the samples. This can lead to an extension of the ranges of some taxa thereby replacing initial

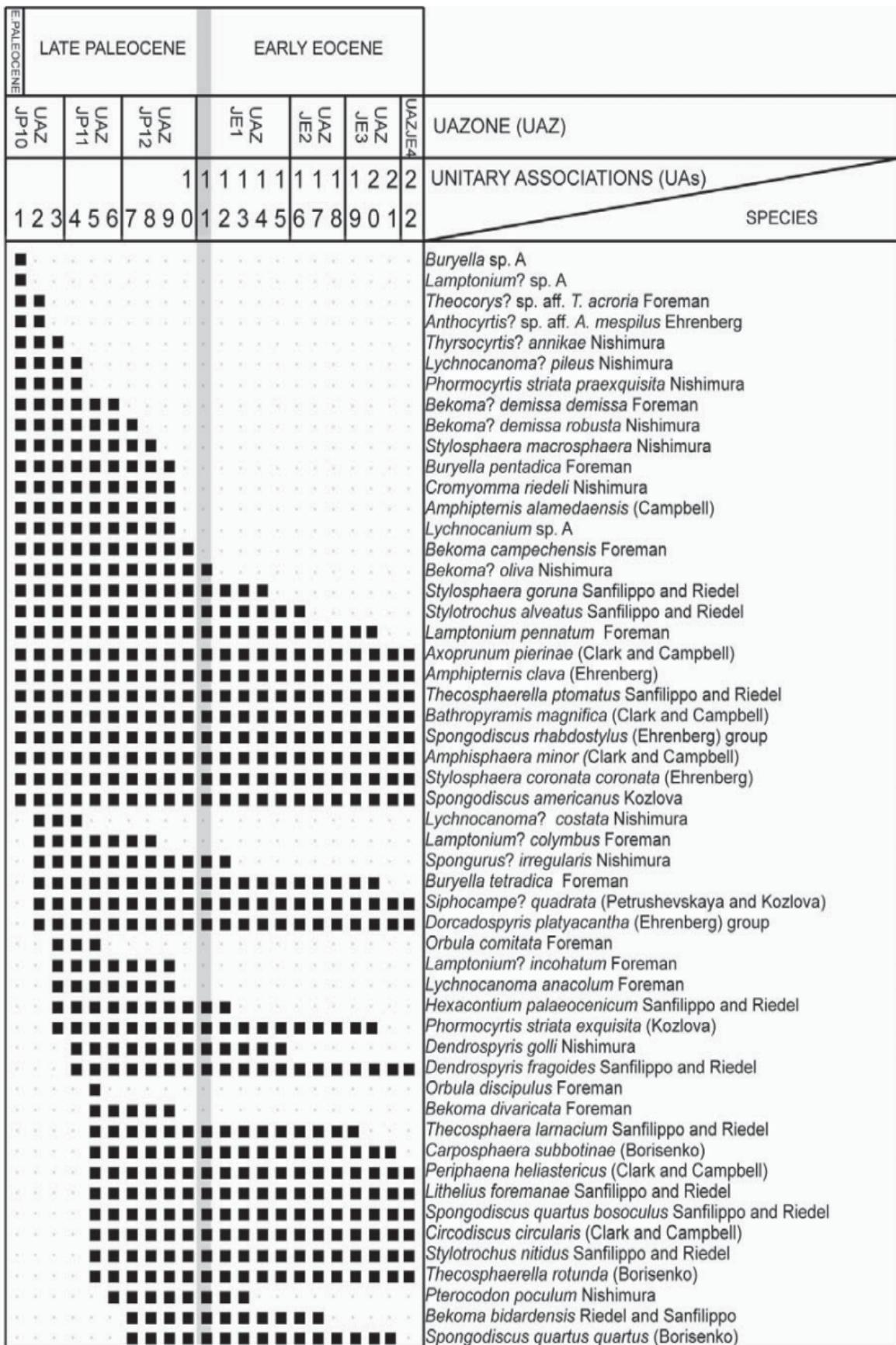
superpositional relationships by real co-existences or it can lead to changing the taxonomic assignment of some taxa. Ideally, it is better to revise the original data, using the information calculated by the program as a guide, than to let the contradictions be removed automatically.

A Unitary Association is a maximal clique defined by the first and last occurrences of the taxa restricted to it. UAs can be united to produce UA Zones with an increased lateral traceability. In our particular problem, constructing UAs allowed us to: 1) date assemblages in accreted terranes where many of the more delicate species within the assemblages are often absent 2) locate critical intervals which are particularly problematic in order to re-examine them, 3) solve contradictions by ignoring less reproducible relationships, which may be due to a poor stratigraphic record, biogeographic constraints or misidentification.

## RESULTS

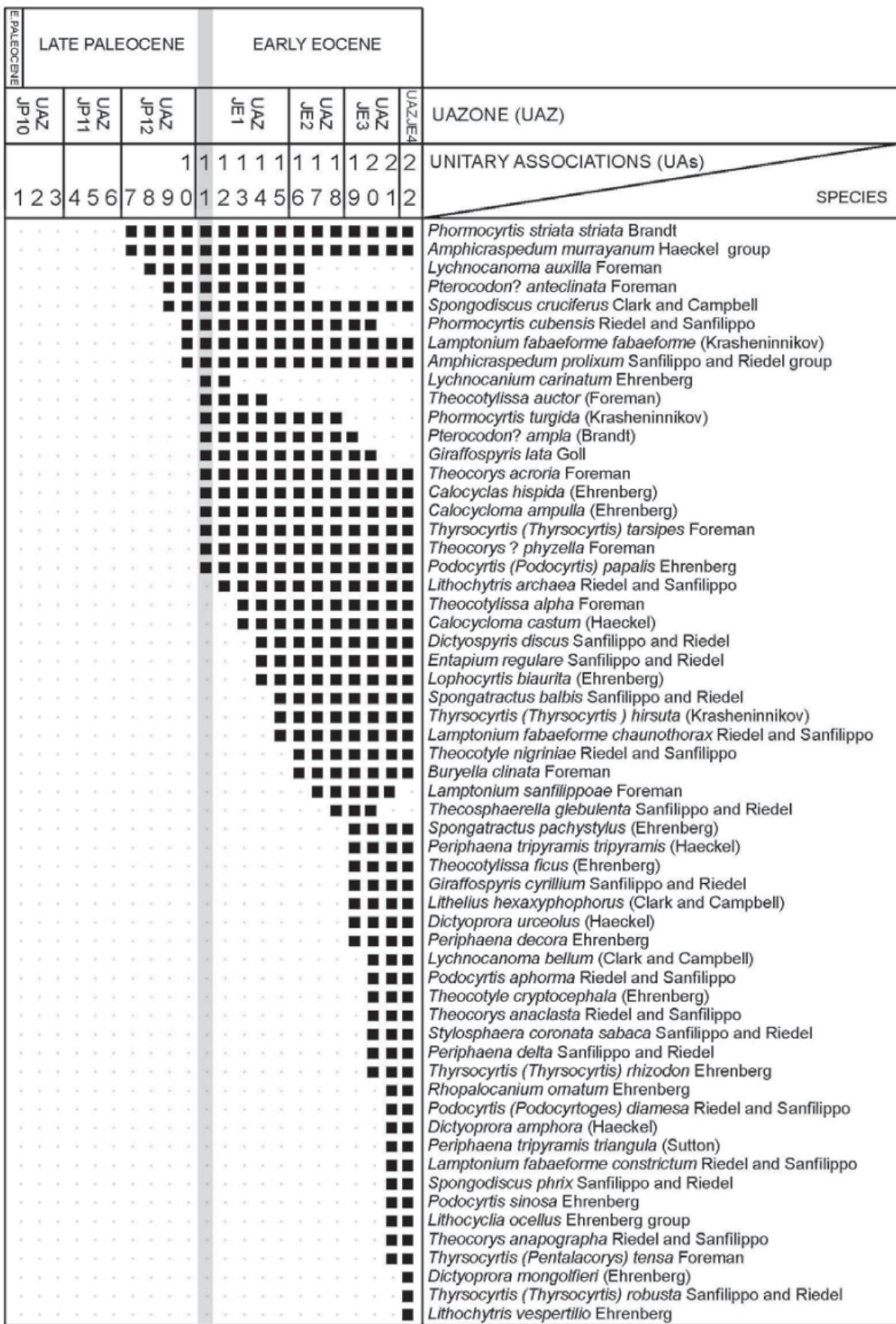
### UA Zone description

The composite range chart representing the succession of the calculated 22 discrete sequences (UAs) of co-existing species is shown in text-figure 3. These have been grouped into seven UA Zones (JP10 to JE4). Zone labels are defined after the initials of the first author followed by the one of the epoch. The UA method produces discrete sequences in which the first or last occurrence of taxa may occur anywhere within the association. As a consequence such a zonation cannot be precisely superimposed on a



### TEXT-FIGURE 3

Composite range chart of taxa versus the identified UAs. Shaded line equals location of the Paleocene/ Eocene boundary within UA1 1.



TEXT-FIGURE 3 (continued)

Composite range chart of taxa versus the identified UAs. Shaded line equals location of the Paleocene/ Eocene boundary within UA1.1.

zonation defined on individual events. Unitary Associations can be strictly identified in certain intervals of some sections. When these intervals have been correlated to existing zonations (Berggren et al. 1995; Bukry 1973, 1975; Okada and Bukry 1980, Sanfilippo and Nigrini 1998a), they provide attachment points and permit a comparison. These relations are individually illustrated in text-figure 4 for each studied core. A synthesis of UA correlations to the Paleogene timescale (Berggren et al. 1995) has been tentatively constructed. In table 1, these results are expressed using UAs as reference. The correlations of the UAs to absolute ages are shown both as a two step process via calcareous microfossils (Sanfilippo and Nigrini 1998a) and using individual radiolarian events directly tied to the paleomagnetic timescale (Nigrini et al. 2006).

Several radiolarian datums appear to be moderately diachronous within the UA framework, such as the FO of *Theocotyle cryptcephala* (FO UA20 (CP11) in Site 1051A and FO UA 22 (CP12) in Site 94). Additional stratigraphically extensive sections would be required to establish the magnitude and relevance of this diachronicity.

#### **UAZ JP 10 (UA1, UA2 and UA3)**

UAZ JP 10 is characterized by the association of *Buryella* sp. A n.sp., *Lamptonium?* sp. A n.sp., *Theocorys?* sp. aff. *T.acroria*, *Anthocyrtis?* sp. aff. *A. mespilus* and *Thrysocyrtis?* *annikae* with *Lychnocanoma?* *costata*, *Lamptonium?* *colymbus*, *Spongurus?* *irregularis*, *Buryella tetradiaca*, *Siphocampe?* *quadrata*, *Dorcadospyris platyacantha* group, *Orbula comitata*, *Lamptonium?* *incohatum*, *Lychnocanoma anacolum*, *Hexacontium palaeocenicum* and *Phormocyrtis striata exquisita*. The lower boundary is presently unknown.

*Samples:* UA1 is found in samples ODP 171B-1051A-73x-2w (23-25cm) to ODP 171B-1051A-65x-1w (52-53cm). UA2 is found in samples ODP 171B-1051A-64x-5w (76-78cm) to ODP 171B-1051A-63x-2w (28-30cm) and DSDP 43-384-11-3 (130-132cm) to DSDP 43-384-10-6 (38-40cm). UA3 is found in samples ODP 171B-1051A-62x-4w (42-44cm) to ODP 171B-1051A-61x-4w (40-42cm) and DSDP 43-384-10-5 (38-40cm) to DSDP 43-384-9-3 (38-40cm).

*Approximate correlation:* CP3-4, P1-4 and RP6.

#### **UAZ JP11 (UA4, UA5 and UA6)**

UAZ JP 11 is characterized by the association of *Lychnocanoma?* *pileus*, *Phormocyrtis striata praexquisita*, *Bekoma?* *demissa*, *demissa*, *Orbula comitata* and *Lychnocanoma?* *costata* with *Dendrospyris golli*, *Dendrospyris fragoides*, *Orbula discipulus*, *Bekoma divaricata*, *Thecosphaera larnacium*, *Carposphaera subbotinae*, *Periphera heliastericus*, *Lithelius foremanae*, *Spongodiscus quartus bosoculus*, *Circodiscus circularis*, *Stylo trochus nitidus*, *Thecosphaerella rotunda* and *Pterocodon poculum*.

*Remarks:* A hiatus of circa 1 Ma occurs between the base of ODP 171B-1051A-58x-CC and ODP 171B-1051A-59x-1w (identified by the absence of nannofossil zone CP6 (Shipboard Scientific Party 1998)), and could explain why UA5 and UA6 are undetermined in this site.

*Samples:* UA4 is found in samples ODP 171B-1051A-61x-2w (33-35cm) to ODP 171B-1051A-59x-4w (50-51cm). UA5 is found in samples DSDP 43-384-9-1 (38-40cm) to DSDP 43-384-8-5 (36-38cm) and DSDP 10-86-8-4 (119-125cm) to DSDP 10-86-8-

1 (114-116cm). UA6 is found in samples 10-86-8-3 (40-42cm) to 10-86-8-1 (80-82cm).

*Approximate correlation:* CP5-6, P4 and RP6.

#### **UAZ JP12 (UA7, UA8, UA9 and UA10)**

UAZ JP 12 is characterized by the association of *Bekoma?* *demissa robusta*, *Stylosphaera macrosphaera*, *Buryella pentadica*, *Cromyomma riedeli*, *Amphipternis alamedaensis*, *Lychnocanium* sp. A, *Bekoma campechensis*, *Lamptonium?* *colymbus*, *Lamptonium?* *incohatum*, *Lychnocanoma anacolum* and *Bekoma divaricata* with *Bekoma bidartensis*, *Spongodiscus quartus quartus*, *Phormocyrtis striata striata*, *Amphicraspedum murrayanum* group, *Lychnocanoma auxilla*, *Pterocodon?* *anteclinata*, *Spongodiscus cruciferus*, *Phormocyrtis cubensis*, *Lamptonium fabaeforme fabaeforme* and *Amphicraspedum prolixum* group.

*Samples:* UA7 is found in sample DSDP 43-384-7-6- (36-38cm) and in sample ODP 171B-1051A-58x-6w (96-97cm). UA8 is found in samples DSDP 43-384-7-4- (38-40cm) to DSDP 43-384-7-3- (36-38cm). UA9 is found in samples DSDP 43-384-7-2- (38-40cm) to DSDP 43-384-6-1- (38-40cm) and ODP 171B-1051A-58-x2w (34-36cm) to ODP 171B-1501A-57x-4w (96-98cm). UA10 is found in samples ODP 171B-1051A-57x-3w (38-40cm) to ODP 171B-1051A-56x-5w (37-38cm).

*Approximate correlation:* CP6-8b, P4-5, RP6-7

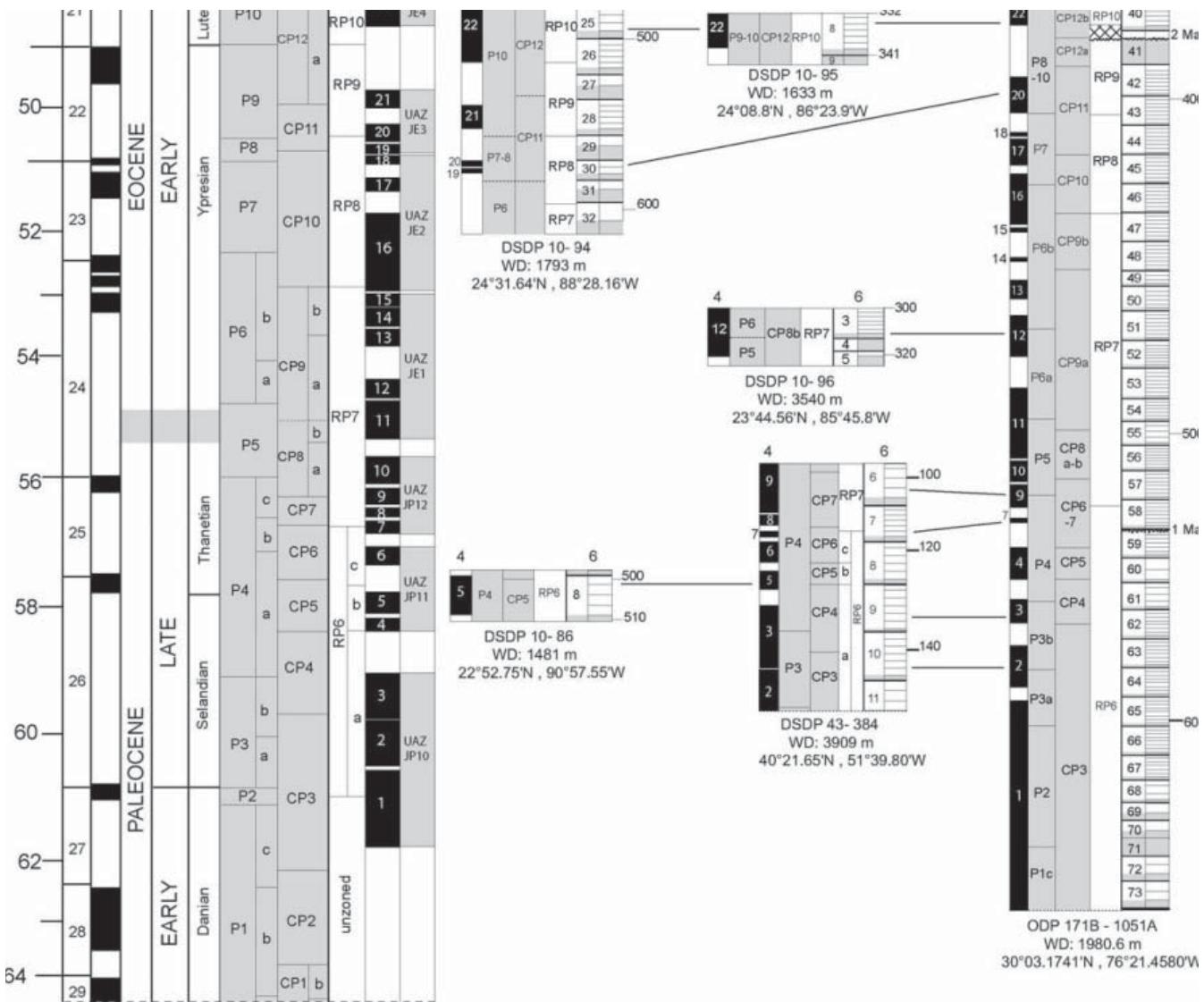
#### **UAZ JE1 (UA11, UA12, UA13, UA14 and UA15)**

UAZ JE1 is characterized by the association of *Bekoma?* *oliva*, *Stylosphaera goruna*, *Spongurus?* *irregularis*, *Hexacontium?* *palaeocenicum*, *Dendrospyris golli* and *Pterocodon poculum* with *Lychnocanium carinatum*, *Theocotylissa auctor*, *Phormocyrtis turgida*, *Pterocodon?* *ampla*, *Girafospyris lata*, *Theocorys acroria*, *Calocyclus hispida*, *Calocyclus ampulla*, *Thrysocyrtis* (*Thrysocyrtis*) *tarsipes*, *Theocorys?* *phyzella*, *Podocyrtis* (*Podocyrtis*) *papalis*, *Lithochytris archaea*, *Theocotylissa alpha*, *Calocyclus castum*, *Dictyospyris discus*, *Entapium regulare*, *Lophocyrtis biaurita*, *Spongatractus balbis*, *Thrysocyrtis* (*Thrysocyrtis*) *hirsuta* and *Lamptonium fabaeforme chaunothorax*.

*Remarks:* The base of the carbon isotope excursion and the benthic foraminifera extinction have been recorded in core ODP 171B-1051A-55x (at 512.90 mbsf and 512.80 mbsf, respectively) (Norris and Rohl 1999). These events are placed at 55.5Ma (Berggren et al. 1995) and occur with UA11 herein. Determination of nannofossil biostratigraphic datums in this interval was hampered by poor preservation (Shipboard Scientific Party 1998). The Paleocene/ Eocene boundary lies between the record of Sites 96 (nannofossil zone CP8b) and 384 (nannofossil zone CP8a).

*Samples:* UA1 1 is found in samples ODP 171B-1051A-56x-4w (37-38cm) to ODP 171B-1051A-53x-5w (34-36cm). UA12 is found in samples ODP 171B-1051A-52x-5w (42-44cm) to ODP 171B-1051A-51x-3w (56-58cm) and DSDP 10-96-5-1 (14-116cm) to DSDP 10-96-3-1 (41-43cm). UA13 is found in samples ODP 171B-1051A-50x-3w (56-58cm) to ODP 171B-1051A-49x-3w (14-16cm). UA14 is found in sample ODP 171B-1051A-48x-5w (33-35cm). UA15 is found in sample ODP 171B-1051A-47x-5w (45-47cm).

*Approximate correlation:* CP8a-9b, P5-6b, RP7



TEXT-FIGURE 4

Correlation of cores based on UAs and an estimate of UAs and UA Zones in the Paleogene timescale (Berggren et al. 1995). 1: Planktonic foraminifera (Berggren et al. 1995), 2: Calcareous Nannofossils (Bukry 1973, 1975; Okada and Bukry 1980), 3: Radiolaria (Sanfilippo and Nigrini 1998a), 4: Unitary Associations (this study), 5: estimate position of UAs and UA Zones in the Paleogene timescale, 6: cores and cores sections, shaded areas represent poor core recovery. Nannofossil data from: DSDP 10 Bukry 1973, DSDP 43 Berggren et al. 2000 and ODP 171B Shipboard Scientific Party zonation assignment 1998. Foraminifera data from: DSDP 10 Shipboard Scientific Party, DSDP 43 Berggren et al. 2000 and ODP 171B Shipboard Scientific Party zonation assignment 1998. Radiolarian RP zone data from: DSDP 10 Foreman 1973, Sanfilippo and Riedel 1973, DSDP 43 Nishimura 1992, Sanfilippo and Nigrini 1998b, ODP 171B Sanfilippo and Blome 2001. Right hand side of cores is mbsf. Wavy lines in core sections represent previously reported hiatuses. Dashed lines within existing zonations indicate a tentative boundary placement. Correlation lines link common occurring UAs.

#### UAZ JE2 (UA16, UA17 and UA18)

UAZ JE2 is characterized by the association of *Stylotrochus alveatus*, *Bekoma bidartensis*, *Lychnocanoma auxilla*, *Pterocodon? anteclinalata* and *Phormocyrtis turgida* with *Theocotyle nigriniae*, *Buryella clinata*, *Lamptonium sanfilippoae* and *Thecosphaerella glebulenta*.

**Samples:** UA16 is found in samples ODP 171B-1051A-47x-4w (56-58cm) to ODP 171B-1051A-45x-5w (84-86cm). UA17 is found in samples ODP 171B-1051A-45x-3w (26-28cm) to ODP 171B-1051A-44x-5w (48-50cm). UA18 is found in sample ODP 171B-1051A-44x-3w (40-42cm).

Approximate correlation: CP9b-1, P6b-7, RP7-8

#### UAZ JE3 (UA19, UA20 and UA21)

UAZ JE3 is characterized by the association of *Lamptonium pennatum*, *Buryella tetradica*, *Phormocyrtis striata exquisita*, *Thecosphaera larnacium*, *Carpospaera subbotinae*, *Spongodiscus quartus quartus*, *Phormocyrtis cubensis*, *Pterocodon? ampla*, *Girafospyris lata*, *Lamptonium sanfilippoae* and *Thecosphaerella glebulenta* with *Spongatractus pachystylus*, *Peripheraena tripyramis tripyramis*, *Theocotylissa ficus*, *Girafospyris cyrillium*, *Theocotyle cryptocephala*, *Lithelius hexaxyphophorus*, *Dictyoprrora urceolus*, *Peripheraena decora*, *Lychnocanoma bellum*, *Podocyrtis aphorma*, *Theocotyle cryptocephala*, *Theocorys*

UAZ UA (this study)	Nannofossils		Consensus correlation		Planktonic foraminifera		Consensus correlation		Radiolarians		Consensus correlation		Absolute ages (Ma)	
	ODP 199	Compiled												
UAZ JE4	UA22	CP12b (1051A), CP12 (95), CP12 (1051A)	CP12	CP12	P8-10 (1051A), P9-10 (95), P10 (94)	P8-10	P8-10	RP10-11 (1051A), RP10 (95), RP10-11 (94)	RP10-11	RP10-11 (1051A)	RP10-11	RP10-11	48.13- 49.91	-49.00
	UA21	CP11 (94)	CP11		P10 (94)	P10		RP9 (94)	RP9					
	UA20	CP11 (94), CP11 (1051A)	CP11		P7-8 (94), P8-10 (1051A)	P7-10		RP8 (94), RP9 (1051A)	RP8-9				~50.30	
UAZ JE3	UA19	CP11 (94)	CP11	CP11	P7-8 (94)	P7-8		RP8 (94)	RP8				53.07- 51.37	
	UA18	CP11 (1051A)	CP11		P7(1051A)	P7	P6b-7	RP8 (1051A)	RP8					
	UA17	CP10-11 (1051A)	CP10-11		P7 (1051A)	P7		RP8 (1051A)	RP8				~52.85	
UAZ JE2	UA16	CP9b-10 (1051A)	CP9b-10	CP9b-11	P6b-7 (1051A)	P6b-7		RP7-8 (1051A)	RP7-8				53.07- 51.37	
	UA15	CP9b (1051A)	CP9b	CP8a- CP9b	P6b (1051A)	P6b	P5-P6b	RP7 (1051A)	RP7					
	UA14	CP9b (1051A)	CP9b		P6b (1051A)	P6b		RP7 (1051A)	RP7					
UAZ JE1	UA13	CP9a (1051A)	CP9a		P6b (1051A)	P6b		RP7 (1051A)	RP7					
	UA12	CP9a (1051A) CP8b (96)	CP8b-9a		P6a-b (1051A) P5-6 (96)	P5-6		RP7 (1051A) RP7 (96)	RP7					
	UA11	CP8a-b-9a (1051A)	CP8a-9a		P6b (1051A)	P6b		RP7 (1051A)	RP7					
UAZ JP12	UA10	CP8a-b (1051A)	CP8a-b	CP6-8b	P5-6 (1051A)	P5-6	P4-5	RP7 (1051A)	RP7					
	UA9	CP6-7 (1051A), CP7-8a (384)	CP6-8a		P4-5 (1051A), P4 (384)	P4-5		RP7 (1051A), RP7 (384)	RP7					
	UA8	CP7 (384)	CP7		P4 (384)	P4		RP7 (384)	RP7				~56.9	
	UA7	CP6-7 (1051A), CP6 (384)	CP6-7		P4 (1051A), P4 (384)	P4		RP6 (1051A), RP7 (384)	RP6-7				53.07- 51.37	
UAZ JP11	UA6	CP6 (384)	CP6	CP5-6	P4 (384)	P4	P4	RP6c (384)	RP6					
	UA5	CP5 (384), CP5 (86)	CP5		P4 (384), P4 (86)	P4		RP6b (384), RP6 (86)	RP6					
	UA4	CP5 (1051A)	CP5		P4 (1051A)	P4		RP6 (1051A)	RP6					
UAZ JP10	UA3	CP4 (1051A), CP3-4 (384)	CP3-4	CP3-4	P3 (1051A), P3-4 (384)	P3-4	P1-4	RP6 (1051A), RP6a (384)	RP6					
	UA2	CP3 (1051A), CP3 (384)	CP3		P3 (1051A), P3 (384)	P3		RP6 (1051A), RP6a (384)	RP6					
	UA1	CP3 (1051A)	CP3		P1-3 (1051A)	P1-3		RP6 (1051A)	RP6					

TABLE 1

Cross-correlation between UA results and existing zonations in study sections. Nannofossils from Bukry 1973, 1975; Okada and Bukry 1980; Planktonic Foraminifera from Berggren et al. 1995 and Berggren et al. 2000; Radiolarian zones from Foreman 1973, Sanfilippo and Riedel 1973 and Sanfilippo and Blome 2001; Absolute ages for Paleogene radiolarian stratigraphic datums from ODP 199 (datums used: FO of *Phormocyrtis striata striata*, *Buryella clinata*, *Theocorytis ficus* and *Lithochytris vespertilio*) from Nigrini et al. 2006 and compiled absolute ages from Sanfilippo and Nigrini 1998a. Note the diachronous occurrence of *Phormocyrtis striata striata* between the Atlantic and Pacific Oceans.

*anaclasta*, *Stylospheara coronata sabaca*, *Peripheraena delta*, *Thrysocyrtis (Thrysocyrtis) rhizodon*, *Rhopalocanium ornatum*, *Podocyrtis (Podocyrtoges) diamesa*, *Dictyoprora amphora*, *Peripheraena tripyramis triangula*, *Lamptonium fabaeforme constrictum*, *Spongodiscus phrix*, *Podocyrtis sinosa*, *Lithocyclia ocellus* group, *Theocorys anapographa* and *Thrysocyrtis (Penatalacorys) tensa*.

**Remarks:** Calcareous nannofossils indicate a hiatus of circa 2 Ma between cores 171B-1051A-40X-CC, and 171B-1051A-41X-CC (Shipboard Scientific Party 1998). Sanfilippo and Blome (2001) confirmed this hiatus with radiolarians stating that most of RP9 and RP10 are missing. Herein UA21 is missing from this core, but it has been identified in DSDP 10-94. Detecting this hiatus in DSDP 10-94 is hampered by large coring gaps in core 27 through to core 29. For further discussion on the correlation and significance of late early Eocene hiatuses see Aubry (1995), Norris et al. (2001) and Sanfilippo and Blome (2001).

**Samples:** UA19 is found in sample DSDP 10-94-30-2 (114-116cm). UA20 is found in sample DSDP 10-94-30-1 (34-35cm) and ODP 171B-1051A-43x-5w (34-36cm) to ODP 171B-1051A-42x-3w (116-118cm). UA21 is found in samples DSDP 10-94-28-5 (43-45cm) to DSDP 10-94-28-2 (42-44cm).

**Approximate correlation:** CP1 1, P7-10, RP8-9

UAZ JE4 (UA22)

UAZ JE4 is characterized by the association of *Dictyoprora monogolfieri*, *Thrysocyrtis (Thrysocyrtis) robusta* and *Lithochytris vespertilio*.

**Samples:** UA22 is found in samples DSDP 10-95-8-6 (43-45cm) to DSDP 10-95-8-1 (42-46cm) and DSDP 10-94-26-4 (43 -45cm) to DSDP 10-94-24-3 (44-46cm) and ODP 171B-1051A-40x-5w (76-78cm) to ODP 171B-1051A-40x-3w (50-52cm).

**Approximate correlation:** CP12, P8-10, RP 10-11

## SUMMARY AND CONCLUSION

The aim of this study is to establish a new late Paleocene to early Eocene low-latitude radiolarian zonation suited for the correlation of accreted terranes. Biostratigraphic data from six cores drilled in the Atlantic Ocean and in the Gulf of Mexico has been compiled and problematic intervals re-examined. Out of the 195 samples considered, 167 samples have been re-studied. Topotypes from DSDP 10 have been imaged by SEM to facilitate the identification of re-crystallized forms found in accreted terranes.

The Unitary Association method has been chosen to carry out this work as it has the property of attributing equal weight to each species occurrence. This approach has the advantage of not being dependant on a limited set of key datums. It is based on a deterministic mathematical model designed to construct a discrete sequence of co-existence intervals of species. Twenty two Unitary Associations have been erected and correlated to the existing age models (given by nannofossils, planktonic foraminifera and radiolarians) for each site. The 22 UAs were united into seven UA Zones (JP10- JE4) to increase lateral traceability. A general consensus correlation with calcareous microfossil groups has been made and used to estimate the position of the UA Zones on the Paleogene timescale. The RP zones and the UA Zones display

good correlation. Unitary Associations permit to distinguish supplementary zonal subdivisions within RP7 and RP6.

## SYSTEMATIC PALEONTOLOGY

Taxonomic notes are given for new species and when taxonomic clarification is required. For other taxa only reference to the author, the first illustration, new combination (if applicable), a paper defining the present species concept and the species code used in this study are given. Taxa are listed in alphabetical order. Species without codes were not considered in the biostratigraphy.

### *Amphicraspedum murrayanum* Haeckel group

Plate 4, figures 1, 2

*Amphicraspedum murrayanum* HAECKEL group 1887, p. 523, pl. 44, fig. 10. – HOLLIS 2006, pl. 1, fig. 1

*Distinguishing characters:* Forms assigned to this species-group include variable morphotypes having an inflated middle part of the test which partially belong to *Amphicraspedum murrayanum*. Distinguished from *A. prolixum* group by its generally shorter test which is more inflated in its median part.

Species code: **amur**

*Amphicraspedum prolixum* Sanfilippo and Riedel  
Plate 4, figures 3-5

*Amphicraspedum prolixum* SANFILIPPO and RIEDEL group 1973, p. 524, pl. 10, figs. 7-11; pl. 28, figs. 3, 4

Species code: **apro**

*Amphipternis alamedaensis* (Campbell and Clark)  
Plate 3, figure 22

*Phormocampe (Cyrtocorys) alamedaensis* CAMPBELL and CLARK 1944, p. 37, pl. 7, fig. 41. – HOLLIS 1997, p. 67, pl. 15, figs. 15, 16

Species code: **asti**

*Amphipternis clava* (Ehrenberg)  
Plate 3, figures 22, 21

*Lithocampe? clava* EHRENBERG 1873, p. 238; 1875, p. 76, pl. 4, fig. 2. – FOREMAN 1973, p. 430, pl. 7, figs. 16, 17; pl. 9, figs. 2, 7

Species code: **acla**

*Amphisphaera minor* (Clark and Campbell)  
Plate 3, figure 13

*Stylosphaera minor* CLARK and CAMPBELL 1942, p. 27, pl. 5, figs. 1, 2, 12; Sanfilippo and Riedel 1973, p. 486, pl. 1, figs. 1-5; pl. 22, fig. 4

Species code: **amin**

*Anthocyrtis mespilus* Ehrenberg  
Plate 1, figure 5

*Anthocyrtis mespilus* EHRENBERG 1854, pl. 36, fig. 13; 1875, p. 66, pl. 6, fig. 4.

*Anthocyrtis?* sp. aff. *A. mespilus* Ehrenberg group. – NISHIMURA 1992, p. 331, pl. 6, figs. 12- 13

Species code: **ames**

*Axoprunum pierinae* (Clark and Campbell)  
Plate 3, figure 14

*Lithatractus pierinae* CLARK and CAMPBELL 1942, p. 34, pl. 5, fig. 25. – HOLLIS 2002, p. 291, pl. 1, fig. 10

Species code: **apie**

*Bathopyramis magnifica* (Clark and Campbell)

Plate 1, figure 16

*Sethopyramis (Cephalopyramis) magnifica* CLARK and CAMPBELL 1942, p. 72, pl. 8, figs. 1, 5, 9. – HOLLIS et al. 1997, p. 57, pl. 6, fig. 22

Species code: **bmag**

*Bekoma bidartensis* Riedel and Sanfilippo

Plate 1, figure 12

*Bekoma bidartensis* RIEDEL and SANFILIPPO 1971, p. 1592, pl. 7, figs. 1-7

Species code: **bbid**

*Bekoma campechensis* Foreman

Plate 1, figure 11

*Bekoma campechensis* FOREMAN 1973, p. 432, pl. 3, fig. 24; pl. 10, figs. 1, 2

Species code: **bcam**

*Bekoma? demissa demissa* Foreman

Plate 1, figure 7

*Bekoma? demissa demissa* FOREMAN 1973, p. 432, pl. 3, fig. 22; pl. 10, fig. 5

Species code: **bdem**

*Bekoma? demissa robusta* Nishimura

Plate 1, figure 6

*Bekoma? demissa robusta* NISHIMURA 1992, p. 333, pl. 6, figs. 2, 3

Species code: **brob**

*Bekoma divaricata* Foreman

*Bekoma divaricata* FOREMAN 1973, p. 433, pl. 3, fig. 23; pl. 10, figs. 3, 4

Species code: **bdiv**

*Bekoma? oliva* Nishimura

Plate 2, figure 4

*Bekoma? oliva* NISHIMURA 1992, p. 333, pl. 5, figs. 10, 11; pl. 13, fig. 6

Species code: **boli**

*Buryella* sp. A

Plate 1, figures 21-23

*Description:* Shell of 5 segments with the lumbar stricture externally expressed. Cephalis subspherical with few circular pores. Collar stricture not externally expressed but with a vertical pore. Third and fourth segments are of comparable width, both conical with one to three rows of circular pores arranged quincuncially. Few unarranged pores are present. The test displays maximum width on the fourth segment and gradually narrows towards the aperture. Fifth segment inverted with 3 rows of circular pores.

Distinguishing characters: This species differs from *Buryella tetradica* by having five segments and by the presence of few quincuncially arranged pores. It differs from the sub-species *B. tetradica* var. A described by Sanfilippo and Blome (2001, p. 210, figs. 8d, 8e) by the third and fourth segments displaying similar proportions and only two to five rows of pores arranged quincuncially, not in longitudinal rows. It differs from *Buryella pentadica* by having an externally expressed lumbar stricture and by the third and fourth segments having similar proportions.

Occurrence: Late early Paleocene to late Paleocene of ODP 171B-1051A

Species code: **bbru**

*Buryella clinata* Foreman

Plate 2, figure 17

*Buryella clinata* FOREMAN 1973, p. 433, pl. 8, figs. 1-3; pl. 9, fig. 19

Species code: **bcli**

*Buryella pentadica* Foreman

Plate 1, figures 14, 15

*Buryella pentadica* FOREMAN 1973, p. 433, pl. 8, fig. 8; pl. 9, figs. 15, 16

Species code: **bpen**

*Buryella tetradica* Foreman

Plate 2, figure 20

*Buryella tetradica* FOREMAN s.s. 1973, p. 433, pl. 8, figs. 4-5; pl. 9, figs. 13, 14

Distinguishing characters: A sub-species of *Buryella tetradica* [Pl. 2, fig. 21], tabulated as *B. tetradica* var. A by Sanfilippo and Blome (2001, p. 210, figs. 8d, 8e), differs from *B. tetradica* s.s. by the fourth segment being transversely segmented. This sub-species has herein not been included in the biostratigraphy.

Species code: **btet**

*Calocyclus hispida* (Ehrenberg)

Plate 2, figure 1

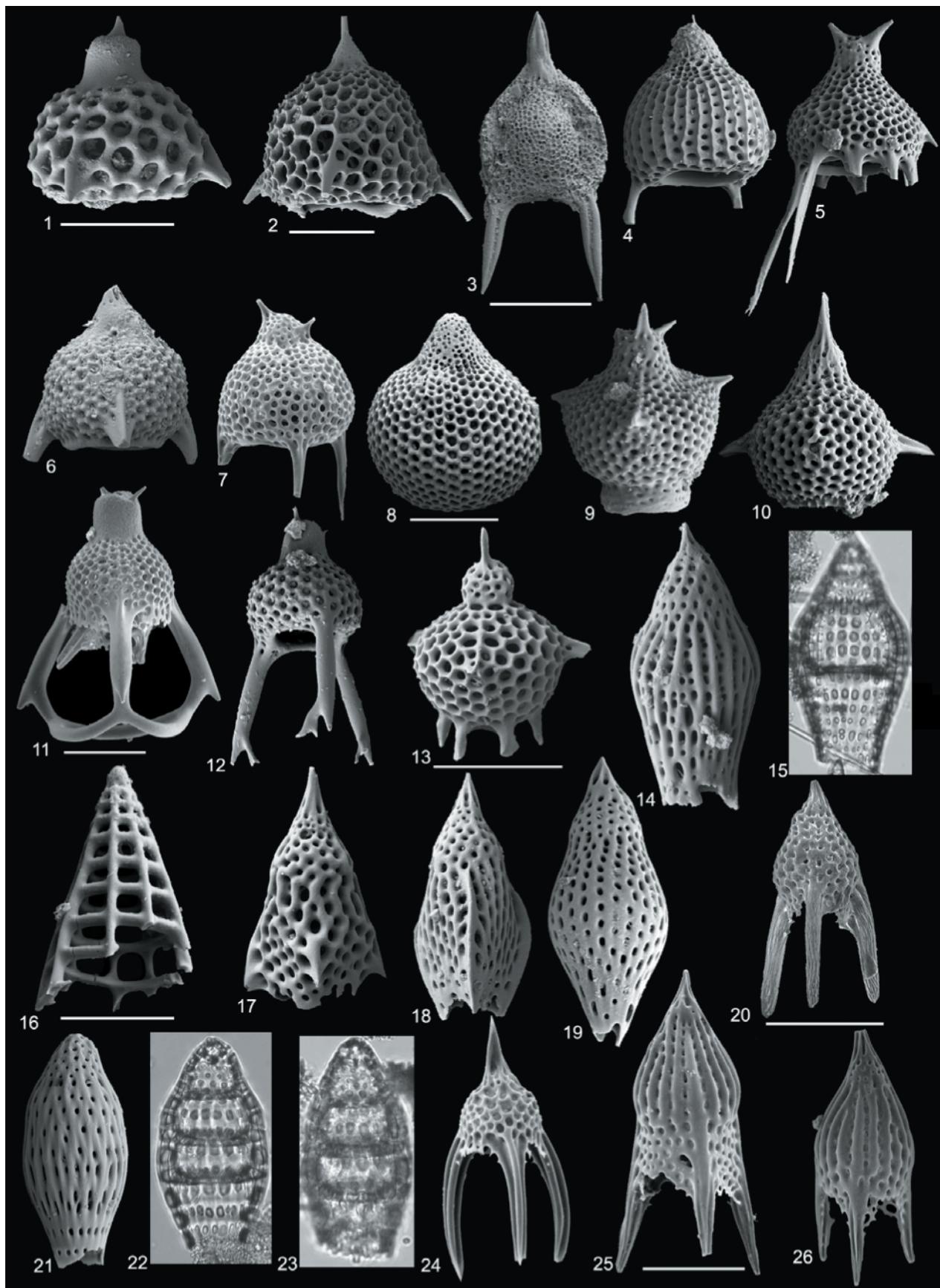
*Anthocyrtis hispida* EHRENBERG 1873 p. 216, pl. 8, fig. 2; 1875, pl. 8, fig. 2. – FOREMAN 1973, p. 434, pl. 1, figs. 12-15; pl. 9, fig. 18

Remarks: Includes forms tabulated by Sanfilippo and Blome

## PLATE 1

All scale bars equal 100µm.

- 1 *Orbula comitata* Foreman. ODP 171B-1051A-61x-4w (40-42cm)
- 2 *Orbula discipulus* Foreman. DSDP 10-86-8-4 (119-125cm)
- 3 *Lychnocanoma? pileus* Nishimura. ODP 171B-1051A-68x-1w (60-62cm)
- 4 *Lychnocanoma? costata* Nishimura. ODP 171B-61x-4w (40-42cm)
- 5 *Anthocyrtis?* sp. aff. *A. mespilus* Ehrenberg. ODP 171B-1051A-67x-1w (34-36cm)
- 6 *Bekoma? demissa robusta* Nishimura. ODP 171B-1051A-68x-1w (60-62cm)
- 7 *Bekoma? demissa demissa* Foreman. DSDP 10-86-8-4 (119-125cm)
- 8 *Lamptonium? columbus* Foreman. ODP 171B-1051A-58x-7w (16-18cm)
- 9 *Lamptonium?* sp. A n.sp. ODP 171B-1051A-67x-2w (40-42cm)
- 10 *Lamptonium pennatum* Foreman. ODP 171B-1051A-58x-7w (16-18cm)
- 11 *Bekoma campechensis* Foreman. ODP 171B-1051A-58x-2w (36-38cm)
- 12 *Bekoma bidartensis* Riedel and Sanfilippo. ODP 171B-1051A-56x-3w (76-78cm)
- 13 *Pterocodon poculum* Nishimura. DSDP 43-384-6-3 (102-109cm)
- 14 *Buryella pentadica* Foreman. ODP 171B-1051A-67x-2w (40-42cm)
- 15 *Buryella pentadica* Foreman. ODP 171B-1051A-67x-1w (34-36cm)
- 16 *Bathropyramis magnifica* (Clark and Campbell). ODP 171B-1051A-67x-1w (34-36cm)
- 17 *Phormocyrtis striata praexquisita* Nishimura. ODP 171B-1051A-67x-1w (34-36cm)
- 18 *Phormocyrtis striata exquisita* (Kozlova). ODP 171B-1051A-58x-7w (16-18cm)
- 19 *Phormocyrtis striata striata* Brandt. ODP 171B-1051A-44x-7w (8-10cm)
- 20 *Lychnocanoma auxilla* Foreman. DSDP 10-96-3-5 (42-44cm)
- 21 *Buryella* sp. A n. sp. ODP 171B-1051A-67x-2w (40-42cm)
- 22 *Buryella* sp. A n. sp. ODP 171B-1051A-67x-1w (34-36cm)
- 23 *Buryella* sp. A n. sp. ODP 171B-1051A-67x-5w (40-42cm)
- 24 *Lychnocanoma anacolum* Foreman. DSDP 10-86-8-4 (119-125cm)
- 25 *Lychnocanum carinatum* Ehrenberg. DSDP 10-96-3-5 (42-44 cm)
- 26 *Lychnocanum* sp. A. ODP 171B-1051A-58x-2w (34-36cm)



(2001, p. 211, fig. 8m) as a sub-species (*Calocyclus hispida* var. A.)

Species code: **chis**

*Calocyclus ampulla* (Ehrenberg)  
Plate 2, figure 2

*Eucyrtidium ampulla* EHRENBURG 1854, pl. 36, figs. 15a-c. – EHRENBURG 1873, p. 225. – EHRENBURG 1875, p. 70, pl. 10, figs. 11, 12; FOREMAN 1973, p. 434, pl. 1, figs. 1-5

Species code: **camp**

*Calocyclus castum* (Haeckel)  
Plate 2, figure 3

*Calocyclus casta* HAECKEL 1887, p. 1384, pl. 73, fig. 10. – FOREMAN 1973, p. 434, pl. 1, figs. 9, 10

Species code: **ccas**

*Carposphaera subbotinae* (Borisenko)  
Plate 3, figure 4

*Cenosphaera subbotinae* BORISENKO 1958, p. 85, pl. 5, figs. 5-7.  
– ABELMANN 1990, p. 691, pl. 1, fig. 9

Species code: **csub**

*Circodiscus circularis* (Clark and Campbell)

Plate 4, figs. 10, 12

*Porodiscus (Trematodiscus) circularis* CLARK and CAMPBELL 1942,  
p. 42, pl. 11, figs. 2, 6, 10. – KOZLOVA 1999, pl. 9, fig. 2

Species code: **ccir**

*Cromyomma riedeli* Nishimura  
Plate 3, figure 7

*Cromyomma riedeli* NISHIMURA 1992, p. 322, pl. 1, figs. 6, 7; pl. 11,  
fig. 7

Species code: **cred**

*Dendrospyris fragoides* Sanfilippo and Riedel

*Dendrospyris fragoides* SANFILIPPO and RIEDEL 1973, p. 526, pl. 15,  
figs. 8-13; pl. 31, figs. 13, 14

Species code: **dfra**

*Dendrospyris golli* Nishimura

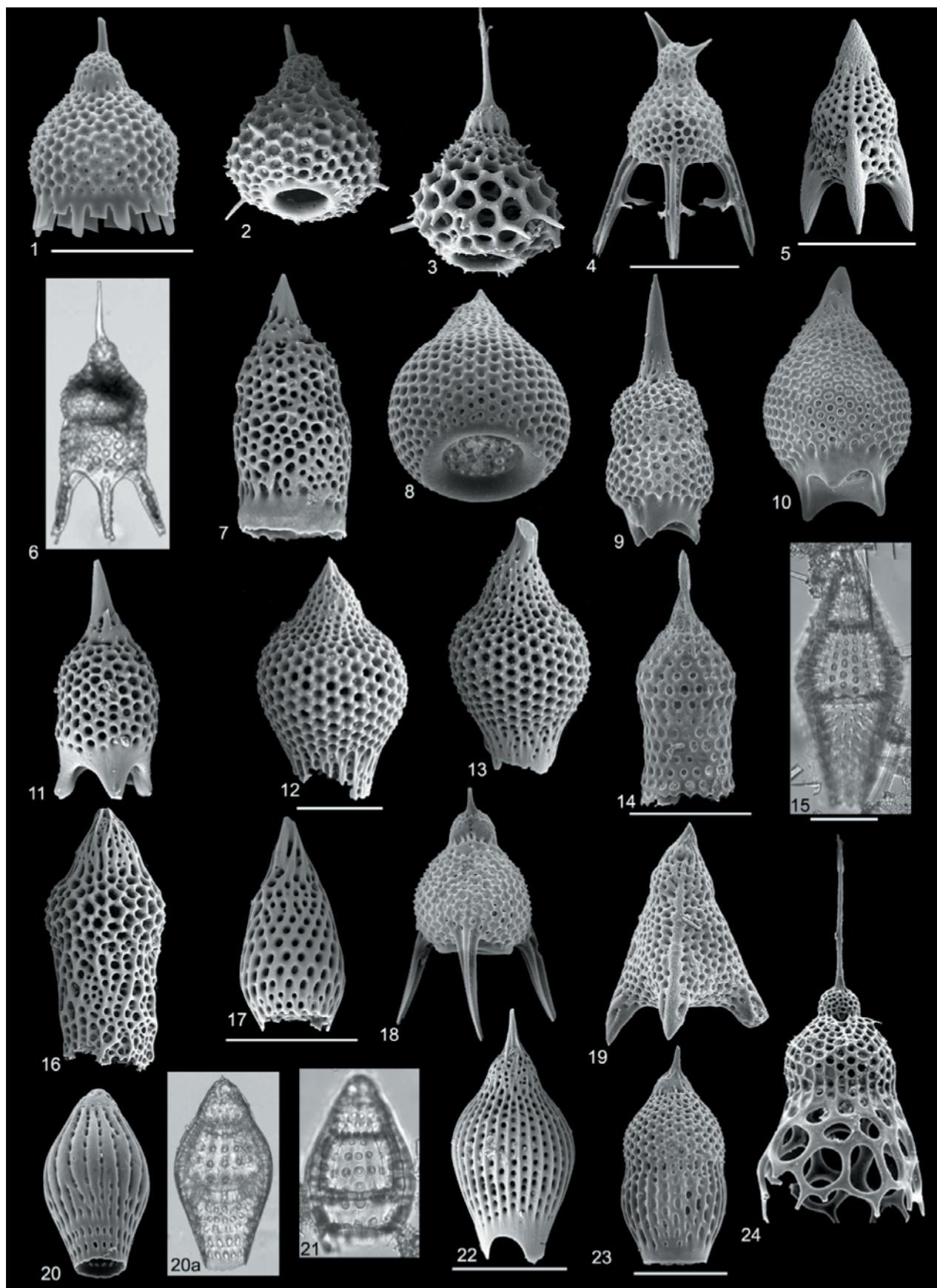
*Dendrospyris golli* NISHIMURA 1992, p. 330, pl. 3, figs. 1, 2; pl. 12,  
fig. 11

Species code: **dgol**

## PLATE 2

All scale bars equal 100µm.

- 1 *Calocyclus hispida* (Ehrenberg) s.s. DSDP 10-94-25-3 (113-115cm)
- 2 *Calocyclus ampulla* (Ehrenberg). ODP 171B-1051A-56x-4w (37-38cm)
- 3 *Calocyclus castum* (Haeckel). ODP 171B-1051A-44x-7w (8-10cm)
- 4 *Bekoma? oliva* Nishimura. DSDP 10-86-8-4 (119-125cm)
- 5 *Lithochytris archaea* Riedel and Sanfilippo. ODP 171B-1051A-44x-7w (8-10cm)
- 6 *Thrysocyrtis (Thrysocyrtis) tarsipes* Foreman. DSDP 10-96-3-5 (42-44 cm)
- 7 *Theocotyle nigriniae* Riedel and Sanfilippo. ODP 171B-1051A-44x-7w (8-10cm)
- 8 *Theocotylissa ficus* (Ehrenberg). DSDP 10-94-25-3 (113-115cm)
- 9 *Theocotylissa auctor* (Foreman). DSDP 10-96-3-5 (42-44 cm)
- 10 *Theocotylissa alpha* (Foreman). ODP 171B-1051A-44x-7w (8-10cm)
- 11 *Thrysocyrtis (Thrysocyrtis) rhizodon* Ehrenberg. ODP 171B-1051A-44x-7w (8-10cm)
- 12 *Lamptonium fabaeforme fabaeforme* (Krasheninnikov). ODP 171B-1051A-54x-5w (40-42cm)
- 13 *Lamptonium fabaeforme chaunothorax* Riedel and Sanfilippo. ODP 171 B- 1051 A-44x-7w (8-10cm)
- 14 *Pterocodon? ampla* (Brandt). DSDP 10-96-3-5 (42-44cm)
- 15 *Phormocyrtis turgida* (Krasheninnikov). ODP 171 B- 1051 A-52x-5w (42-44cm)
- 16 *Phormocyrtis cubensis* (Riedel and Sanfilippo). ODP 171B-1051A-44x-7w (8-10cm)
- 17 *Buryella clinata* Foreman. ODP 171B-1051A-44x-7w (8-10cm)
- 18 *Lychocanoma bellum* (Clark and Campbell). ODP 171B-1051A-44x-7w (8-10cm)
- 19 *Lithochytris vespertilio* Ehrenberg. DSDP 10-94-25-3 (113-115cm)
- 20 *Buryella tetradica* Foreman s.s. DSDP 10-96-3-5 (42-44cm)
- 21 *Buryella tetradica* Foreman var. A. ODP 171B-1051A-65x-5w (40-42cm)
- 22 *Podocyrtis papalis* (Ehrenberg). ODP 171 B- 1051 A-44x-7w (8-10cm)
- 23 *Theocorys? phyzella* Foreman. DSDP 10-96-3-5 (42-44cm)
- 24 *Theocorys acroria* Foreman. DSDP 10-96-3-5 (42-44cm)



*Dictyoprora amphora* (Haeckel)  
Plate 3, figure 18

*Dictyocephalus amphora* HAECKEL 1887, p. 1305, pl. 62, fig. 4. – NI-  
GRINI 1977, p. 250, pl. 4, figs. 1, 2

Species code: **damp**

*Dictyoprora mongolfieri* (Ehrenberg)  
*Eucyrtidium mongolfieri* EHRENBURG 1854, pl. 36, fig. 18.  
– NIGRINI 1977, p. 250, pl. 4, fig. 7

Species code: **dmon**

*Dictyoprora urceolus* (Haeckel)  
Plate 3, figure 17

*Dictyocephalus urceolus* HAECKEL 1887, p. 1305. – NIGRINI 1977, p.  
251, pl. 4, figs. 9, 10

Species code: **durc**

*Dictyospyris discus* Sanfilippo and Riedel  
*Dictyospyris discus* SANFILIPPO and RIEDEL 1973, p. 527, pl. 16,  
figs. 4-8; pl. 32, figs. 4-7

Species code: **ddis**

*Dorcadospyris platyacantha* (Ehrenberg) group  
Plate 4, figure 17

*Petalospyris platyacantha* EHRENBURG 1873, p. 247; 1875, pl. 22, fig.  
8. – SANFILIPPO and RIEDEL 1973, pl. 17, figs. 11-15; pl. 33, fig.  
2

Species code: **dpla**

*Entapium regulare* Sanfilippo and Riedel  
*Entapium regulare* SANFILIPPO and RIEDEL 1973, p. 492, pl. 2, figs.  
10-19; pl. 24, figs. 1-3

Species code: **ereg**

*Girafospyris cyrillum* Sanfilippo and Riedel  
*Girafospyris cyrillum* SANFILIPPO and RIEDEL 1973, p. 528, pl. 18,  
figs. 1-3; pl. 33, fig. 3

Species code: **gcyr**

*Girafospyris lata* Goll  
Plate 4, figure 16

*Girafospyris lata* GOLL 1969, p. 334, pl. 58, figs. 22, 24–26

Species code: **glat**

*Hexacontium? palaeocenicum* Sanfilippo and Riedel  
Plate 3, figure 5

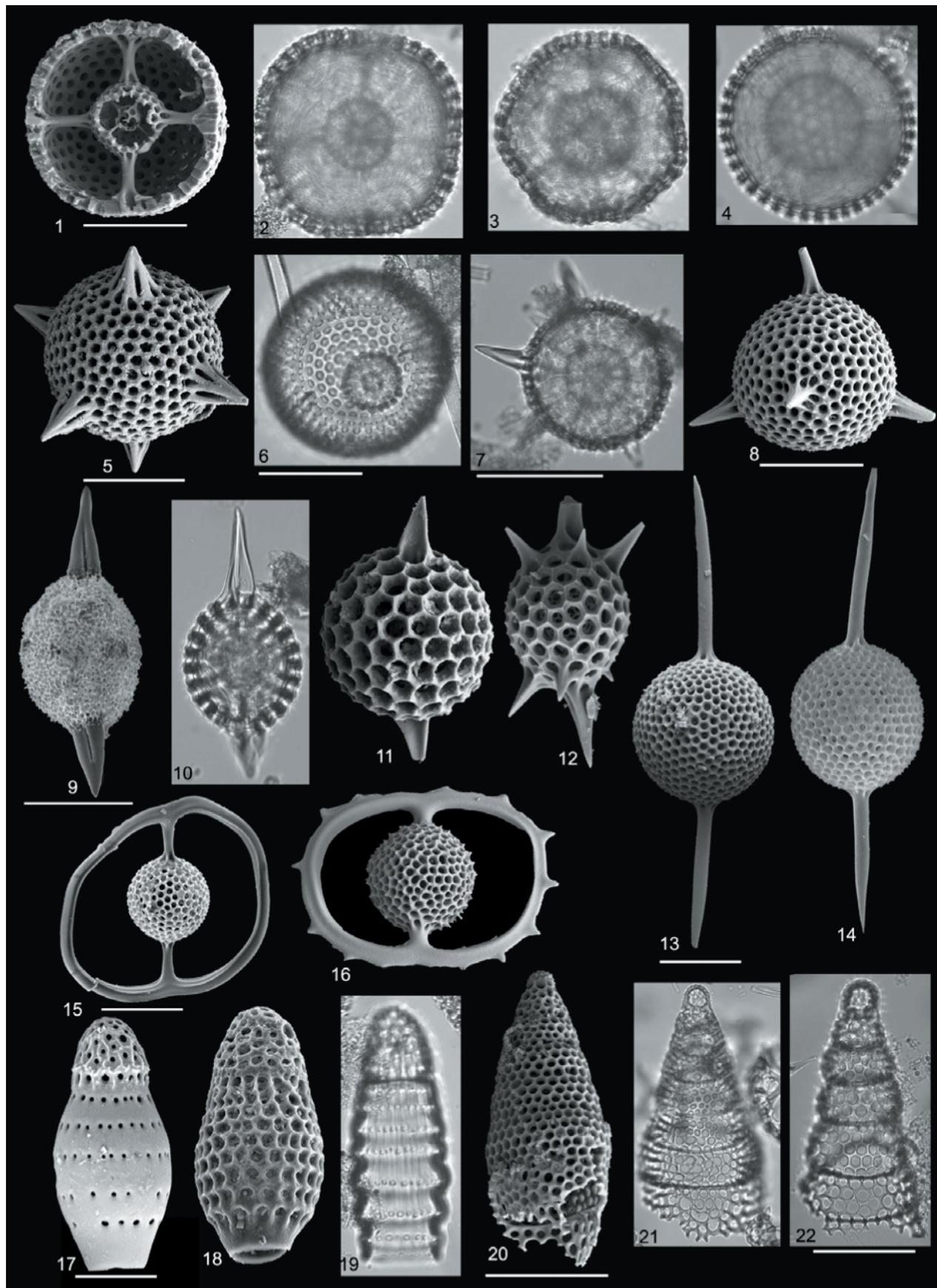
*Hexacontium? palaeocenicum* SANFILIPPO and RIEDEL 1973, p.  
492, pl. 4, fig. 2; pl. 24, fig. 4

Species code: **hpal**

### PLATE 3

All scale bars equal 100µm.

- 1 *Thecosphaera larnacium* Sanfilippo and Riedel. ODP 171B-1051A-56x-4w (37-38cm)
- 2 *Thecosphaera larnacium* Sanfilippo and Riedel. ODP 171B-1051A-44x-5w (48-50cm)
- 3 *Thecosphaerella glebulenta* Sanfilippo and Riedel. ODP 171B-1051A-42x-4w (22-23cm)
- 4 *Carposphaera subbotinae* (Borisenko). DSDP 43-384-7-6 (119-125cm)
- 5 *Hexacontium? palaeocenicum* Sanfilippo and Riedel. ODP 171B-1051A-57x-6w (40-42cm)
- 6 *Thecosphaerella ptomatus* Sanfilippo and Riedel. ODP 171B-1051A-67x-1w (34-36cm)
- 7 *Cromyomma riedeli* Nishimura. ODP 171B-1051A-65x-5w (40-42cm)
- 8 *Pseudostaurosphaera* sp. ODP 171B-1051A-44x-7w (8-10cm)
- 9 *Spongatractus pachystylus* (Ehrenberg). DSDP 10-94-32-CC
- 10 *Stylosphaera coronata coronata* Ehrenberg. ODP 171B-1051A-47x-7w (28-30cm)
- 11 *Stylosphaera macrosphaera* Nishimura. ODP 171B-1051A-61x-4w (40-42cm)
- 12 *Stylosphaera goruna* Sanfilippo and Riedel. ODP 171B-1051A-67x-1w (34-36cm)
- 13 *Amphisphaera minor* (Clark and Campbell). ODP 171B-1051A-67x-2w (40-42cm)
- 14 *Axoprunum pierinae* (Clark and Campbell). ODP 171B-1051A-58x-7w (16-18cm)
- 15 *Saturnalis circularis* Haeckel. DSDP43-384-7-6 (119-125cm)
- 16 *Saturnalis* sp. A aff. *trochoides* Haeckel. DSDP 43-384-6-3-(102-109cm)
- 17 *Theocampe urceolus* (Haeckel). DSDP 10-94-32-CC
- 18 *Theocampe amphora* (Haeckel). DSDP 10-94-32-CC
- 19 *Siphocampe altamontensis* (Campbell and Clark). ODP 171B-1051A-64x-2w (51-52cm)
- 20 *Amphipternis clava* (Ehrenberg). ODP 171B-1051A-67x-1w (34-36cm)
- 21 *Amphipternis clava* (Ehrenberg). ODP 171B-1051A-59x-2w (36-38cm)
- 22 *Amphipternis alamedaensis* (Campbell and Clark). DSDP 43-384-6-3 (102-109cm)



**Lamptonium? columbus** Foreman  
Plate 1, figure 8

*Lamptonium? columbus* FOREMAN 1973, p. 435, pl. 6, fig. 2; pl. 11,  
figs. 15, 19

Species code: **Icol**

**Lamptonium? sp. A**

Plate 1, figure 9

**Description:** Large shell of three segments, inflated thorax bearing three pronounced wings. Sparsely perforate subspherical cephalis. The majority of specimens have only one horn which is never observed complete. By-spines, arising from the massive shell material on the cephalis or the uppermost part of the thorax, are observed in some specimens. The wings extend outward from the upper third of the thorax. The shape of the thorax varies somewhat depending on the position of the wings, from inflated pyriform to conical proximally and inflated distally. The subcylindrical, fragile abdomen consists of a hyaline band, varying in width proximally and a porous part extending distally. Pores on this part are circular to sub-circular, irregular in size and distribution.

**Distinguishing characters:** This species is distinguished from *Lamptonium? columbus* by having more prominent thoracic wings and a distinctive abdominal hyaline band. It differs from *Lamptonium pennatum* by the higher position of the wings on the thorax, having an abdominal hyaline band not a fringe and in the nature of the apical horn.

**Occurrence:** Late early Paleocene to late Paleocene of ODP 171B-1051A

Species code: **Imar**

**Lamptonium? incohatum** Foreman

*Lamptonium? incohatum* FOREMAN 1973, p. 436, pl. 6, fig. 1; pl. 11,  
fig. 18

Species code: **linc**

**Lamptonium fabaeforme chaunothorax** Riedel and Sanfilippo

Plate 2, figure 13

*Lamptonium (?) fabaeforme (?) chaunothorax* RIEDEL and SANFILIPPO 1970, p. 524, pl. 5, figs. 8-9

Species code: **Icha**

**Lamptonium fabaeforme constrictum** Riedel and Sanfilippo

*Lamptonium (?) fabaeforme (?) constrictum* RIEDEL and SANFILIPPO 1970, p. 523, pl. 5, fig. 7

Species code: **Icon**

**Lamptonium fabaeforme fabaeforme** (Krasheninnikov)

Plate 2, figure 12

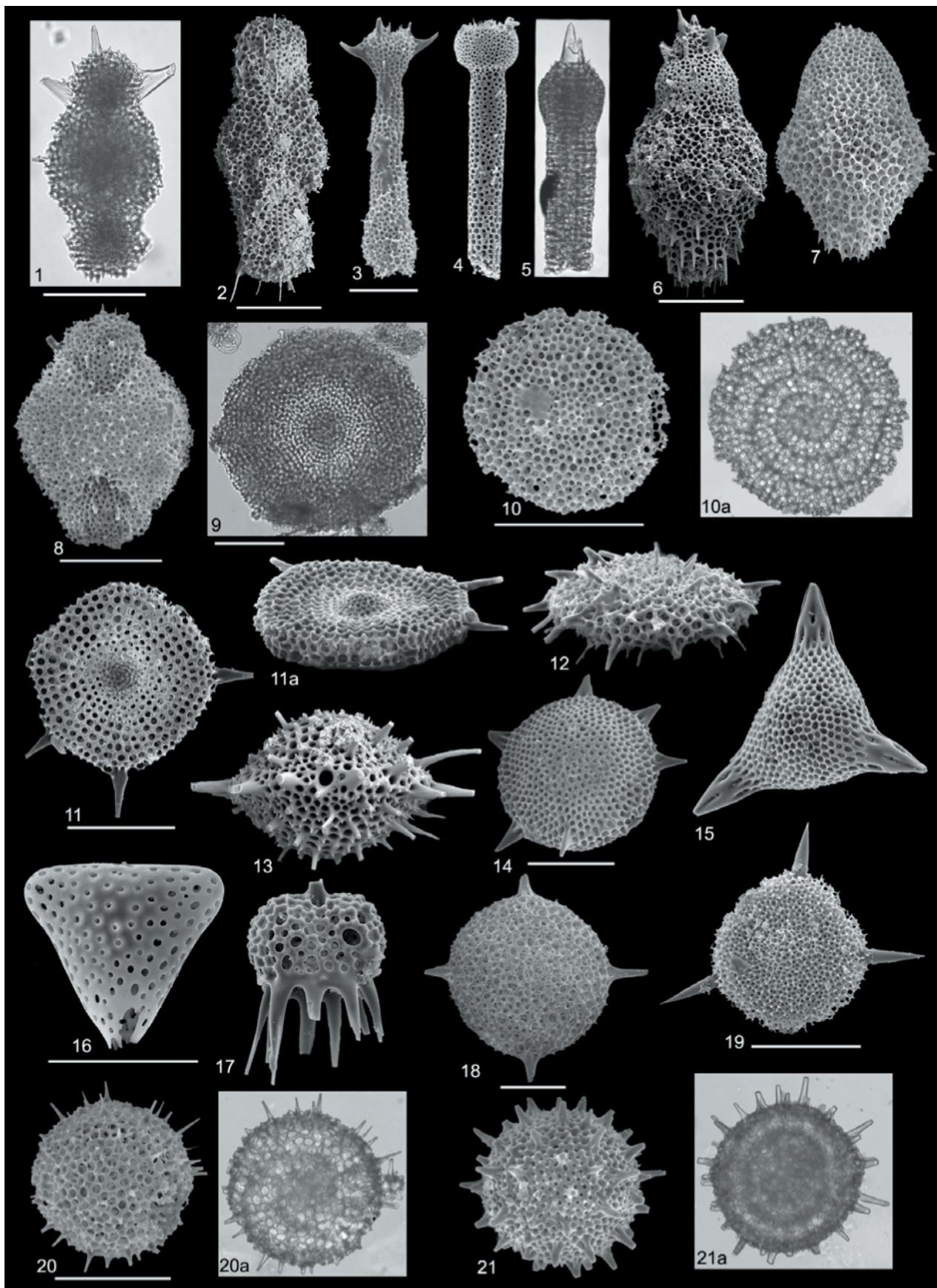
*Cyrtocalpis fabaeforme* KRASHENINNIKOV 1960, p. 296, pl. 3, fig. 11.  
– RIEDEL and SANFILIPPO 1970, p. 523, pl. 5, fig. 6

Species code: **Ifab**

#### PLATE 4

All scale bars equal 100µm.

- 1 *Amphicraspedum murrayanum* Haeckel group.  
ODP 171B-1051A-44x-7w (8-10cm)
- 2 *Amphicraspedum murrayanum* Haeckel group.  
ODP 171B-1051A-56x-4w (37-38cm)
- 3 *Amphicraspedum prolixum* Sanfilippo and Riedel group. DSDP 10-94-25-3 (113-115cm)
- 4 *Amphicraspedum prolixum* Sanfilippo and Riedel group. DSDP 10-96-3-5 (42-44cm)
- 5 *Amphicraspedum prolixum* Sanfilippo and Riedel group. ODP 171B-1051A-44x-7w (8-10cm)
- 6 *Spongurus? irregularis* Nishimura. ODP 171B-1051A-57x-6w (40-42cm)
- 7 *Spongurus? irregularis* Nishimura. DSDP 43-384-7-6 (119-125cm)
- 8 *Spongodiscus cruciferus* (Clark and Campbell).  
DSDP 10-94-25-3 (113-115cm)
- 9 *Spongodiscus americanus* Kozlova . ODP 171B-1051A-66-CC
- 10 *Circodiscus circularis* (Clark and Campbell).  
DSDP 43-384-7-6 (119-125cm)
- 11 *Spongodiscus quartus quartus* (Borisenko).
- 12 *Circodiscus circularis* (Clark and Campbell).  
DSDP 10-96-3-5 (42-44cm)
- 13 *Stylocrochus nitidus* Sanfilippo and Riedel. ODP 171B-1051A-44x-7w (8-10cm)
- 14 *Periphrena heliastericus* (Clark and Campbell).  
DSDP 10-86-8-4 (119-125cm)
- 15 *Periphrena tripyramis triangula* (Haeckel).  
DSDP 10-94-32-CC
- 16 *Girafospyris lata* Goll. DSDP 10-96-3-5 (42-44cm)
- 17 *Dorcadospyris platyacantha* (Ehrenberg) group.  
DSDP 10-96-3-5 (42-44cm)
- 18 *Spongodiscus rhabdostylus* (Ehrenberg) group.  
ODP 171B-1051 A-58x-7w (16-18cm)
- 19 *Spongodiscus rhabdostylus* (Ehrenberg) group.  
DSDP 10-94-25-3 (113-115cm)
- 20 *Lithelius foremanae* Sanfilippo and Riedel. ODP 171B-1051A-58x-2w (34-36cm)
- 21 *Lithelius hexaxyphophorus* (Clark and Campbell).  
DSDP 10-94-25-3 (113-115cm)



**Lamptonium pennatum** Foreman

Plate 1, figure 10

*Lamptonium pennatum* FOREMAN 1973, p. 436, pl. 6, figs. 3-5; pl. 11, fig. 13

Species code: **lpen**

**Lamptonium sanfilippiae** Foreman

*Lamptonium sanfilippiae* FOREMAN 1973, p. 436, pl. 6, figs. 15-16; pl. 11, figs. 16-17

Species code: **lsan**

**Lithelius foremanae** Sanfilippo and Riedel

Plate 4, figure 20

*Lithelius foremanae* SANFILIPPO and RIEDEL 1973, p. 522, pl. 7, figs. 1-6; pl. 26, figs. 4, 5

Species code: **lfor**

**Lithelius hexaxyphophorus** (Clark and Campbell)

Plate 4, figure 21

*Stylosphaera hexaxyphophora* CLARK and CAMPBELL 1942, p. 28, pl. 6, figs. 4, 5, 7, 11, 12. – SANFILIPPO and RIEDEL 1973, p. 522, pl. 7, figs. 7-9; pl. 26, figs. 6-7

Species code: **lhex**

**Lithochytris archaea** Riedel and Sanfilippo

Plate 2, figure 5

*Lithochytris archaea* RIEDEL and SANFILIPPO 1970, p. 528, pl. 9, fig. 7

Species code: **larc**

**Lithochytris vespertilio** Ehrenberg

Plate 2, figure 19

*Lithochytris vespertilio* EHRENBURG 1873, p. 239; 1875, pl. 4, fig. 10; RIEDEL and SANFILIPPO 1970, p. 528, pl. 9, figs. 8-9

Species code: **lves**

**Lithocyclia ocellus** Ehrenberg group

*Lithocyclia ocellus* EHRENBURG group 1854, pl. 36, fig. 30; 1873, p. 240; RIEDEL and SANFILIPPO 1973, p. 744, pl. 2, figs. 7-8

Species code: **loce**

**Lophocyrtis biaurita** (Ehrenberg)

*Eucyrtidium biaurita* EHRENBURG 1873, p. 226; 1875, p. 70, pl. 10, figs. 7-8. – FOREMAN 1973, p. 442, pl. 8, figs. 23-26

Species code: **ibia**

**Lychnocanium carinatum** Ehrenberg

Plate 1, figure 25

*Lychnocanium carinatum* EHRENBURG 1875, p. 78, pl. 8, fig. 5. – NISHIMURA 1987, p. 727, pl. 3, fig. 11, non 6

*Distinguishing characters:* *Lychnocanium* is defined by two thick walled first segments bearing an apical horn, a slightly to heavily ribbed thorax from which extend three solid feet and usually a thin-walled third segment connecting them. *Lychnocanium* Ehrenberg differs from *Lychnocanoma* Haeckel by having a smaller cephalis and a ribbed thorax.

Species code: **lcar**

**Lychnocanium** sp. A

Plate 1, figure 26

*Theopodium* sp. aff. *Lychnocanium carinatum* Ehrenberg. – NISHIMURA 1992, p. 354, pl. 7, fig. 11. – JIANBING and AITCHISON 2002, p. 150, pl. 2, fig. 6

*Distinguishing characters:* This form is distinguished from *Lychnocanium carinatum* Ehrenberg by its shorter feet and very finely latticed shell below the thorax. Feet are slender and triradiate. It is thought that *Lychnocanium carinatum* originated from this form by elongation of the legs and development of a third segment.

*Occurrence:* Late early Paleocene to late Paleocene of ODP 171B-1051A and late Paleocene of DSDP 43-3 84

Species code: **lyca**

**Lychnocanoma anacolum** Foreman

Plate 1, figure 24

*Lychnocanoma anacolum* FOREMAN 1973, p. 437, pl. 1, fig. 19; pl. 11, fig. 7

Species code: **lana**

**Lychnocanoma auxilla** Foreman

Plate 1, figure 20

*Lychnocanoma auxilla* FOREMAN 1973, p. 437, pl. 2, fig. 6; pl. 11, figs. 1, 2

Species code: **laux**

**Lychnocanoma bellum** (Clark and Campbell)

Plate 2, figure 18

*Lychnocanum bellum* CLARK and CAMPBELL 1942, p. 72, pl. 9, figs. 35, 39. – FOREMAN 1973, p. 437, pl. 1, fig. 17; pl. 11, fig. 9

Species code: **lbel**

**Lychnocanoma? costata** Nishimura

Plate 1, figure 4

*Lychnocanoma? costata* NISHIMURA 1992, p. 342, pl. 6, figs. 4-6

Species code: **lcos**

**Lychnocanoma? pileus** Nishimura

Plate 1, figure 3

*Lychnocanoma? pileus* NISHIMURA 1992, p. 344, pl. 6, figs. 7, 8; pl. 13, fig. 5

Species code: **lipil**

**Orbula comitata** Foreman

Plate 1, figure 2

*Orbula comitata* FOREMAN 1973, p. 437, pl. 3, fig. 11; pl. 10, figs. 7, 8

Species code: **ocom**

**Orbula discipulus** Foreman

Plate 1, figure 1

*Orbula discipulus* FOREMAN 1973, p. 438, pl. 3, fig. 10 ; pl. 10, figs. 9, 10

Species code: **odis**

*Peripaena decora* Ehrenberg

*Peripaena decora* EHRENBERG 1873, p. 246; 1875, pl. 28, fig. 6.  
– SANFILIPPO and RIEDEL 1973, p. 523, pl. 8, figs. 8-10; pl. 27,  
figs. 2-5

Species code: **pdec**

*Peripaena delta* Sanfilippo and Riedel

*Peripaena delta* SANFILIPPO and RIEDEL 1973, p. 523, pl. 8, figs. 11,  
12; pl. 27, figs. 6, 7

Species code: **pdel**

*Peripaena heliastericus* (Clark and Campbell)

Plate 4, figure 14

*Heliodiscus heliastericus* CLARK and CAMPBELL 1942, p. 39, pl. 3,  
figs. 10, 11. – KOZLOVA 1999, p. 87, pl. 34, figs. 2, 11, 12; pl. 40, fig.  
4

Species code: **phel**

*Peripaena tripyramis triangula* (Sutton)

Plate 4, figure 15

*Phacotriactis triangula* SUTTON 1896, p. 61 (unillustrated)

*Triactis tripyramis triangula* RIEDEL and SANFILIPPO 1970, p. 521,  
pl. 4, figs. 9, 10 – SANFILIPPO and RIEDEL 1973, p. 523, pl. 9, figs.  
10, 11

Species code: **ptri**

*Peripaena tripyramis tripyramis* (Haeckel)

*Triactiscus tripyramis* HAECKEL 1887, p. 432, pl. 33, fig. 6 – SAN-  
FILIPPO and RIEDEL 1973, p. 523, pl. 9, figs. 7-9

Species code: **ptrp**

*Phormocyrtis cubensis* (Riedel and Sanfilippo)

Plate 2, figure 16

*Eucyrtidium cubense* RIEDEL and SANFILIPPO 1971, p. 1594, pl. 7,  
figs. 10, 11

Species code: **pcub**

*Phormocyrtis striata exquisita* (Kozlova)

Plate 1, figure 18

*Podocyrtis exquisita* Kozlova in KOZLOVA and GORBOVETS 1966,  
p. 106, pl. 17, fig. 2. – FOREMAN 1973, p. 438, pl. 7, figs 1-4, 7-8; pl.  
12, fig. 5

Species code: **pexq**

*Phormocyrtis striata praexquisita* Nishimura

Plate 1, figure 17

*Phormocyrtis striata praexquisita* NISHIMURA 1992, p. 346, pl. 9, figs.  
1-3

Species code: **ppra**

*Phormocyrtis striata striata* Brandt

Plate 1, figure 19

*Phormocyrtis striata* Brandt in WETZEL 1935, p. 55, pl. 9, fig. 12. –  
FOREMAN 1973, p. 438, pl. 7, figs. 5, 6, 9

Species code: **pstr**

*Phormocyrtis turgida* (Krasheninnikov)

Plate 2, figure 15

*Lithocampe turgida* KRASHENINNIKOV 1960, p. 301, pl. 3, fig. 17  
– FOREMAN 1973, p. 438, pl. 7, fig. 10; pl. 12, fig. 6

Species code: **ptur**

*Pterocodon? ampla* (Brandt)

Plate 2, figure 14

*Theocyrtis ampla* Brandt in WETZEL 1935, p. 56, pl. 9, figs. 13–15.  
– FOREMAN 1973, p. 438, pl. 5, figs. 3-5

Species code: **ptam**

*Pterocodon? anteclinata* Foreman

*Pterocodon? anteclinata* FOREMAN 1975, p. 621, pl. 9, figs. 32-34

Species code: **pant**

*Pterocodon poculum* Nishimura

Plate 1, figure 13

*Pterocodon poculum* NISHIMURA 1992, p. 350, pl. 8, figs. 1-3; pl. 13,  
fig. 13

Species code: **ppoc**

*Podocyrtis (Lampterium) sinuosa* Ehrenberg

*Podocyrtis sinuosa* EHRENBERG 1873, p. 253; 1875, pl. 15, fig. 5 –  
RIEDEL and SANFILIPPO 1970, p. 534, pl. 11, figs. 3, 4

Species code: **psin**

*Podocyrtis (Podocyrtoges) aphorma* Riedel and Sanfilippo

*Podocyrtis (Lampterium) aphorma* RIEDEL and SANFILIPPO 1970,  
p. 534, pl. 11, fig. 2

Species code: **paph**

*Podocyrtis (Podocyrtoges) diamesa* Riedel and Sanfilippo

*Podocyrtis (Podocyrtis) diamesa* RIEDEL and SANFILIPPO 1970, p.  
533, pl. 12, fig. 4, non figs. 5, 6

Species code: **pdim**

*Podocyrtis (Podocyrtis) papalis* Ehrenberg

Plate 2, figure 22

*Podocyrtis papalis* EHRENBERG 1847, p. 55, fig. 2. – SANFILIPPO  
and RIEDEL 1973, p. 531, pl. 20, figs. 11-14, pl. 36, figs. 2, 3

Species code: **ppap**

*Rhopalocanium ornatum* Ehrenberg

*Rhopalocanium ornatum* EHRENBERG 1847, fig. 3; 1854, pl. 36, fig. 9.  
– FOREMAN 1973, p. 439, pl. 2, figs. 8-10; pl. 12, fig. 3

Species code: **rorn**

*Saturnalis circularis* Haeckel

Plate 3, figure 15

*Saturnalis circularis* HAECKEL 1887, p. 131 (unillustrated). – NIGRI-  
NI 1967 p. 24, pl. 1, fig. 9

**Saturnalis** sp. A aff. *S. trochoides* Haeckel  
Plate 3, figure 10

*Saturnalis* sp. A aff. *S. trochoides* HAECKEL 1887, p. 132 (unillustrated);  
Saturnalin with spiny ring, SANFILIPPO and RIEDEL 1973, pl. 22, fig. 3

**Siphocampe altamontensis** (Campbell and Clark)  
Plate 3, figure 19

*Tricolocampe (Tricolocamptra) altamontensis* CAMPBELL and CLARK 1944, p. 33, pl. 7, figs. 24, 26. – LING 1991, p. 320, pl. 1, fig. 12

**Siphocampe? quadrata** (Petrushevskaya and Kozlova)  
*Lithamphora sacculifera quadrata* PETRUSHEVSKAYA and KOZLOVA 1972, p. 539, pl. 30, figs. 4–6

Species code: **squa**

**Spongatractus balbis** Sanfilippo and Riedel  
*Spongatractus balbis* SANFILIPPO and RIEDEL 1973, p. 518, pl. 2, figs. 1–3; pl. 25, figs. 1, 2

Species code: **sbab**

**Spongatractus pachystylus** (Ehrenberg)  
Plate 3, figure 9

*Spongospaera pachystyla* EHRENBERG 1873, p. 256; 1875, pl. 26, fig. 3. – SANFILIPPO and RIEDEL 1973, p. 519, pl. 2, figs. 4–6; pl. 25, fig. 3

Species code: **spac**

**Stylosphaera coronata coronata** Ehrenberg  
Plate 3, figure 10

*Stylosphaera coronata* EHRENBERG 1873, p. 258; 1875, pl. 25, fig. 4; SANFILIPPO and RIEDEL 1973, p. 520, pl. 1, figs. 13–17; pl. 25, fig. 4

Species code: **scor**

**Stylosphaera coronata macrospheara** Nishimura  
Plate 3, fig. 11

*Stylosphaera coronata macrospheara* NISHIMURA 1992, p. 325, pl. 1, figs. 3, 4; pl. 11, fig. 1

Species code: **smac**

**Stylosphaera coronata sabaca** Sanfilippo and Riedel  
*Stylosphaera coronata sabaca* SANFILIPPO and RIEDEL 1973, p. 521, pl. 1, fig. 18; pl. 25, figs. 7, 8

Species code: **ssab**

**Stylosphaera goruna** Sanfilippo and Riedel  
Plate 3, figure 12

*Stylosphaera goruna* SANFILIPPO and RIEDEL 1973, p. 521, pl. 1, figs. 20–22; pl. 25, figs. 9, 10

Species code: **sgor**

**Spongodiscus americanus** Kozlova  
Plate 4, figure 9

*Spongodiscus americanus* Kozlova in KOZLOVA and GORBOVETZ 1966, p. 88, pl. 14, figs. 1, 2

Species code: **same**

**Spongodiscus cruciferus** (Clark and Campbell)  
Plate 4, figure 8

*Spongastericus cruciferus* CLARK and CAMPBELL 1942, p. 50, pl. 1, figs. 1–6, 8, 10, 11, 16–18. – SANFILIPPO and RIEDEL 1973, p. 524, pl. 11, figs. 14–17; pl. 28, figs. 10, 11

Species code: **scr**

**Spongodiscus phrix** Sanfilippo and Riedel  
*Spongodiscus phrix* SANFILIPPO and RIEDEL 1973, p. 525, pl. 12, figs. 1, 2; pl. 29, fig. 2

Species code: **spfr**

**Spongodiscus rhabdostylus** (Ehrenberg) group  
Plate 4, figures 18, 19

*Spongospaera rhabdostylus* EHRENBERG 1873, p. 256; 1875, pl. 26, figs. 1, 2; SANFILIPPO and RIEDEL 1973, p. 525, pl. 13, figs. 1–3; pl. 30, figs. 1, 2

**Remarks:** This species-group includes forms having 3 or 4 bladed or conical strong radial spines. The oldest specimens in this work from ODP leg 171B tend to have 4 conical radial spines.

Species code: **srha**

**Spongodiscus quartus bosoculus** Sanfilippo and Riedel  
*Spongodiscus quartus bosoculus* SANFILIPPO and RIEDEL 1973, p. 525, pl. 12, figs. 8–10; pl. 29, fig. 7

Species code: **sbos**

**Spongodiscus quartus quartus** (Borisenko)  
Plate 4, figure 11

*Staurodictya quartus* BORISENKO 1958, p. 96, pl. 2, fig. 5. – SANFILIPPO and RIEDEL 1973, p. 525, pl. 12, figs. 6, 7; pl. 29, figs. 5, 6

Species code: **spqu**

**Spongurus? irregularis** Nishimura  
Plate 4, figures 6, 7

*Spongurus? irregularis* NISHIMURA 1992, p. 327, pl. 2, figs. 7–9; pl. 12, figs. 3, 7

Species code: **sirr**

**Stylotrochus alveatus** Sanfilippo and Riedel  
*Stylotrochus alveatus* SANFILIPPO and RIEDEL 1973, p. 525, pl. 13, figs. 4, 5; pl. 30, figs. 3, 4

Species code: **salv**

**Stylotrochus nitidus** Sanfilippo and Riedel  
Plate 4, figure 13

*Stylotrochus nitidus* SANFILIPPO and RIEDEL 1973, p. 525, pl. 13, figs. 9–14; pl. 30, figs. 7–10

Species code: **snit**

***Theocorys acoria*** Foreman

Plate 2, figure 24

*Theocorys acoria* FOREMAN 1973, p. 439, pl. 5, figs. 11–13; pl. 12, fig. 2Species code: **tacr*****Theocorys anaclasta*** Riedel and Sanfilippo*Theocorys anaclasta* RIEDEL and SANFILIPPO 1970, p. 530, pl. 10, figs. 2, 3Species code: **tana*****Theocorys anapographa*** Riedel and Sanfilippo*Theocorys anapographa* RIEDEL and SANFILIPPO 1970, p. 530, pl. 10, fig. 4Species code: **tanp*****Theocorys? phyzella*** Foreman

Plate 2, figure 23

*Theocorys? phyzella* FOREMAN 1973, p. 440, pl. 5, fig. 8; pl. 12, fig. 1Species code: **tphy*****Theocorys?* sp. aff. *T. acoria*** Foreman*Theocorys?* sp. aff. *T. acoria* Foreman. – NISHIMURA 1992, pl. 4, fig. 7Species code: **tafa*****Thecosphaera larnacium*** Sanfilippo and Riedel

Plate 3, figure 1, 2

*Thecosphaera larnacium* SANFILIPPO and RIEDEL 1973, p. 521, pl. 3, figs. 4–6; pl. 25, figs. 13, 14Species code: **tlar*****Thecosphaerella glebulenta*** Sanfilippo and Riedel

Plate 3, figure 3

*Thecosphaerella glebulenta* SANFILIPPO and RIEDEL 1973, p. 521, pl. 3, figs. 12, 13; pl. 26, fig. 1Species code: **tgle*****Thecosphaerella ptomatus*** Sanfilippo and Riedel

Plate 3, figure 6

*Thecosphaerella ptomatus* SANFILIPPO and RIEDEL 1973, p. 521, pl. 3, figs. 14–18; pl. 26, fig. 2Species code: **tpto*****Thecosphaerella rotunda*** (Borisenko)*Thecosphaera rotunda* BORISENKO 1960, p. 222, pl. 1, fig. 3; pl. 3, figs. 2, 3. – SANFILIPPO and RIEDEL 1973, pl. 3, figs. 7–11; pl. 36, fig. 3Species code: **trot*****Theocotyle cryptocephala*** (Ehrenberg)*Eucyrtidium cryptocephalum* EHRENBERG 1873, p. 227; 1875, p. 70, pl. 11, fig. 11. – SANFILIPPO and RIEDEL 1982, p. 178, pl. 2, figs. 4–7Species code: **tcty*****Theocotyle nigriniae*** Riedel and Sanfilippo

Plate 2, figure 7

*Theocotyle cryptocephala* (?) *nigriniae* RIEDEL and SANFILIPPO 1970, p. 525, pl. 6, fig. 5, non 6Species code: **tnig*****Theocotylissa alpha*** Foreman*Theocotylissa alpha* FOREMAN 1973, p. 441, pl. 4, figs. 13–15, non 14; pl. 12, fig. 16 Plate 2, figure 10Species code: **talp*****Theocotylissa auctor*** Foreman

Plate 2, figure 9

*Theocotyle* (*Theocotylissa*) *auctor* FOREMAN 1973, p. 441, pl. 4, figs. 8–10; pl. 12, fig. 13Species code: **tauc*****Theocotylissa ficus*** (Ehrenberg)

Plate 2, figure 8

*Eucyrtidium ficus* EHRENNBERG 1873, p. 228; 1875, pl. 11, fig. 19 – SANFILIPPO and RIEDEL 1982, p. 180, pl. 2, figs. 19, 20Species code: **tfic*****Thrysocyrtis? annikae*** Nishimura*Thrysocyrtis?* *annikae* NISHIMURA 1992, p. 356, pl. 7, figs. 4–6 Species code: **tann*****Thrysocyrtis (Pentalacorys) tensa*** Foreman*Thrysocyrtis hirsuta* FOREMAN 1973, p. 442, pl. 3, figs. 13–16; pl. 12, fig. 8Species code: **tten*****Thrysocyrtis (Thrysocyrtis) hirsuta*** (Krasheninnikov)*Podocyrtis hirsutus* KRASHENINNIKOV 1960, p. 300, pl. 3, fig. 16. – SANFILIPPO and RIEDEL 1982, p. 173, pl. 1, figs. 3, 4Species code: **thir*****Thrysocyrtis (Thrysocyrtis) rhizodon*** Ehrenberg

Plate 2, figure 11

*Thrysocyrtis rhizodon* EHRENBERG 1873, p. 262; 1875, p. 94, pl. 12, fig. 1. – SANFILIPPO and RIEDEL 1982, p. 173, pl. 1, figs. 14–16Species code: **trhi*****Thrysocyrtis (Thrysocyrtis) robusta*** Riedel and Sanfilippo*Thrysocyrtis hirsuta robusta* RIEDEL and SANFILIPPO 1970, p. 526, pl. 8, fig. 1Species code: **trob*****Thrysocyrtis (Thrysocyrtis) tarsipes*** Foreman*Thrysocyrtis (Thrysocyrtis) tarsipes* FOREMAN 1973, p. 442, pl. 3, fig. 9; pl. 12, fig. 14, Plate 2, figure 6Species code: **ttar**

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## REFERENCES

- ABELMANN, A., 1990. Oligocene to Middle Miocene radiolarian stratigraphy of southern high latitudes from Leg 113, Sites 689 and 690, Maud Rise. In: Barker, P.F., Kennett, J.P. et al. Eds., *Proceedings of the Ocean Drilling Program, Scientific Results 113*, 675-708. College Station TX: Ocean Drilling Program.
- AUBRY, M.-P., 1995. From chronology to stratigraphy: interpreting the lower and middle Eocene stratigraphic record in the Atlantic Ocean. In: Berggren, W. A., Kent, D. V., Aubry, M.-P. and Hardenbol, J. Eds., *Geochronology, Time Scales and Global Stratigraphic Correlation. A Unified Temporal Framework for a Historical Geology*, 213-274. Tulsa: SEPM Special Publication, 54.
- BERGGREN, W.A., KENT, D.V., SWISHER, C.C., III, and AUBRY, M.-P., 1995. A revised Cenozoic geochronology and chronostratigraphy. In: Berggren, W.A., Kent, D.V., Aubry, M.-P., and Hardenbol, J. Eds., *Geochronology, Time Scales and Global Stratigraphic Correlation. A Unified Temporal Framework for a Historical Geology*, 129-212. Tulsa: SEPM Special Publication, 54.
- BERGGREN, W.A., AUBRY, M.-P., VAN FOSSEN, M., KENT, D.V., NORRIS, R.D and QUILLEVERE, F. 2000. Integrated Paleocene calcareous plankton magnetobiochronology and stable isotope stratigraphy: DSDP Site 384 (NW Atlantic Ocean). *Palaeogeography, Palaeoclimatology, Palaeoecology*, 159: 1-51.
- BORISENKO, N. I., 1958. Palaeocene Radiolaria of Western Kuban. In: Krylov, A. P. Ed. Problems in Geology, Drilling and Exploitation of Wells, *Trudy Vsesoyuznyi Neftegazovyi Nauchno-Issledovatel'skii Institut (VNI), Krasnodarskii Filial*, 2:81-100.
- , 1960. New radiolarians from the Paleocene deposits of the Kuban. *Trudy Vsesoyuznyi Neftegazovyi Nauchno-Issledovalelskii Institut (VNI), Krasnodarskii Filial*, 4: 199-207.
- BRANDT, R., 1935. Die Mikropalaeontologie des Heiligenhafener, Kieseltones (Ober-Eozän) Radiolarien, Systematik. In: Wetzel, E.O., Ed., *Jahresbericht des Niedersächsischen Geologischen Vereins*, 27: 48-59
- BUKRY, D., 1973. Low latitude coccolith biostratigraphic zonation. In: Edgar, N.T., Saunders, J.B. et al., Eds., *Initial Reports of the Deep Sea Drilling Project 15*. Washington, DC: US Government Printing Office, 684-703
- , 1975. Coccolith and silicoflagellate stratigraphy, northwestern Pacific Ocean, Ocean Drilling Project Leg 32. In: Larson, R. L., Moberly, R. et al., Eds., *Initial Reports of the Deep Sea Drilling Project, 32*, Washington, DC: US Government Printing Office, 685-677.
- CAMPBELL, A. S. and CLARK, B. L., 1944. Radiolaria from Upper Cretaceous of Middle California. *Geological Society of America, Special Papers*, 57: 1-61.
- CANDE S.C. and KENT D.V., 1995. Revised calibration of the geomagnetic polarity timescale for the Late Cretaceous and Cenozoic. *Journal of Geophysical Research*, 97(B10): 13917-13951.
- CAULET, J.P. 1991. Radiolarians from the Kerguelen Plateau, Leg 119. In: J. Barron, B. Larsen et al., Eds., *Proceedings of the Ocean Drilling Program, Scientific Results 119*. College Station, TX: Ocean Drilling Program, 513-546
- CLARK, B.L., AND CAMPBELL, A.S., 1942. Eocene radiolarian faunas from the Mt. Diablo area, California. *Geological Society of America, Special Papers*, 39: 1-112
- DE WEVER, P., DUMITRICA, P., CAULET, J-P., NIGRINI C. and CARIDROIT, M., 2001. *Radiolarians in the sedimentary record*, Amsterdam: Gordon and Breach Science Publishers, 533 pp.
- DEFLANDRE, G., 1953. Radiolaires fossiles. In: Grassé P.P., Ed., *Traité de Zoologie*, 1 (2), 389-436. Paris: Masson.
- DISERENS, M.-O., BAUMGARTNER, P. O. and DUMITRICA, P., 2003. Age determination of late Cretaceous radiolites in orogenic environments: an example from accreted terrains of southern Costa Rica. Interrad X 2003, *Abstracts & Programme*, 49-50. University of Lausanne, Switzerland.
- DUMITRICA, P. 1985. Internal morphology of the Saturnalidae (Radiolaria): systematic and phylogenetic consequences. *Revue de Micropaléontologie*, 28 (3): 181-196
- , 1989. Internal skeletal structures of the superfamily Pylonacea (Radiolaria), a basis of a new systematics. *Revista española de Micropaleontología* 21 (2): 207-264.
- EHRENBERG, C. G., 1847. Über die mikroskopischen kieselschaligen Polycystinen als mächtige Gebirgsmasse von Barbados und über das Verhältniss deraus mehr als 300 neuen Arten bestehenden ganz eignethümlichen Formengruppe jener Felssmasse zu den jetzt lebenden Thieren und zur Kreidebildung. Eine neue Anregung zur Erforschung des Erdlebens. *Königliche Preussische Akademie der Wissenschaften zu Berlin, Monats Bericht, Jahre 1847*: 40-60.
- EHRENBERG, C.G., 1854. *Mikrogeologie: Das Erden und Felsen schafende Wirken des unsichtbar kleinen selbständigen Lebens auf der Erde*. Leipzig: Leopold Voss, 374 pp.
- , 1873. Größere Felsproben des Polycystinen-Mergels von Barbados mit weiteren Erläuterungen. *Königliche preussischen Akademie der Wissenschaften zu Berlin, Monatsberichte, Jahre 1873*: 213-263.
- , 1875. Fortsetzung der mikrogeologischen Studien als Gesammt-Uebersicht der mikroskopischen Palaontologie gleichartig analysirter Gebirgsarten der Erde, mit specieller Rücksicht auf den Polycystinen-Mergel von Barbados. *Königliche Akademie der Wissenschaften zu Berlin, Abhandlungen, Jahre, 1875*: 1-225.
- FLORES ALBIN, E., 1983. Radiolarios de algunas formaciones del Cretacico-Paleogeno Inferior de Cuba occidental. Translated Title: Radiolarians from Cretaceous and lower Paleogene formations in western Cuba. *Ciencias de la Tierra y del Espacio*, 7: 3-19.
- FLORES ALBIN, E. and FERNANDEZ-RODRIGUEZ, G., 1985. Acerca de la bioestratigrafía del límite entre 10s depósitos del paleoceno-eoceno inferior (formaciones Alcázar y Capdevila) en la provincia Ciudad Habana. *Revista Tecnológica*, 15, Serie: Geología, 1: 8-18.
- FOREMAN, H.P., 1968. Upper Maestrichtian Radiolaria of California. *Special Papers in Palaeontology*, 31: 1-82.
- , 1973. Radiolaria of Leg 10 with systematics and ranges for the families Amphipyndacidae, Artostrobiidae and Theoperidae. In: Worzel, J. L., Bryant, W. et al., Eds., *Initial Reports of the Deep Sea Drilling Project, 10*, 407-474. Washington, DC: US Government Printing Office.

- , 1975. Radiolaria from the North Pacific, Deep Sea Drilling Project, Leg 32. In : Larson, R. L., Moberly, R. et al., Eds., *Initial Reports of the Deep Sea Drilling Project 32*, 579–676. Washington, DC: US Government Printing Office.
- GUEX, J. 1977. Une nouvelle méthode d'analyse biochronologique. *Bulletin de Géologie*, Lausanne, No 224: 309–322
- , 1991. *Biochronological Correlations*. Berlin/Heidelberg/New York: Springer-Verlag, 250 pp.
- GOLL, R. M., 1969. Classification and phylogeny of Cenozoic Trisocyclidae (Radiolaria) in the Pacific and Caribbean basins. Part II. *Journal of Paleontology*, 43(2): 322–339.
- GOLL, R. M. and BJORKLUND K.R., 1989. A new radiolarian biostratigraphy for the Neogene of the Norwegian Sea; ODP Leg 104. In: Eldholm, O., Thiede, J., Taylor, E. et al. Eds., *Proceedings of the Ocean Drilling Program, Scientific Results*, 104, 697–737. College Station, TX: Ocean Drilling Program.
- HAECKEL, E., 1862. *Die Radiolarien (Rhizopoda Radiolaria)- Eine monographie*. Berlin: Reimar, 572 p.
- , 1881. Entwurf eines Radiolarien-Systems auf Grund von Studien der Challenger. *Jenaische Zeitschrift für Naturwissenschaft*, 15 (8.3): 418–472.
- , 1887. Report on the Radiolaria collected by H.M.S. Challenger during the years 1873–1876. *Report on the Scientific Results of the Voyage of the H.M.S. Challenger, Zoology*, 18, clxxxviii + 1803 pp.
- HOLLIS, C. J., 1993. Latest Cretaceous to late Paleocene radiolarian biostratigraphy; a new zonation from the New Zealand region. *Marine Micropaleontology*, 21 (4): 295–327.
- , 1997. *Cretaceous-Paleocene Radiolaria from eastern Marlborough, New Zealand*. Lower Hutt, New Zealand: Institute of Geological and Nuclear Sciences Limited, Monograph 17, 152 pp.
- , 2002. Biostratigraphy and paleoceanographic significance of Paleocene radiolarians from offshore eastern New Zealand. *Marine Micropaleontology*, 46 (3-4): 265–316.
- , 2006. Radiolarian faunal turnover through the Paleocene-Eocene transition, Mead Stream, New Zealand. *Eclogae Geologicae Helvetiae*, 99, Supplement 1: S79–S99.
- HOLLIS, C.J., DICKENS G., FIELD B., JONES C., STRONG C., 2005. The Paleocene-Eocene transition at Mead Stream, New Zealand: a southern Pacific record of early Cenozoic global change, *Palaeogeography, Palaeoclimatology, Palaeoecology*, 215, 3 13–343.
- HOLLIS, C. J., WAGHORN, D. B., STRONG, C. P. and CROUCH, E. M., 1997. *Integrated Paleocene biostratigraphy of DSDP Site 277 (Leg 29): Foraminifera, calcareous nannofossils, Radiolaria and palynomorphs*. Lower Hutt, New Zealand: Institute of Geological and Nuclear Sciences Limited, Report 97/7, 87 pp.
- HAMMER, O., GUEX, J. and SAVARY, J. <http://folk.uio.no/ohammer/uagraph>, last access October 2007.
- JACKETT, S-J. and BAUMGARTNER, P.O., 2007. New imaging techniques applied to Paleogene Radiolaria. *Micropaleontology*, 53(3): 239–247.
- JIANBING, L. and AITCHISON, J. C. 2002. Upper Paleocene radiolarians from the Yamdrok melange, south Xizang (Tibet), China. *Micropaleontology*, 48 (1): 145–154.
- JOHNSON, D. A. and NIGRINI, C. A., 1985. Time-transgressive late Cenozoic Radiolarian events of the Equatorial Indo-Pacific. *Science*, 230 (4725): 53 8–540
- KELLER, G., ADATTE, T., HOLLIS, C. J., ORDONEZ, M., ZAMBRAZO, I., JIMENEZ, N., STINNESBECK, W., ALEMAN, A. and HALE-ELRICH, W., 1997. The Cretaceous/Tertiary boundary event in Ecuador; reduced biotic effects due to eastern boundary current setting. *Marine Micropaleontology* 31 (3-4): 97–133.
- KOZLOVA, G. E., 1999. Paleogene boreal radiolarians from Russia. Practical manual on microfauna, 9, St Petersburg, All-Russia Petroleum Research Exploration Institute (VNIGRI): 323 pp.
- KOZLOVA, G. E. and GORBOVETZ, A. N., 1966. Radiolarians of the Upper Cretaceous and Upper Eocene deposits of the West Siberian Lowland. *Proceedings of the All Union Petroleum Scientific Research Institute for Geological Survey (VNIGRI)*, 248: 1–159.
- KOZUR, H., and MOSTLER, H. 1982. Entactinaria: A new radiolarian suborder. *Geologisch-Paläontologische Mitteilungen Innsbruck*, 11 (1): 399–414.
- KRASHENINNIKOV, V. A., 1960. Nekotorye Radiolyarii Nizhnego i Srednego Eotsena Zapadnogo Predkavkaza. *Mineralogicko-Geologicka i Okhrana Nadr SSSR Vsesoyuznogo Nauchno-Issledovatel'skogo Geologorazved Neftyanogo Instituta*, 16: 271–308.
- LAZARUS, D.B., 1992. Antarctic Neogene radiolarians from the Kerguelan Plateau, Legs 119 and 120. In: Wise, S-W. Jr., Schlisch, R. et al. Eds., *Proceedings of the Ocean Drilling Program, Scientific Results 120*, 785–809. College Station, TX: Ocean Drilling Program.
- LING, H.-Y. 1991. Cretaceous (Maestrichtian) radiolarians. In: Ciesielski, P.F., Kristoffersen, Y. et al. Eds. *Proceedings of the Ocean Drilling Program, Scientific Results 114*, 317–324. College Station, TX: Ocean Drilling Program.
- MOORE, T.C. Jr., SHACKLETON, N.J. and PISIAS, N.G., 1993. Paleoceanography and the diachrony of radiolarian events in the eastern Equatorial Pacific. *Paleoceanography*, 8 (5): 567–586.
- MORLEY, J.J. and NIGRINI C., 1995. Miocene to Pleistocene radiolarian biostratigraphy of North Pacific sites 881, 884, 885, 886, and 887. In: Rea, D.K. Basov, I.A. et al., Eds., *Proceedings of the Ocean Drilling Program, Scientific Results 145*, 55–91. College Station, TX: Ocean Drilling Program.
- NIGRINI, C., 1967. Radiolaria in pelagic sediments from the Indian and Atlantic Oceans. *Bulletin of the Scripps Institution of Oceanography, University of California, San Diego*, 11: 125
- , 1977. Tropical Cenozoic Artostrobiidae (Radiolaria). *Micropaleontology*, 23 (3): 241–269.
- NIGRINI, C. and CAULET, J-P., 1992. Late Neogene radiolarian assemblages characteristic of Indo-Pacific areas of upwelling. *Micropaleontology* 38 (2): 139–164.
- NIGRINI, C., SANFILIPPO, A. and MOORE, T.J., Jr., 2006. Cenozoic radiolarian biostratigraphy: a magnetobiostratigraphic chronology of Cenozoic sequences from ODP Sites 1218, 1219, and 1220, Equatorial Pacific. In: Wilson, P. A., Lyle, M., Firth, J.V. et al. Eds., *Proceedings of the Ocean Drilling Program, Scientific Results 199*, Ocean Drilling Program, College Station, TX: 1–76.
- NISHIMURA, A., 1987. Cenozoic Radiolaria in the western North Atlantic, site 603, leg 93 of the Deep Sea Drilling Project. In: Van Hinte, J.E., et al., Eds., *Initial Reports of the Deep Sea Drilling*

- Project Leg 93, Washington, DC: US Government Printing Office, 713-737
- , 1992. Paleocene radiolarian biostratigraphy in the northwest Atlantic at Site 384, Leg 43, of the Deep Sea Drilling Project. *Micropaleontology*, 38(4): 317–362.
- NORRIS, R. D. and ROHL, U. 1999. Carbon cycling and chronology of climate warming during the Palaeocene/Eocene transition, *Nature*, 401: 775-778.
- NORRIS, R. D., KLAUS, A. and KROON, D. 2001. Mid-Eocene deep water, the Late Palaeocene Thermal Maximum and continental slope mass wasting during the Cretaceous-Palaeogene impact. In: Kroon, D., Norris, R.D., and Klaus, A. Eds., *Western North Atlantic Palaeogene and Cretaceous Palaeoceanography*. Special Publication of Geological Society, London, 183: 23-49.
- OKADA, H. and BUKRY, D. 1980. Supplementary modification and introduction of code numbers to the low latitude coccolith biostratigraphic zonation (Bukry 1973; 1975). *Marine Micropaleontology*, 5: 32 1-325.
- PINDELL, J., KENNAN, L., MARESCH, W.V., STANEK, K.-P., DRAPER, G., and HIGGS, R., 2005, Plate-kinematics and crustal dynamics of circum-Caribbean arc-continent interactions: Tectonic controls on basin development in Proto-Caribbean margins. In: Avé Lallement, H.G., and Sisson, V.B., Eds., *Caribbean-South American plate interactions, Venezuela*, 7–52. Geological Society of America Special Paper, 394.
- PETRUSHEVSKAYA, M. G. 1981. *Radiolyarii otryada Nassellaria Mirovogo Okeana*. Izdavaemye Zoologicheskim Institutom Akademii Nauk SSSR, 128: 406 pp.
- PETRUSHEVSKAYA, M. G. and KOZLOVA, G. E. 1972. Radiolaria: Leg 14, Deep Sea Drilling Project. In: Hayes, D. E., Pimm, A. C. et al. Eds., *Initial Reports of the Deep Sea Drilling Project, 14*. Washington, DC: US Government Printing Office, 495-648.
- RIEDEL, W.R., and SANFILIPPO, A., 1970. Radiolaria, Leg 4, Deep Sea Drilling Project. In Bader, R.G., Gerard, R.D., et al. Eds., *Initial Reports of the Deep Sea Drilling Project, 4*. Washington, DC: US Government Printing Office, 503-575.
- , 1971. Cenozoic Radiolaria from the western tropical Pacific, Leg 7. In: Winterer, E. L., Riedel, W. R. et al. Eds., *Initial Reports of the Deep Sea Drilling Project, 7*. Washington, DC: US Government Printing Office, 1529-1672.
- , 1973. Cenozoic Radiolaria from the Caribbean, Deep Sea Drilling Project, Leg 15. In; Edgar, N.T., Saunders, J.B., et al. Eds., *Initial Reports of the Deep Sea Drilling Project 15*. Washington, DC: US Government Printing Office, 705–751.
- SANFILIPPO, A. and BLOME, C.D., 2001. Biostratigraphic implications of mid-latitude Paleocene–Eocene radiolarian faunas from Hole 1051A, Ocean Drilling Program Leg 171B, Blake Nose, western North Atlantic. In: Kroon, D., Norris, R.D., and Klaus, A. Eds., *Western North Atlantic Palaeogene and Cretaceous Palaeoceanography*, 185-224. Geological Society of London Special Publication, 183.
- SANFILIPPO, A. and NIGRINI, C., 1998a. Code numbers for Cenozoic low latitude radiolarian biostratigraphic zones and GPTS conversion tables. *Marine Micropaleontology*, 33(1-2): 109–156
- , 1998b. Upper Paleocene-Lower Eocene deep-sea radiolarian stratigraphy and the Paleocene/Eocene Series boundary. In: Aubry, M.-P. et al., Eds., *Late Paleocene-Early Eocene climatic and biotic events in the marine and terrestrial records*, 244–276. New York: Columbia University Press.
- SANFILIPPO, A. and RIEDEL, W. R., 1973. Cenozoic Radiolaria (exclusive of theoperids, arstrobiids and amphydyacids) from the Gulf of Mexico, DSDP Leg 10. In: Worzel, J. L., Bryant, W., et al., Eds., *Initial Reports of the Deep Sea Drilling Project 10*, 475–611. Washington, DC: US Government Printing Office.
- , 1980. A revised generic and suprageneric classification of the Artiscins (Radiolaria). *Journal of Paleontology*, 54(5): 1008–1011.
- , 1982. Revision of the radiolarian genera *Theocotyle*, *Theocotylissa*, and *Thrysocyrtis*. *Micropaleontology*, 28: 170-188.
- SANFILIPPO, A., WESTBERG-SMITH, M. J., and RIEDEL, W. R. 1985. Cenozoic Radiolaria. In: Bolli, H. M., Saunders, J. B., and Perch-Nielsen, K., Eds., *Plankton stratigraphy*, 631-712. Cambridge: Cambridge University Press.
- SHILOV, V.V., 1995. Eocene-Oligocene radiolarians from Leg 145, North Pacific. In: Rea D.K., Basov, I.A. et al. Eds. *Proceedings of the Ocean Drilling Program, Scientific Results 145*, 117-132. College Station, TX: Ocean Drilling Program.
- SHIPBOARD SCIENTIFIC PARTY, 1973. Sites 86, 94, 95 and 96. In: Worzel, J. L., Bryant, W. et al., Eds., *Initial Reports of the Deep Sea Drilling Project, 10*. Washington, DC: US Government Printing Office.
- , 1998. Site 1051. In: Norris, R. D., Kroon, O., Klaus, A. et al. Eds., *Proceedings of the Ocean Drilling Program, Initial Reports, 17IB*, 171-239. College Station, TX: Ocean Drilling Program.
- STRONG C., HOLLIS C. J. and WILSON G., 1995. Foraminiferal, radiolarian, and dinoflagellates biostratigraphy of Late Cretaceous to middle Eocene pelagic sediments (Muzzle Group), Mead Stream, Marlborough, New Zealand. *New Zealand Journal of Geology and Geophysics* 38: 171–209.
- SUTTON, H. J., 1896. Radiolaria: a new genus from Barbados. *American Monthly Microscopical Journal*, 17(194): 61-62.
- TAKEMURA, A. 1986. Classification of Jurassic Nassellarians (Radiolaria). *Paläontographica Abteilung A: Paläozoologie-Stratigraphie* 195(1-3): 29-74.