

SHORT REPORT

A *de novo* nonsense *PDGFB* mutation causing idiopathic basal ganglia calcification with laryngeal dystonia

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Idiopathic basal ganglia calcification (IBGC) is characterized by brain calcification and a wide variety of neurologic and psychiatric symptoms. In families with autosomal dominant inheritance, three causative genes have been identified: *SLC20A2*, *PDGFRB*, and, very recently, *PDGFB*. Whereas in clinical practice sporadic presentation of IBGC is frequent, well-documented reports of true sporadic occurrence are rare. We report the case of a 20-year-old woman who presented laryngeal dystonia revealing IBGC. Her healthy parents' CT scans were both normal. We identified in the proband a new nonsense mutation in exon 4 of *PDGFB*, c.439C>T (p.Gln147*), which was absent from the parents' DNA. This mutation may result in a loss-of-function of PDGF-B, which has been shown to cause IBGC in humans and to disrupt the blood-brain barrier in mice, resulting in brain calcification. The c.439C>T mutation is located between two previously reported nonsense mutations, c.433C>T (p.Gln145*) and c.445C>T (p.Arg149*), on a region that could be a hot spot for *de novo* mutations. We present the first full demonstration of the *de novo* occurrence of an IBGC-causative mutation in a sporadic case.

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INTRODUCTION

Idiopathic basal ganglia calcification (IBGC) is defined by the presence of brain calcification affecting at least the basal ganglia, after the exclusion of known causes. There is wide intra- and inter-familial diversity of symptoms and ages of onset. The most common signs are cognitive impairment, psychiatric signs, movement disorders, gait disorder, dysarthria, cerebellar syndrome, pyramidal signs, and seizures.¹ Most familial cases are dominantly inherited. Three causative genes have recently been discovered. First, loss-of-function mutations in *SLC20A2*, causing IBGC in approximately 40% families,^{2,3} might have an impact on the inorganic phosphate metabolism in the brain. Then, we recently identified *PDGFRB*, which encodes the transmembrane receptor platelet derived growth factor receptor β (PDGFR β), as a second IBGC causative gene.⁴ Third, loss of function of PDGF-B (encoded by the *PDGFB* gene), the main ligand of PDGFR β , has recently been shown to cause IBGC in humans and brain calcification through a disruption of the blood–brain barrier integrity in mice.⁵ Sporadic presentation of IBGC is not uncommon. Nevertheless, true sporadic cases, defined by the absence of basal ganglia calcification in the parents' CT scans after the age of 50,⁶ were rarely reported.⁷ We report the case of a 20-year-old woman

presenting laryngeal dystonia revealing IBGC, due to a *de novo* nonsense mutation within *PDGFB*.

MATERIALS AND METHODS

The proband and her parents gave informed, written consent. The study was approved by our local ethics committee. Genomic DNA was extracted from whole peripheral blood using the Flexigen DNA kit (Qiagen, Hilden, Germany). In the proband's DNA, the entire coding sequence and intron/exon boundaries of *SLC20A2* (NM_006749.3), *PDGFRB* (NM_002609.3), and *PDGFB* (NM_002608.2) were PCR-amplified. PCR products were sequenced using the BigDye V3.1 Terminator Kit (Applied Biosystems, Courtaboeuf, France) on an automated sequencer (ABI 3100; Applied Biosystems). After the identification of the nonsense mutation in exon 4 of *PDGFB* (the exons were numbered according to the reference NM_002608.2 in this report; exon 4 corresponds to exon 5 using NG_012111.1 as reference) in the proband, we performed PCR amplification and sequencing of this exon in the parents.

To screen the presence of the mutation in controls, we used the 6503 exomes provided by the Exome Variant Server (EVS, NHLBI GO exome sequencing project, accessed in August, 2013), and a series of 173 in-house exomes from individuals with no neuropsychiatric disorder. To ascertain the parenthood, we used four informative microsatellites (D1S439, D9S1784, D14S986, and D19S913) and genotyped the proband and her parents using a multiplex fluorescent PCR.

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To search for nonsense-mediated RNA decay (NMD), we reverse transcribed total mRNA collected in PAXgene Blood RNA tubes (Qiagen) from the patient and her parents (Fisher Scientific, Illkirch, France) and performed PCR sequencing of the complementary DNA of Exon 4 of *PDGFB*. We then designed a reverse transcription-quantitative multiplex PCR of short fluorescent fragments (RT-QMPSF) of two amplicons of *PDGFB*, with *TOP1* and *SF3A1* genes as controls.

All primer sequences are reported in Supplementary Material. The mutation was submitted to the Leiden Open Variation Database (LOVD).

RESULTS

Case report

A 20-year-old woman was hospitalized for dysphagia and stridor. She had no familial medical history, and her personal medical history was marked by migraine without aura, leading to the identification of faint calcification of both lenticular nuclei in a CT scan performed at 10 years old during a migraine episode. Direct laryngoscopy showed bilateral vocal cord dysfunction: when the patient breathed in, the vocal cords moved in adduction. Three days later, she presented an acute exacerbation of dyspnea, with vocal cord paralysis in adduction, leading to oro-tracheal intubation. As it was not possible to deprive the patient from the ventilator, she underwent a tracheotomy. Neurological examination, laryngeal CT scan, and electromyography were all normal. No loco-regional cause was found. The cerebral CT scan showed calcification of both lenticular and caudate nuclei, and FLAIR-weighted magnetic resonance imaging only showed several punctiform hyperintensities in the supratentorial white matter and discrete anterior periventricular hyperintensities (Figure 1). An extensive etiological assessment of brain calcification (according to Nicolas *et al*⁴) was negative. We retained the diagnosis of IBGC. Her healthy parents had normal CT-scans, performed at the age of 52 years (mother) and 66 years (father). Muscle biopsy (performed as a search for mitochondrial disease) did not show any calcification.

Genetics

No potentially pathogenic variant was identified in *SLC20A2* and *PDGFRB* in the proband. We found a heterozygous single-nucleotide substitution in exon 4 of *PDGFB*, c.439C>T, predicted to introduce a premature stop codon at position 147 in the protein (p.Gln147*). This mutation was not retrieved in the parents and was absent in controls. Microsatellite analysis was compatible with paternity. We retrieved the mutation at the RNA level, r.439c>u (NM_002608.2). We did not find any decrease in the expression of the mutated allele in the proband by RT-PCR sequencing and RT-QMPSF (Supplementary Data). However, as no SNP was present in the *PDGFB*-coding sequence, we could not use a more sensitive approach, such as an SNaPshot technique, to detect NMD.

In addition, with the aim to find more mutations of *PDGFB*, we also sequenced the entire coding region and exon-intron boundaries of *PDGFB* in the 12 index cases from IBGC families and the 17 cases with sporadic presentation of IBGC with no mutation within *PDGFRB* and *SLC20A2* from our French case series.¹ We only found two mutations: the first one was already reported within the original article,⁵ and the second one is the currently presented mutation.

DISCUSSION

Our patient presented laryngeal dystonia, which is defined by involuntary contraction of the vocal cords, responsible for dysphonia.⁸ Severe laryngeal dystonia may also be responsible for life-threatening dyspnea. Laryngeal dystonia is a rare manifestation of IBGC⁹ and such a severe and acute revelation of IBGC has never been reported, to our knowledge.

IBGC is usually inherited according to an autosomal dominant pattern. As clinical expression of brain calcification is not constant, clinically sporadic presentation of IBGC may be due to mutations inherited from an asymptomatic parent in some cases.

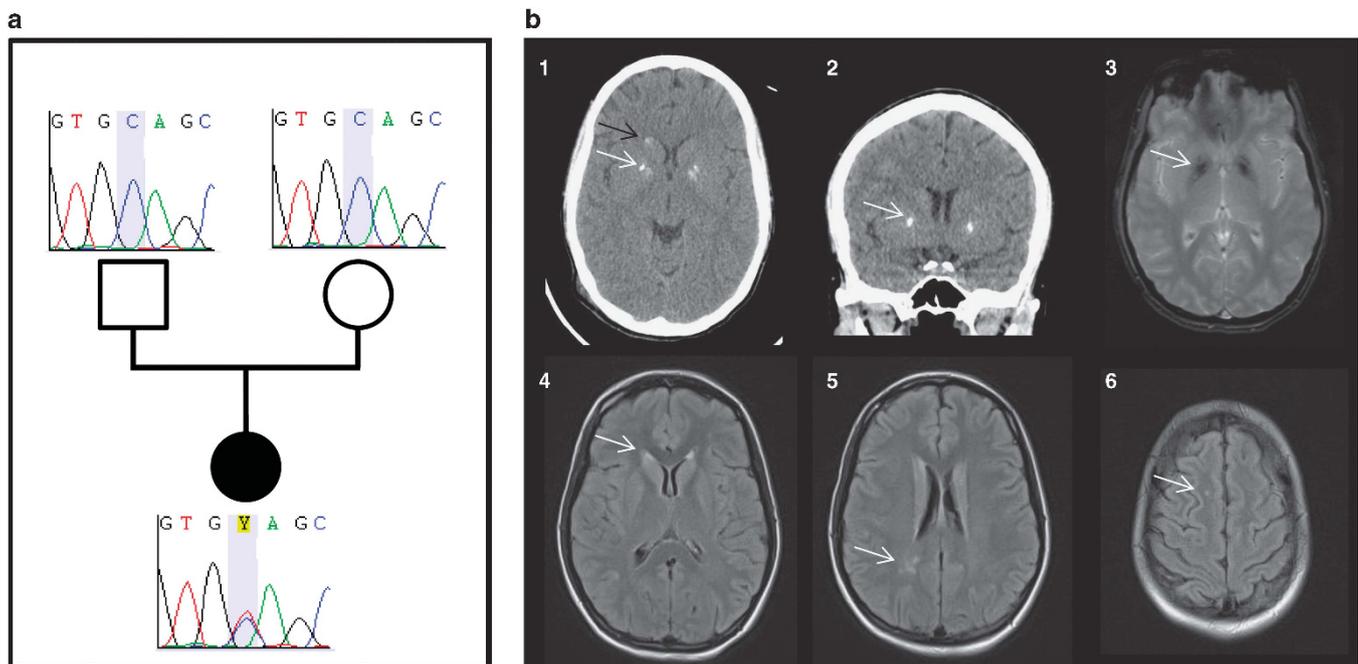


Figure 1 Electropherograms and brain imaging. Electropherograms show a heterozygous substitution of a C by a T in the proband, and a wild-type allele (C) in both parents (a). Brain CT scans on axial (b-1) and coronal (b-2) views show moderate calcifications in the lenticular nuclei (white arrows) and faint calcifications in the caudate nuclei (black arrow). On T2-weighted MRI, calcifications appear hypointense (b-3) while no signal modification is seen in the lenticular nuclei in FLAIR-weighted images, showing hyperintensities on anterior periventricular regions (b-4) and subcortical white matter (b-5, b-6).

We demonstrate here that a true sporadic occurrence of IBGC, due to a causative *de novo* mutation, can occur.

The mutation occurred at position c.439 (p.147), just between two recently reported nonsense mutations in *PDGFB* (resulting in p.Gln145* and p.Arg149*). Interestingly, the c.445C>T (p.Arg149*) mutation also most likely occurred *de novo*, although, to our knowledge, paternity was not verified.⁵ This suggests a clustering of *de novo* mutations in exon 4 of the *PDGFB* gene. Such a phenomenon has been studied using pangenomic data from concordant monozygotic twins with autism, and a mutability index (MI) was calculated for each exon as an estimate of relative mutation rate at single-nucleotide resolution.¹⁰ The mean MI is 2.41 for exon 4 of *PDGFB* (chr22:37,957,573_37,957,779, hg18), the genome-wide arithmetic mean of MI being 1, and the mean MI of all other exons of *PDGFB* being less than 1.54, except for exon 5 (2.15). In accordance with these estimates, our data suggest that exon 4 of *PDGFB* could be a hotspot for single-nucleotide substitutions. It is also well-recognized that an older paternal age is an extrinsic factor favoring *de novo* point mutations.¹⁰ Although the parental origin of the mutated allele could not be documented in our case, we note that the father was aged 44 and the mother 29 at their daughter's date of birth.

Surprisingly, we did not find further mutations of *PDGFB* with the exception of the previously reported one,⁵ among the familial cases and those with sporadic presentation of our case series. *De novo* occurrence of a *PDGFB* mutation is therefore probably a quite rare event among the IBGC cases. Nevertheless, among the seven mutations reported within *PDGFB* so far, two occurred *de novo*, which is a relevant result.

This c.439C>T (p.Gln147*) mutation might not be subject to efficient NMD because it is located close to the exon4–exon5 junction,¹¹ which could explain that we did not find mRNA decay. However, the mutation is clearly detrimental, as it is predicted to result in a shortened protein with loss of functionally critical domains including interface-contributing residues of the PDGF-B:PDGFR β complex.¹²

To conclude, we demonstrate true sporadic occurrence of IBGC due to a *de novo* nonsense mutation. Our results further support the involvement of the loss of function of PDGF-B in IBGC and suggest a possible hotspot for single-nucleotide substitutions in exon 4 of *PDGFB*.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

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