

RESEARCH ARTICLE

Genotype-Phenotype Associations of the CD-Associated Single Nucleotide Polymorphism within the Gene Locus Encoding Protein Tyrosine Phosphatase Non-Receptor Type 22 in Patients of the Swiss IBD Cohort

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Data Availability Statement: The data used for this study is obtained from the Swiss IBD cohort. According to Swiss ethic's law, data including patient data can only be accessed upon approval by an ethics board. Data collected in the Swiss IBD cohort are accessible upon ethical approval and a request to the Head of the cohort: SwissIBD Cohort Study, Prof. Dr. med. Dr. phil. Gerhard Rogler, Division of Gastroenterology and Hematology, Raemistrasse 100, 8091 Zurich, Switzerland. For further data requests

Abstract

Background

Protein tyrosine phosphatase non-receptor type 22 (PTPN22) plays an important role in immune cell function and intestinal homeostasis. The single nucleotide polymorphism (SNP) rs2476601 within the *PTPN22* gene locus results in aberrant function of PTPN22 protein and protects from Crohn's disease (CD). Here, we investigated associations of PTPN22 SNP rs2476601 in inflammatory bowel disease (IBD) patients in the Swiss IBD Cohort Study (SIBDCS).

Methods

2'028 SIBDCS patients (1173 CD and 855 ulcerative colitis (UC) patients) were included. The clinical characteristics were analysed for an association with the presence of the PTPN22 SNP rs2476601 genotypes 'homozygous variant' (AA), 'heterozygous' (GA) and 'homozygous wild-type' (GG).

Results

13 patients (0.6%) were homozygous variant (AA) for the PTPN22 polymorphism, 269 (13.3%) heterozygous variant (GA) and 1'746 (86.1%) homozygous wild-type (GG). In CD, AA and GA genotypes were associated with less use of steroids and antibiotics, and reduced prevalence of vitamin D and calcium deficiency. In UC the AA and GA genotype

please see: <http://ibdcohort.ch/index.php/informationen-fuer-forscher.html>.

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was associated with increased use of azathioprine and anti-TNF antibodies, but significantly less patients with the *PTPN22* variant featured malabsorption syndrome ($p = 0.026$).

Conclusion

Our study for the first time addressed how presence of SNP rs2476601 within the *PTPN22* gene affects clinical characteristics in IBD-patients. Several factors that correlate with more severe disease were found to be less common in CD patients carrying the A-allele, pointing towards a protective role for this variant in affected CD patients. In UC patients however, we found the opposite trend, suggesting a disease-promoting effect of the A-allele.

Introduction

A single nucleotide polymorphism (SNP) within the gene locus encoding protein tyrosine phosphatase non-receptor type 22 (*PTPN22*; SNP ID rs2476601) has been associated with an increased risk to develop autoimmune disorders, including rheumatoid arthritis (RA)[1–3], systemic lupus erythematosus (SLE)[4–6], Graves disease[7], and type-I diabetes (T1D)[7, 8]. Interestingly, genome-wide association studies (GWAS) that addressed genes associated with inflammatory bowel disease (IBD), revealed that the very same SNP reduces the risk to develop Crohn's disease (CD)[9–12]. While there was no association found with ulcerative colitis (UC) in most of these studies, one of them found a moderate decrease in UC disease risk, which was attributed to correlation with reduced TNF serum levels[9]. In contrast to classical autoimmune or auto-inflammatory disorders, where the adaptive immune system attacks the body's own cells/tissues, current hypothesis suggest that IBD is driven by inflammatory reactions against the harmless commensal microbiota in the intestine[13–15]. It has been suggested that genetic factors result in a defective innate immune response towards invading intestinal pathogens ultimately driving an over-activation of the adaptive arm of the immune system, what finally causes severe chronic and/or relapsing intestinal inflammation[13, 14, 16–18]. Although up to date over 200 gene loci have been associated with an altered risk to develop IBD[19], and for several of them, basic research has provided important mechanistic insight, it is still not known how presence of these SNPs affects clinical outcome and/or disease characteristics in IBD patients.

The CD-associated SNP rs2476601 is located in exon 14 of the *PTPN22* gene locus and results in the substitution of arginine 620 with a tryptophan residue in the *PTPN22* protein product (*PTPN22*-620W). Although initial studies demonstrated that presence of the variant results in increased *in vitro* dephosphorylation capacity[20], the *PTPN22*-620W variant is nowadays regarded to lead to an altered-function protein, since more recent studies demonstrated that mice designed to express the murine orthologue of *PTPN22*-620W, feature increased T cell receptor signaling and enhanced levels of autoreactive T cells, phenocopying the findings in *PTPN22* deficient animals[21, 22]. Later, these changes in T cell receptor signaling were attributed to altered substrate specificity of the *PTPN22*-620W variant[23].

PTPN22 is expressed in all immune cells, including B and T lymphocytes as well as myeloid immune cells such as monocytes, dendritic cells and macrophages[24], but not in non-hematopoietic cells such as intestinal epithelial cells or fibroblasts ([24] and own unpublished data). In T and B cells, *PTPN22* activity attenuates antigen receptor signaling[20, 22, 25], ultimately promoting proliferation and aberrant activation of T and B cells[25–27]. The function of *PTPN22* in innate immune cells is less studied, although it seems to be importantly involved in intestinal

homeostasis: we have found that PTPN22 is reduced in intestinal biopsies of IBD patients when compared to healthy subjects[28]. This reduction was mainly due to decreased expression of PTPN22 in CD68+ cells of the monocyte/macrophage lineage, while its expression in B and T cells remained unchanged[28]. Loss of PTPN22 in monocytes results in misbalanced secretion of inflammatory cytokines in response to IFN- γ and the bacterial cell wall product muramyl dipeptide, characterized by enhanced levels of IL-6 and IL-8, but decreased IL-12 and IFN- γ [28, 29]. Further, loss of PTPN22 and presence of PTPN22-620W in macrophages favors generation of pro-inflammatory M1 macrophages[30], and attenuates toll-like receptor (TLR)4 and TLR7 signaling, resulting in decreased Type-I interferon responses[31, 32]. The importance of PTPN22 in intestinal homeostasis is further demonstrated by the fact that loss of PTPN22 results in increased dextran sodium sulfate (DSS)-induced acute colitis[30, 31].

Taken together, these data describe an important role for PTPN22 in regulating inflammatory events in the intestine, but up to date, it has not been addressed how presence of the minor (A) allele influences clinical course or disease characteristics in affected patients. Therefore, here we aimed to address how SNP rs2476601 in *PTPN22* influences clinical parameters in patients suffering from IBD. Since SNP rs2476601 is differentially associated with IBD than with classical inflammatory disorders, we believe that this can give important insight to understand why SNP rs2476601 is negatively associated with CD. Further, a better understanding of the association between IBD risk loci and the complex pathophysiology of IBD might result in better prediction of the disease course and therefore might have an important impact on treatment decisions.

Using the patient collective of the Swiss IBD Cohort Study (SIBDCS), we investigated, whether presence of the CD-associated *PTPN22* variant, rs2476601 is associated with distinctive disease characteristics in Swiss IBD patients.

Results

Distribution of PTPN22 alleles in the SIBDCS CD and UC patient cohorts

We analysed a total of 2'028 IBD patients from the SIBDCS, consisting of 1173 (57.8%) CD and 855 (42.2%) UC patients. Of the entire set of patients, 13 patients (0.6%) featured the AA genotype of the *PTPN22* polymorphism, 269 (13.3%) carried the heterozygous form (GA) and 1'746 (86.1%) the homozygous wild-type (GG).

In the group of 1173 CD patients 1034 (88.2%) carried the homozygous wild-type allele (GG) of PTPN22, 136 (11.6%) the heterozygous form (GA) and 3 (0.3%) the AA genotype. In the UC group 712 (83.3%) carried the homozygous wild-type allele (GG), 133 (15.6%) the heterozygous form (GA) and 10 (1.2%) the AA genotype. The groups GA and AA were merged together to compare existence of the A-allele to its non-existence. When comparing the distribution of these genotypes between the CD and UC group, according to a chi-squared test, the distributions of these genotypes were significantly different ($p = 0.002$; Table 1).

Table 1. Distribution of genotypes in UC and CD patients.

Number (%)	GG	GA or AA	p-value (chi2)
Diagnosis			
Crohn (1173 patients)	1034 (59.22%)	139 (49.29%)	0.002
UC or IC (855 patients)*	712 (40.78%)	143 (50.71%)	

*There are 804 UC and 51 IC patients.

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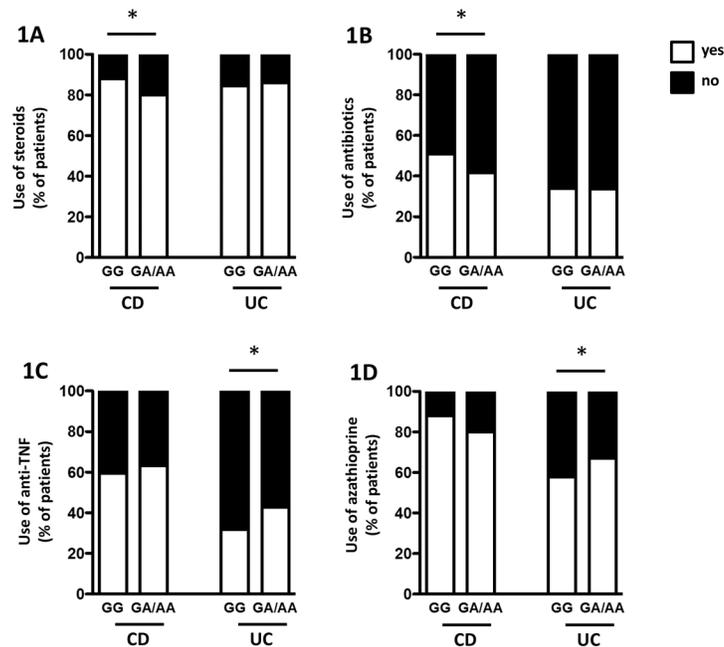


Fig 1. Use of steroids, antibiotics, anti-TNF treatment, and azathioprine in CD and UC patients carrying the A-allele in SNP rs2476601. The graphs show percentage of CD (left two bars in each graph) and UC patients (right two bars in each graph) treated (white area) or not treated (black area) with **A:** steroids, **B:** antibiotics, **C:** anti-TNF medication, **D:** or azathioprine.

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CD patients carrying the A-allele are treated less often with steroids or antibiotics

Disease course and response to a certain treatment are crucial clinical parameters to determine further treatment options, and they might be a factor to evaluate overall disease severity (e.g. anti-TNF antibodies are frequently used in patients refractory to other treatment approaches). Therefore, we next addressed whether presence of the A-allele was associated with specific medications and/or response to medication. In the CD group, the distribution of genotypes significantly differed between patients with steroid therapy (910 GG / 111 GA or AA) and those without steroid therapy (124 GG / 28 GA or AA), which was verified using the chi-squared test (p-value: 0.007; Fig 1A, S1 Table). This was also statistically significant when comparing the number of follow-ups with a therapy using steroids (p-value 0.034 by a chi-squared test; S1 Table). Also for the use of antibiotics the distribution of genotypes for those patients with antibiotic therapy (525 GG / 58 GA or AA) significantly differed from the CD patients not using antibiotics (509 GG / 81 GA or AA) with a p-value of 0.045 (Fig 1B; S1 Table). When analysing the use of anti-TNF antibodies, failure or non-response to anti-TNF therapy, non-response to steroids, use of azathioprine and/or 6-mercaptopurine, as well as use of methotrexate, cyclosporine and/or tacrolimus, no significant difference could be detected between the genotypes (GG versus GA/AA; Fig 1C+1D and S1 Table).

In UC patients use of anti-TNF antibodies and use of steroids is enhanced with the AA or GA genotype

In the UC group, the distribution of genotypes for patients with use of anti-TNF therapy (227 GG / 61 GA or AA) significantly differed from the UC patients not using anti-TNF therapy

(485 GG / 82 GA or AA) with a p-value of 0.013 by a chi-squared test (Fig 1C). The allele distribution in UC patients using azathioprine, but not in those using 6-mercaptopurine, also differed significantly (411 GG / 96 GA or AA versus 301 GG / 47 GA or AA; p-value 0.037; Fig 1D). This stayed significant when combining the use of azathioprine/6-mercaptopurine (437 GG / 100 GA or AA with AZA/6-MP versus 275 GG / 43 GA or AA without; p-value 0.049; S2 Table). When analysing the failure or non-response to anti-TNF therapy, use of steroids, number of follow-ups with a therapy of steroids, non-response to steroids, use of antibiotics, as well as use of methotrexate, cyclosporine and/or tacrolimus showed no significant difference when comparing genotypes (GG versus GA/AA; Fig 1 and S2 Table).

Presence of the A-allele is not associated with markers predicting complicated disease course

Since factors associated with a more severe disease course (e.g. IL-10R polymorphisms[33, 34], NOD2 variants[35, 36]) may also result in an earlier disease onset, we next analysed whether the age at first diagnosis is different in patients carrying the A-allele (AA and GA genotype) from those who do not (GG-genotype). In the CD group, median age at diagnosis was 24.6 years (q25-q75: 18.3–34.5; min-max: 0.5–81.4) in the GG group, and 25.4 years (q25-q75: 18.7–36.6; min-max: 6.5–73.7) in the GA or AA group, hence there was no statistically difference detectable (p = 0.28). Also in the UC group there was no significant difference in the age at diagnosis between the genotypes (GG versus GA/AA) with a median age at diagnosis of 29.0 years (q25-q75: 20.4–39.2; min-max: 3.1–79.6) within the GG genotype and 30.3 years (q25-q75: 23.0–38.5; min-max: 5.7–74.1) within the GA or AA genotype (p = 0.53; S3 Table; Fig 2A+2B). Next, we addressed demographic parameters and clinical phenotypes including gender, initial or current disease location, history of surgery, history of stenosis or fistulae and extra-intestinal manifestations in CD patients, but no significant differences between the genotypes were detected when using a chi-squared test (Table 2).

Association with malabsorption in UC patients and vitamin D and calcium deficiency in CD patients

We next analysed, whether PTPN22 variation might be associated with malabsorption and vitamin deficiency in IBD patients. In the UC group, malabsorption syndrome showed a statistically different distribution between the genotypes with 24 (3.37%) patients with

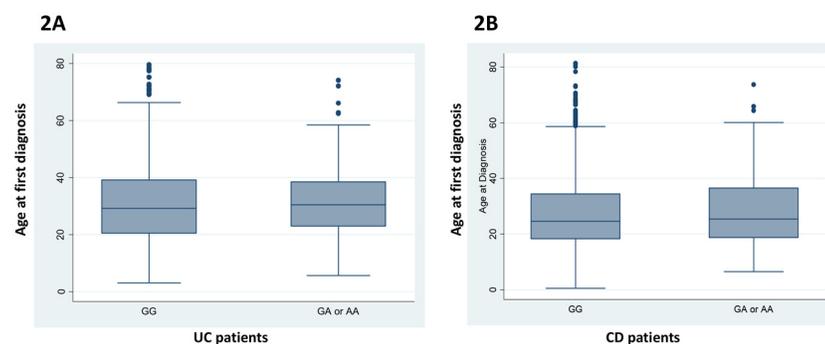


Fig 2. No difference in age at diagnosis in UC and CD patients carrying the A-allele in SNP rs2476601. The graphs show median age at first diagnosis (bold horizontal line), values within the 25% and 75% percentile (Box borders), and outliers (dots) in (A) UC, and (B) CD patients homozygous for the major (G) allele or heterozygous/homozygous carriers of the minor (A) allele in PTPN22 SNP rs2476601.

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Table 2. Association of PTPN22 rs2476601 SNP with clinical parameters in CD.

Number (%)	GG	GA or AA	p-value (chi2)
Gender distribution			
Men	524 (50.68%)	69 (49.64%)	0.818
Women	510 (49.32%)	70 (50.36%)	
Disease location			
Initial disease location			
L1	203 (21.50%)	29 (22.66%)	
L2	209 (22.14%)	27 (21.09%)	
L3	523 (55.40%)	69 (53.91%)	0.548
L4	9 (0.95%)	3 (2.34%)	
Unknown	90 missing	11 missing	
Current disease location			
L1	221 (23.14%)	33 (25.78%)	
L2	211 (22.09%)	28 (21.88%)	
L3	514 (53.82%)	64 (50.00%)	0.449
L4	9 (0.94%)	3 (2.34%)	
Unknown	79 missing	11 missing	
Fistula, stenosis, surgery			
Perianal fistula			
No	778(75.24%)	102 (73.38%)	0.634
Yes	256 (24.76%)	37 (26.62%)	
Other fistula			
No	816 (78.92%)	119 (85.61%)	0.065
Yes	218 (21.08%)	20 (14.39%)	
Anal fissure			
No	896 (86.65%)	126 (90.65%)	0.187
Yes	138 (13.35%)	13 (9.35%)	
Abscess			
No	753 (72.82%)	98 (70.50%)	0.565
Yes	281 (27.18%)	41 (29.50%)	
Summary of History of fistula			
No	525 (50.77%)	72 (51.80%)	0.820
Yes	509 (49.23%)	67 (48.20%)	
Stenosis			
No	576 (55.71%)	75 (53.96%)	0.697
Yes	458 (44.29%)	64 (46.04%)	
Surgery			
No	498 (48.16%)	66 (47.48%)	0.956
Yes	536 (51.84%)	73 (52.52%)	
Psoriasis and TBC			
Psoriasis			
No	923 (89.26%)	18 (12.7)	0.836
Yes	111 (10.74%)	124 (87.3)	
TBC infection			
No	924 (89.36%)	18 (12.7)	0.810
Yes	110 (10.64%)	124 (87.3)	
Summary of Psoriasis and TBC infection			
No	925 (89.46%)	18 (12.7)	0.784
Yes	109 (10.54%)	124 (87.3)	

(Continued)

Table 2. (Continued)

Number (%)	GG	GA or AA	p-value (chi2)
Complications			
Osteopenia (–porosis)			
No	777 (75.15%)	114 (82.01%)	0.075
Yes	257 (24.85%)	25 (17.99%)	
Deep Venous Thrombosis			
No	1010 (97.68%)	135 (97.12%)	0.686
Yes	24 (2.32%)	4 (2.88%)	
Pulmonary Embolism			
No	1019 (98.55%)	138 (99.28%)	0.485
Yes	15 (1.45%)	1 (0.72%)	
Gallstone			
No	973 (94.10%)	132 (94.96%)	0.683
Yes	61 (5.90%)	7 (5.04%)	
Nephrolithiasis			
No	981 (94.87%)	136 (97.84%)	0.123
Yes	53 (5.13%)	3 (2.16%)	
Malabsorption syndrome			
No	956 (92.46%)	130 (93.53%)	0.652
Yes	78 (7.54%)	9 (6.47%)	
Perforation / Peritonitis			
No	989 (95.65%)	131 (94.24%)	0.455
Yes	45 (4.35%)	8 (5.76%)	
Adv. Effect of Treatment			
No	899 (86.94%)	122 (87.77%)	0.785
Yes	135 (13.06%)	17 (12.23%)	
Summary of Complications			
No	590 (57.06%)	82 (58.99%)	0.665
Yes	444 (42.94%)	57 (41.01%)	
Extraintestinal manifestations			
Peripheral arthritis			
No	514 (49.71%)	71 (51.08%)	0.908
Yes	520 (50.29%)	68 (48.92%)	
Uveitis / Iritis			
No	900 (87.04%)	125 (89.93%)	0.314
Yes	134 (12.96%)	14 (10.07%)	
Pyoderma gangrenosum			
No	1019 (98.55%)	137 (98.56%)	0.987
Yes	15 (1.45%)	2 (1.44%)	
Erythema nodosum			
No	958 (92.65%)	127 (91.37%)	0.600
Yes	76 (7.35%)	12 (8.63%)	
Aphthous oral ulcers			
No	877 (84.82%)	124 (89.21%)	0.245
Yes	157 (15.18%)	15 (10.79%)	
Ankylosing spondylitis			
No	942 (91.10%)	127 (91.37%)	0.871
Yes	92 (8.90%)	12 (8.63%)	

(Continued)

Table 2. (Continued)

Number (%)	GG	GA or AA	p-value (chi2)
Prim. scler. cholangitis			
No	1026 (99.23%)	136 (97.84%)	0.113
Yes	8 (0.77%)	3 (2.16%)	
Other			
No	974 (94.20%)	133 (95.68%)	0.469
Yes	60 (5.80%)	6 (4.32%)	
Summary of Extraintestinal manifestations			
No	412 (39.85%)	58 (41.73%)	0.793
Yes	622 (60.15%)	81 (58.27%)	

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malabsorption syndrome in the GG group compared to 0% in the GA or AA group ($P = 0.026$; Fig 3A and S3 Table). Malabsorption is neither well defined in the Swiss IBD cohort nor in the gastroenterological literature in general, and its occurrence in UC patients is rather uncommon. Further, deficiency of vitamin K and vitamin D are associated with active intestinal inflammation [37, 38]. Analysis of these factors revealed that CD patients carrying the A-allele suffered less often from vitamin D and calcium deficiency (Fig 3B+3C and Table 3). In UC patients, however, presence of the A-allele was associated with increased occurrence of vitamin D deficiency (Fig 3B and Table 4), while for none of the other analyzed factors the distribution was significantly different between the genotypes (Tables 3+4).

Discussion

In our study, we analyzed whether presence of SNP rs2476601 within *PTPN22* is associated with disease characteristics in patients suffering from IBD. Using the longitudinal and prospectively obtained data from the SIBDC, we found that CD patients carrying the A-allele need less often steroids and/or antibiotic treatment, while no difference was detected regarding the use of anti-TNF antibodies. CD patients with the GA or AA genotype further suffer less often from vitamin D and calcium deficiency. In UC patients, presence of the A-allele was associated with enhanced use of anti-TNF medication and reduced prevalence of malabsorption syndrome, but at the same time—and in line with more severe disease—vitamin D deficiency was more common in those patients. In our study population, however, no significant difference could be found between genotypes when analysing other markers of (severe) disease, such as gender,

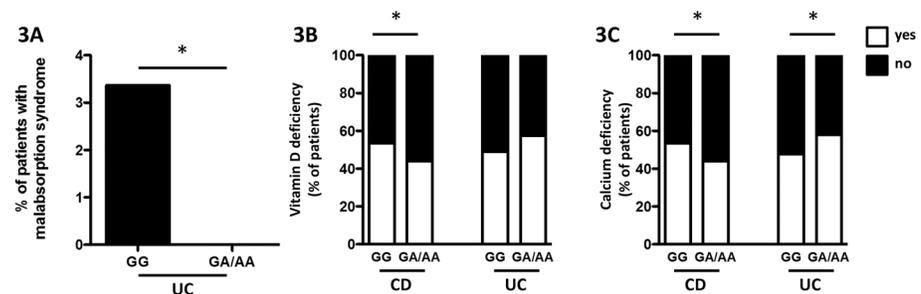


Fig 3. Presence of SNP rs2476601 affects malabsorption, vitamin D and calcium deficiency. A: percentage of UC patients with malabsorption syndrome. **B+C:** percentage of CD (left two bars) and UC patients (right two bars) featuring (white area) or not featuring (black area) **B:** vitamin D deficiency, or **C:** calcium deficiency.

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Table 3. Association of PTPN22 rs2476601 SNP with BMI and micronutrient deficiency in CD patients.

Crohn's disease patients	GG	GA or AA	p-value (Wilcoxon)
BMI			
Median (q25–q75, Min–Max)	22.7 (20.2–25.8, 12.5–48.1)	22.3 (19.5–25.9, 15.4–37.0)	0.424
Unknown: 11	9 missing	2 missing	
Hb level			
Median (q25–q75, Min–Max)	136.7 (128.2–145.8, 83–179)	138.2 (129–147, 97.5–168)	0.428
Unknown: 6	4 missing	2 missing	
Vit. B12 deficiency			
No	684 (76.68%)	88 (76.52%)	
Yes	208 (23.32%)	27 (23.48%)	0.970
Unknown: 166	142 missing	24 missing	
Folate deficiency			
No	499 (64.14%)	64 (68.82%)	
Yes	279 (35.86%)	29 (31.18%)	0.372
Unknown: 302	256 missing	46 missing	
Anaemia			
No	625 (60.68%)	89 (64.96%)	
Yes	405 (39.32%)	48 (35.04%)	0.334
Unknown: 6	4 missing	2 missing	
Iron def. Anaemia			
No	787 (84.26%)	106 (86.89%)	
Yes	147 (15.74%)	16 (13.11%)	0.451
Unknown: 117	100 missing	17 missing	
Iron def. + chronic disease anemia			
No	920 (98.50%)	122 (100%)	
Yes	14 (1.50%)	0 (0%)	0.173
Unknown: 117	100 missing	17 missing	
Calcium			
No	523 (50.58%)	83 (59.71%)	
Yes	511 (49.42%)	56 (40.29%)	0.043
Unknown: 0			
Vitamin D			
No	480 (46.42%)	78 (56.12%)	
Yes	554 (53.58%)	61 (43.88%)	0.032
Unknown: 0			
Folic acid			
No	733 (70.89%)	101 (72.66%)	
Yes	301 (29.11%)	38 (27.34%)	0.665
Unknown: 0			

doi:10.1371/journal.pone.0160215.t003

initial or current disease location, surgery, history of stenosis or fistula and extra-intestinal manifestations.

GWAS previously associated SNP rs2476601 with reduced risk for developing CD, since this variant is less prevalent in CD patients than in the normal population. In the here presented study, we expanded this knowledge to affected IBD patients, where we found that even in patients suffering from CD, SNP rs2476601 seems to have some protective effects: steroids are usually used in more severe disease, and the use of antibiotics typically results from complications and/or infections, hence reduced use of these two medications indicate that the

Table 4. Association of PTPN22 rs2476601 SNP with BMI and micronutrient deficiency in UC patients.

UC / IC patients	GG	GA or AA	p-value (Wilcoxon)
BMI			
Median (q25 –q75, Min–Max)	23.2 (20.9–26.2, 14.3–45.9)	22.9 (21.0–25.9, 13.6–48.8)	0.758
Unknown: 8	6 missing	2 missing	
Hb level			
Median (q25 –q75, Min–Max)	136 (127.5–145.7, 70–175)	137 (127.6–147.3, 86.5–168)	0.337
Unknown: 22	20 missing	2 missing	
Vit. B12 deficiency			
No	463 (85.42%)	84 (80.00%)	0.159
Yes	79 (14.58%)	21 (20.00%)	
Unknown: 208	170 missing	38 missing	
Folate deficiency			
No	328 (68.19%)	59 (62.77%)	0.305
Yes	153 (31.81%)	35 (37.23%)	
Unknown: 280	231 missing	49 missing	
Anaemia			
No	406 (58.67%)	87 (61.70%)	0.504
Yes	286 (41.33%)	54 (38.30%)	
Unknown: 22	20 missing	2 missing	
Iron def. Anaemia			
No	505 (83.33%)	108 (87.10%)	0.298
Yes	101 (16.67%)	16 (12.90%)	
Unknown: 125	106 missing	19 missing	
Iron def. + chronic disease anemia			
No	581 (95.87%)	116 (93.55%)	0.256
Yes	25 (4.13%)	8 (6.45%)	
Unknown: 125	106 missing	19 missing	
Calcium			
No	373 (52.39%)	60 (41.96%)	0.023
Yes	339 (47.61%)	83 (58.04%)	
Unknown: 0			
Vitamin D			
No	364 (51.12%)	61 (42.66%)	0.065
Yes	348 (48.88%)	82 (57.34%)	
Unknown: 0			
Folic acid			
No	596 (83.71%)	119 (83.22%)	0.885
Yes	116 (16.29%)	24 (16.78%)	
Unknown: 0			

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existence of the A-allele might protect from (severe and/or complicated) CD or might lead to a milder/less complicated disease course. Nevertheless, mechanistic data directly supporting these findings are lacking, hence our conclusion regarding the influence on disease severity should be regarded with caution. Since vitamin D deficiency is associated with active disease and a more severe disease course, the reduced abundance of vitamin D deficiency in CD patients with the GA or AA genotype further supports our hypothesis that the A-allele might be protective in CD. From basic research, it is not obvious why presence of SNP rs2476601 would result in reduced disease severity in CD. Most studies demonstrated that presence of the

A-allele results in changes in T-cell responses, ultimately promoting inflammatory T cell subsets[23, 25, 39], and during innate immune reactions, presence of the A-allele has been shown to promote inflammatory macrophages[30], also indicating an inflammation-prone phenotype. However, in contrast to other inflammatory disorders, in the intestine rapid clearance of invading pathogens is crucial for homeostasis, and it might well be that the more inflammation prone nature of the (first) immune response in A-allele carriers might result in a faster clearance of infections in an early stage of the disease, preventing the development of more severe infections needing antibiotic treatment, as well as the development of progressed chronic inflammation.

Of special interest are the findings in the UC patient group: even though in most GWAS no association of *PTPN22* SNP rs2476601 with UC was found[11, 12], and one study in a Danish IBD cohort even found reduced risk to develop UC upon presence of SNP rs2476601[9], our study in contrast suggests that the A-allele might have a disease-promoting role. In contrary to the before mentioned studies, we addressed clinical associations, i.e. how the variants influences disease course in IBD, rather than the risk to develop the disease in a first place, which might explain these opposed observations. In particular, we found that UC patients with the GA or AA genotype needed anti-TNF medication more often than patients with the GG genotype. Since anti-TNF medication is usually used in more severe, treatment refractory disease, this might be an indication that presence of the A-allele possibly results in a more pronounced disease course in UC patients. Direct mechanistic data to support this finding are lacking, but some data, describing how *PTPN22* affects cellular pathways involved in IBD, have been published recently[28, 29, 40] and are reviewed elsewhere[41]. There was no difference for the use of 6-mercaptopurine (6-MP), but the use of azathioprine (AZA) was significantly enhanced in UC patients carrying the A-allele, what again indicates a promoting role for the variant. Taken together this suggests that presence of the A-allele might have relevance not only for CD but also for UC patients. However, presence of the A-allele was not associated with altered response neither to the use of anti-TNF medication, nor the use of antibiotics nor steroids. This is consistent with previous findings in the above-mentioned Danish cohort, where the A-allele was also not found to be associated with changes in the response to anti-TNF treatment [9].

It might be surprising that the same genetic variant shows opposite effects on disease severity in CD and UC. The evidence pointing towards enhanced disease severity in UC is well in line with the A-allele being associated with increased risk for other autoimmune disorders. On the other hand, it is not surprising that the A-allele has a protective effect in CD, since GWAS have associated this allele with reduced risk to develop CD.

From a mechanistic point of view, the opposite findings on CD and UC disease characteristics might be explained by the fact that the *PTPN22* variant affects T cell biology[22, 25], as well as pro-inflammatory signalling in tissue macrophages [30]. It is clear that the role of T cell biology is different between UC and CD, with UC classically being regarded as a Th2-mediated disorder, while in CD Th1-signature cytokines play a dominant role[42]. Therefore, changes in T cell biology, as induced by presence of the *PTPN22* variant, likely have different effects on UC and CD development.

Malabsorption was significantly less often found in UC patients carrying the A-allele. Malabsorption in IBD patients is mainly caused through the presence of severe inflammation of the ileum and subsequent insufficient nutrient absorption as well as previous intestinal resection. In UC, the small intestine is typically not affected, however, malabsorption is known to occur in patients with high numbers of bowel movements thus indicating more severe disease. Micronutrition deficiency is common in IBD patients, although less prevalent in UC patients than in CD patients[43, 44]. Since micronutrition deficiencies are associated with severe disease course[43, 45], the finding that less UC patients carrying the A-allele show malabsorption,

somehow contradicts severe disease in those patients. However, UC patients carrying the A-allele showed calcium deficiency more often, and no other factor associated with malabsorption was affected. Taken together this again supports the hypothesis that UC patients carrying the A-allele might suffer from more severe disease.

CD patients with the GA or the AA genotype suffered less often from vitamin D and calcium deficiencies. This is of interest, since vitamin D deficiency is known as a risk factor for IBD, and is associated with active disease[37, 46]. Animal studies have further shown that vitamin D and vitamin D receptor (VDR) are important regulators of immune homeostasis: vitamin D reduces the proliferation of CD8+ cytotoxic T cells [47], and shifts the T helper cell balance away from (pro-inflammatory) Th1 and Th17 cells towards IL-10 producing Th2 and regulatory T cells[48, 49]. Further, vitamin D influences several pathways involved in IBD pathogenesis, such as NOD2 signalling and autophagy[48, 50]. Therefore, a positive influence on vitamin D levels upon presence of the A-allele might also contribute to a less severe disease course.

A drawback of our study might be that we only addressed one single SNP, and did not take other genetic variants in account that might be present in some of the patients. Of special interest in this regard is the fact that, aside the here addressed SNP rs2476601, another variant in the gene locus encoding PTPN22 (SNP rs33996649) has been described to affect susceptibility for IBD. This variant results in a loss of PTPN22 phosphatase function, and has been described to protect from the onset of UC[12]. Unfortunately the patients enrolled in the Swiss IBD cohort have not been genotyped for SNP rs33996649, therefore analysing phenotype changes associated with this variant was not possible.

Given the number of IBD-associated SNPs, it is likely that a significant number of patients might be carrying not only one, but several disease-associated SNPs. Since presence of several SNPs might have cumulative or even multiplying effects on clinical outcome, it would be of great interest to stratify patients carrying the PTPN22 SNP rs2476601 according to the presence of other genetic variants. However, since SNP rs2476601 is rather rare, there are not enough A-allele carriers within the Swiss IBD cohort to draw meaningful conclusions from such analysis.

A limitation of our study might be that we did not include healthy subjects, especially since the occurrence of SNP rs2476601 is rather low with only 0.6% in CD patients. However, in the healthy population, the SNP is more frequent (between 1–2%), and genetic variance is rather low in a small country such as Switzerland. The main focus of our study was to determine how presence of SNP rs2476601 affects disease characteristics in IBD patients; hence including healthy controls would not add significant value to achieve this aim. Further, it has already been described thoroughly that SNP rs2476601 is associated with IBD[9, 11, 12]. For these reasons, we refrained from including healthy controls in our study.

Despite these limiting factors, we can conclude that in summary, significantly fewer patients in our cohort with the PTPN22-620W variant (GA or AA genotype) were treated with steroids and antibiotics in CD, but more with azathioprine and anti-TNF antibodies in UC. Although no disease-promoting association of the PTPN22 SNP rs2476601 with UC was described before, we demonstrated, that significantly fewer UC patients carrying the variant developed malabsorption syndrome, but vitamin D and calcium deficiency was more common. These findings might suggest a milder disease course of CD but aggravated disease in UC in A-allele carriers. This opposite influence of the A-allele on CD and UC disease development supports the hypothesis that these two forms of IBD are distinct disease entities. Since PTPN22 is involved in immune cell regulation, our findings are in line with previous findings showing that UC and CD are distinct in their immunological signature[42]. Our findings are of interest, since presence of the A-allele in PTPN22 SNP rs2476601 is associated with several autoimmune

disorders, but, to the best of our knowledge, it is currently not known how the A-allele influences disease course or treatment characteristics in any of these disorders. Therefore, our study is the first to address a clinical relevance of SNP rs2476601, and helps to better understand its effect on disease course and treatment options in IBD patients.

Materials and Methods

Study Design

Patient data were obtained from the register of the nationwide SIBDCS, in which patients with IBD from all regions of Switzerland have prospectively been included since 2006[51]. The cohort study is supported by the Swiss National Science Foundation. The cohort goals and methodology are described elsewhere[51].

We included 2028 IBD patients that were enrolled in the study at time of data acquisition and had been previously genotyped for the CD-associated risk variant rs2476601 within the *PTPN22* gene locus. Genotyping was performed as part of an analysis of the whole Swiss IBD cohort for all SNPs that are currently known to be associated with IBD. Since UC and IC share several disease characteristics and indeterminate colitis is often managed the same as patients who have UC, UC and IC patients were pooled for the analysis in order to increase sample size. The *PTPN22* polymorphism rs2476601 occurs in three possible isoforms: homozygous wild-type (GG), heterozygous (GA), and homozygous variant (AA). The goal of this study was to analyze whether the presence of the GA- or AA-form is associated with clinical characteristics of IBD patients.

Clinical phenotypes of CD were classified regarding disease location, which was stratified into 1 of 4 groups according to the Montreal classification and analyzed separately for initial location and current location: ileal disease with or without disease limitation to the cecum (L1), a disease limited to the colon (L2), an ileal disease with disease of the colon beyond the cecum (L3), or disease of the upper gastrointestinal tract (L4). Patients with fistulae were classified into four groups: perianal fistula, other type fistula (non-perianal fistula), multiple fistulae (>1) and any type fistula. Presence of any intestinal stenosis was included in the analysis as positive for stenosis. Location of UC was classified according to the Montreal classification into proctitis (L1), left-sided colitis (L2), pancolitis (L3) or “location unknown”[52]. We also included history of intestinal surgery. Gender, age at diagnosis, smoking history, and presence of extraintestinal manifestations were taken into account. We further obtained data about current and prior treatment with 5-aminosalicylate, antibiotics, steroids, immunosuppressants (namely azathioprine/6-mercaptopurine), calcineurin inhibitors (tacrolimus, cyclosporine), and anti-TNF drugs (infliximab, adalimumab, and certolizumab) at enrollment or according to the term “ever treated with”. Anti-TNF non-response was defined as one of the following: (1) breakthrough / loss of response, (2) primary non-response (never effective), (3) therapy stop due to side effects / intolerance.

We further analysed whether A-allele carriers suffer from micronutrient deficiencies, such as iron, vitamin B12, vitamin D, calcium, or folate deficiency, which might all result from defective absorption due to severe inflammation. A further consequence of malabsorption would be a decreased body-mass-index and decreased Hb levels. Malnutrition and micronutrient deficiencies are common in IBD patients[43, 44], and are associated with more severe disease[43, 45] and longer hospitalization times[53].

Statistical Analysis

Clinical data were retrieved from the data center of the Swiss IBD Cohort Study at the University of Lausanne. These data and additional data obtained from a review of the patients' files

were entered into a database (Access 2000; Microsoft Switzerland Ltd Liab. Co., Wallisellen, Switzerland). The Statistical Package for the Social Sciences (version 21; SPSS, Chicago, IL) was used for the statistical analysis.

Crude differences about the association of the *PTPN22* variant in relation to fistulae, stenosis, smoking status, disease location, age at diagnosis, medications and history of intestinal resection surgery were assessed using the Pearson's [chi]² test or the Fisher's exact test (Fisher's exact test used if strata comprised a sample size ≤ 5). A multiple logistic regression model was calculated to identify the associations for this gene variant. Differences about the association of the *PTPN22* variant in relation to age at diagnosis were assessed using a Wilcoxon rank-sum test. A p-value smaller than 0.05 was considered significant.

Ethical considerations

The Swiss IBD cohort study is approved by the local ethical committees (IRB approval number: EK-1316, approved on 05.02.2007 by the Cantonal Ethics Committee of the Canton Zürich, Switzerland). Written informed consent was obtained before inclusion in the cohort.

Supporting Information

S1 Table. Association of PTPN22 rs2476601 SNP with treatment characteristics of CD.
(DOCX)

S2 Table. Association of PTPN22 rs2476601 SNP with treatment characteristics of UC.
(DOCX)

S3 Table. Association of PTPN22 rs2476601 SNP with age at diagnosis.
(DOCX)

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References

1. Begovich AB, Carlton VE, Honigberg LA, Schrodin SJ, Chokkalingam AP, Alexander HC, et al. A missense single-nucleotide polymorphism in a gene encoding a protein tyrosine phosphatase (PTPN22) is associated with rheumatoid arthritis. *Am J Hum Genet.* 2004 Aug; 75(2):330–7. PMID: [15208781](#)
2. Dieude P, Teixeira VH, Pierlot C, Cornelis F, Petit-Teixeira E, Ecras. Testing for linkage and association with rheumatoid arthritis a ptpn22 promoter polymorphism reported to be associated and linked with type 1 diabetes in the Caucasian population. *Ann Rheum Dis.* 2008 Jun; 67(6):900–1. doi: [10.1136/ard.2007.077180](#) PMID: [18474664](#)
3. Lt Michou, Lasbleiz S, Rat A-C, Migliorini P, Balsa A, Westhovens R, et al. Linkage proof for PTPN22, a rheumatoid arthritis susceptibility gene and a human autoimmunity gene. *Proceedings of the National Academy of Sciences.* 2007 January 30, 2007; 104(5):1649–54.
4. Ramirez M, Quintana G, Diaz-Gallo LM, Caminos J, Garcés M, Cepeda L, et al. The PTPN22 C1858T variant as a risk factor for rheumatoid arthritis and systemic lupus erythematosus but not for systemic sclerosis in the Colombian population. *Clin Exp Rheumatol.* 2012 Jul-Aug; 30(4):520–4. PMID: [22704547](#)
5. Moez P, Soliman E. Association of PTPN22 gene polymorphism and systemic lupus erythematosus in a cohort of Egyptian patients: impact on clinical and laboratory results. *Rheumatol Int.* 2012 Sep; 32(9):2753–8. doi: [10.1007/s00296-011-2063-z](#) PMID: [21818561](#)
6. Eliopoulos E, Zervou MI, Andreou A, Dimopoulou K, Cosmidis N, Voloudakis G, et al. Association of the PTPN22 R620W polymorphism with increased risk for SLE in the genetically homogeneous population of Crete. *Lupus.* 2011 Apr; 20(5):501–6. doi: [10.1177/0961203310392423](#) PMID: [21543514](#)
7. Zhebrun D, Kudryashova Y, Babenko A, Maslyansky A, Kunitskaya N, Popcova D, et al. Association of PTPN22 1858T/T genotype with type 1 diabetes, Graves' disease but not with rheumatoid arthritis in Russian population. *Aging (Albany NY).* 2011 Apr; 3(4):368–73.
8. Bottini N, Musumeci L, Alonso A, Rahmouni S, Nika K, Rostamkhani M, et al. A functional variant of lymphoid tyrosine phosphatase is associated with type I diabetes. *Nat Genet.* 2004 Apr; 36(4):337–8. PMID: [15004560](#)
9. Bank S, Skytt Andersen P, Burisch J, Pedersen N, Roug S, Galsgaard J, et al. Polymorphisms in the inflammatory pathway genes TLR2, TLR4, TLR9, LY96, NFKBIA, NFKB1, TNFA, TNFRSF1A, IL6R, IL10, IL23R, PTPN22, and PPARG are associated with susceptibility of inflammatory bowel disease in a Danish cohort. *PLoS One.* 2014; 9(6):e98815. doi: [10.1371/journal.pone.0098815](#) PMID: [24971461](#)

10. Waterman M, Xu W, Stempak JM, Milgrom R, Bernstein CN, Griffiths AM, et al. Distinct and overlapping genetic loci in Crohn's disease and ulcerative colitis: correlations with pathogenesis. *Inflamm Bowel Dis*. 2011 Sep; 17(9):1936–42. doi: [10.1002/ibd.21579](https://doi.org/10.1002/ibd.21579) PMID: [21830272](https://pubmed.ncbi.nlm.nih.gov/21830272/)
11. Genome-wide association study of 14,000 cases of seven common diseases and 3,000 shared controls. *Nature*. 2007 Jun 7; 447(7145):661–78. PMID: [17554300](https://pubmed.ncbi.nlm.nih.gov/17554300/)
12. Diaz-Gallo LM, Espino-Paisan L, Fransen K, Gomez-Garcia M, van Sommeren S, Cardena C, et al. Differential association of two PTPN22 coding variants with Crohn's disease and ulcerative colitis. *Inflamm Bowel Dis*. 2011 Nov; 17(11):2287–94. doi: [10.1002/ibd.21630](https://doi.org/10.1002/ibd.21630) PMID: [21287672](https://pubmed.ncbi.nlm.nih.gov/21287672/)
13. de Souza HS, Fiocchi C. Immunopathogenesis of IBD: current state of the art. *Nat Rev Gastroenterol Hepatol*. 2015 Dec 2.
14. Abraham C, Medzhitov R. Interactions between the host innate immune system and microbes in inflammatory bowel disease. *Gastroenterology*. 2011 May; 140(6):1729–37. doi: [10.1053/j.gastro.2011.02.012](https://doi.org/10.1053/j.gastro.2011.02.012) PMID: [21530739](https://pubmed.ncbi.nlm.nih.gov/21530739/)
15. Li YR, Zhao SD, Li J, Bradfield JP, Mohebnasab M, Steel L, et al. Genetic sharing and heritability of paediatric age of onset autoimmune diseases. *Nat Commun*. 2015; 6:8442. doi: [10.1038/ncomms9442](https://doi.org/10.1038/ncomms9442) PMID: [26450413](https://pubmed.ncbi.nlm.nih.gov/26450413/)
16. Maloy KJ, Powrie F. Intestinal homeostasis and its breakdown in inflammatory bowel disease. *Nature*. 2011 Jun 16; 474(7351):298–306. doi: [10.1038/nature10208](https://doi.org/10.1038/nature10208) PMID: [21677746](https://pubmed.ncbi.nlm.nih.gov/21677746/)
17. Kaser A, Zeissig S, Blumberg RS. Inflammatory bowel disease. *Annu Rev Immunol*. 2010; 28:573–621. doi: [10.1146/annurev-immunol-030409-101225](https://doi.org/10.1146/annurev-immunol-030409-101225) PMID: [20192811](https://pubmed.ncbi.nlm.nih.gov/20192811/)
18. Khor B, Gardet A, Xavier RJ. Genetics and pathogenesis of inflammatory bowel disease. *Nature*. 2011 Jun 16; 474(7351):307–17. doi: [10.1038/nature10209](https://doi.org/10.1038/nature10209) PMID: [21677747](https://pubmed.ncbi.nlm.nih.gov/21677747/)
19. Liu JZ, van Sommeren S, Huang H, Ng SC, Alberts R, Takahashi A, et al. Association analyses identify 38 susceptibility loci for inflammatory bowel disease and highlight shared genetic risk across populations. *Nat Genet*. 2015 Sep; 47(9):979–86. doi: [10.1038/ng.3359](https://doi.org/10.1038/ng.3359) PMID: [26192919](https://pubmed.ncbi.nlm.nih.gov/26192919/)
20. Vang T, Congia M, Macis MD, Musumeci L, Orru V, Zavattari P, et al. Autoimmune-associated lymphoid tyrosine phosphatase is a gain-of-function variant. *Nat Genet*. 2005 Dec; 37(12):1317–9. PMID: [16273109](https://pubmed.ncbi.nlm.nih.gov/16273109/)
21. Zhang J, Zahir N, Jiang Q, Miliotis H, Heyraud S, Meng X, et al. The autoimmune disease-associated PTPN22 variant promotes calpain-mediated Lyp/Pep degradation associated with lymphocyte and dendritic cell hyperresponsiveness. *Nat Genet*. 2011 Sep; 43(9):902–7. doi: [10.1038/ng.904](https://doi.org/10.1038/ng.904) PMID: [21841778](https://pubmed.ncbi.nlm.nih.gov/21841778/)
22. Vang T, Liu WH, Delacroix L, Wu S, Vasile S, Dahl R, et al. LYP inhibits T-cell activation when dissociated from CSK. *Nat Chem Biol*. 2012 May; 8(5):437–46. doi: [10.1038/nchembio.916](https://doi.org/10.1038/nchembio.916) PMID: [22426112](https://pubmed.ncbi.nlm.nih.gov/22426112/)
23. Dai X, James RG, Habib T, Singh S, Jackson S, Khim S, et al. A disease-associated PTPN22 variant promotes systemic autoimmunity in murine models. *J Clin Invest*. 2013 May 1; 123(5):2024–36. doi: [10.1172/JCI66963](https://doi.org/10.1172/JCI66963) PMID: [23619366](https://pubmed.ncbi.nlm.nih.gov/23619366/)
24. Arimura Y, Yagi J. Comprehensive expression profiles of genes for protein tyrosine phosphatases in immune cells. *Sci Signal*. 2010; 3(137):rs1. doi: [10.1126/scisignal.2000966](https://doi.org/10.1126/scisignal.2000966) PMID: [20807954](https://pubmed.ncbi.nlm.nih.gov/20807954/)
25. Hasegawa K, Martin F, Huang G, Tumas D, Diehl L, Chan AC. PEST domain-enriched tyrosine phosphatase (PEP) regulation of effector/memory T cells. *Science*. 2004 Jan 30; 303(5658):685–9. PMID: [14752163](https://pubmed.ncbi.nlm.nih.gov/14752163/)
26. Maine CJ, Hamilton-Williams EE, Cheung J, Stanford SM, Bottini N, Wicker LS, et al. PTPN22 alters the development of regulatory T cells in the thymus. *J Immunol*. 2012 Jun 1; 188(11):5267–75. doi: [10.4049/jimmunol.1200150](https://doi.org/10.4049/jimmunol.1200150) PMID: [22539785](https://pubmed.ncbi.nlm.nih.gov/22539785/)
27. Maine CJ, Marquardt K, Cheung J, Sherman LA. PTPN22 controls the germinal center by influencing the numbers and activity of T follicular helper cells. *J Immunol*. 2014 Feb 15; 192(4):1415–24. doi: [10.4049/jimmunol.1302418](https://doi.org/10.4049/jimmunol.1302418) PMID: [24453256](https://pubmed.ncbi.nlm.nih.gov/24453256/)
28. Spalinger MR, Lang S, Weber A, Frei P, Fried M, Rogler G, et al. Loss of protein tyrosine phosphatase nonreceptor type 22 regulates interferon-gamma-induced signaling in human monocytes. *Gastroenterology*. 2013 May; 144(5):978–88 e10. doi: [10.1053/j.gastro.2013.01.048](https://doi.org/10.1053/j.gastro.2013.01.048) PMID: [23380085](https://pubmed.ncbi.nlm.nih.gov/23380085/)
29. Spalinger MR, Lang S, Vavricka SR, Fried M, Rogler G, Scharl M. Protein tyrosine phosphatase nonreceptor type 22 modulates NOD2-induced cytokine release and autophagy. *PLoS One*. 2013; 8(8):e72384. doi: [10.1371/journal.pone.0072384](https://doi.org/10.1371/journal.pone.0072384) PMID: [23991106](https://pubmed.ncbi.nlm.nih.gov/23991106/)
30. Chang HH, Miaw SC, Tseng W, Sun YW, Liu CC, Tsao HW, et al. PTPN22 modulates macrophage polarization and susceptibility to dextran sulfate sodium-induced colitis. *J Immunol*. 2013 Sep 1; 191(5):2134–43. doi: [10.4049/jimmunol.1203363](https://doi.org/10.4049/jimmunol.1203363) PMID: [23913970](https://pubmed.ncbi.nlm.nih.gov/23913970/)

31. Wang Y, Shaked I, Stanford SM, Zhou W, Curtsinger JM, Mikulski Z, et al. The Autoimmunity-Associated Gene PTPN22 Potentiates Toll-like Receptor-Driven, Type 1 Interferon-Dependent Immunity. *Immunity*. 2013 Jul 25; 39(1):111–22. doi: [10.1016/j.immuni.2013.06.013](https://doi.org/10.1016/j.immuni.2013.06.013) PMID: [23871208](https://pubmed.ncbi.nlm.nih.gov/23871208/)
32. Wang Y, Ewart D, Crabtree JN, Yamamoto A, Baechler EC, Fazeli P, et al. PTPN22 Variant R620W Is Associated With Reduced Toll-like Receptor 7-Induced Type I Interferon in Systemic Lupus Erythematosus. *Arthritis Rheumatol*. 2015 Sep; 67(9):2403–14. doi: [10.1002/art.39211](https://doi.org/10.1002/art.39211) PMID: [26018863](https://pubmed.ncbi.nlm.nih.gov/26018863/)
33. Moran CJ, Walters TD, Guo CH, Kugathasan S, Klein C, Turner D, et al. IL-10R polymorphisms are associated with very-early-onset ulcerative colitis. *Inflamm Bowel Dis*. 2013 Jan; 19(1):115–23. doi: [10.1002/ibd.22974](https://doi.org/10.1002/ibd.22974) PMID: [22550014](https://pubmed.ncbi.nlm.nih.gov/22550014/)
34. Kotlarz D, Beier R, Murugan D, Diestelhorst J, Jensen O, Boztug K, et al. Loss of interleukin-10 signaling and infantile inflammatory bowel disease: implications for diagnosis and therapy. *Gastroenterology*. 2012 Aug; 143(2):347–55. doi: [10.1053/j.gastro.2012.04.045](https://doi.org/10.1053/j.gastro.2012.04.045) PMID: [22549091](https://pubmed.ncbi.nlm.nih.gov/22549091/)
35. Russell RK, Drummond HE, Nimmo EE, Anderson N, Smith L, Wilson DC, et al. Genotype-phenotype analysis in childhood-onset Crohn's disease: NOD2/CARD15 variants consistently predict phenotypic characteristics of severe disease. *Inflamm Bowel Dis*. 2005 Nov; 11(11):955–64. PMID: [16239840](https://pubmed.ncbi.nlm.nih.gov/16239840/)
36. Connelly TM, Berg AS, Harris L 3rd, Brinton D, Deiling S, Koltun WA. Genetic determinants associated with early age of diagnosis of IBD. *Dis Colon Rectum*. 2015 Mar; 58(3):321–7. doi: [10.1097/DCR.000000000000274](https://doi.org/10.1097/DCR.000000000000274) PMID: [25664710](https://pubmed.ncbi.nlm.nih.gov/25664710/)
37. Torki M, Gholamrezaei A, Mirbagher L, Danesh M, Kheiri S, Emami MH. Vitamin D Deficiency Associated with Disease Activity in Patients with Inflammatory Bowel Diseases. *Dig Dis Sci*. 2015 Oct; 60(10):3085–91. doi: [10.1007/s10620-015-3727-4](https://doi.org/10.1007/s10620-015-3727-4) PMID: [26031421](https://pubmed.ncbi.nlm.nih.gov/26031421/)
38. Nowak JK, Grzybowska-Chlebowczyk U, Landowski P, Szaflarska-Poplawska A, Klinciewicz B, Adamczak D, et al. Prevalence and correlates of vitamin K deficiency in children with inflammatory bowel disease. *Sci Rep*. 2014; 4:4768. doi: [10.1038/srep04768](https://doi.org/10.1038/srep04768) PMID: [24759680](https://pubmed.ncbi.nlm.nih.gov/24759680/)
39. Fiorillo E, Orru V, Stanford SM, Liu Y, Salek M, Rapini N, et al. Autoimmune-associated PTPN22 R620W variation reduces phosphorylation of lymphoid phosphatase on an inhibitory tyrosine residue. *J Biol Chem*. 2010 Aug 20; 285(34):26506–18. doi: [10.1074/jbc.M110.111104](https://doi.org/10.1074/jbc.M110.111104) PMID: [20538612](https://pubmed.ncbi.nlm.nih.gov/20538612/)
40. Spalinger MR, Kasper S, Gottier C, Lang S, Atrott K, Vavricka SR, et al. NLRP3 tyrosine phosphorylation is controlled by protein tyrosine phosphatase PTPN22. *The Journal of Clinical Investigation*. 2016/04/04; 126(5).
41. Spalinger MR, McCole DF, Rogler G, Scharl M. Role of protein tyrosine phosphatases in regulating the immune system: implications for chronic intestinal inflammation. *Inflamm Bowel Dis*. 2015 Mar; 21(3):645–55. doi: [10.1097/MIB.000000000000297](https://doi.org/10.1097/MIB.000000000000297) PMID: [25581833](https://pubmed.ncbi.nlm.nih.gov/25581833/)
42. Zenewicz LA, Antov A, Flavell RA. CD4 T-cell differentiation and inflammatory bowel disease. *Trends Mol Med*. 2009 May; 15(5):199–207. doi: [10.1016/j.molmed.2009.03.002](https://doi.org/10.1016/j.molmed.2009.03.002) PMID: [19362058](https://pubmed.ncbi.nlm.nih.gov/19362058/)
43. Massironi S, Rossi RE, Cavalcoli FA, Della Valle S, Fraquelli M, Conte D. Nutritional deficiencies in inflammatory bowel disease: therapeutic approaches. *Clin Nutr*. 2013 Dec; 32(6):904–10. doi: [10.1016/j.clnu.2013.03.020](https://doi.org/10.1016/j.clnu.2013.03.020) PMID: [23602613](https://pubmed.ncbi.nlm.nih.gov/23602613/)
44. Weissshof R, Chermesh I. Micronutrient deficiencies in inflammatory bowel disease. *Curr Opin Clin Nutr Metab Care*. 2015 Nov; 18(6):576–81. doi: [10.1097/MCO.000000000000226](https://doi.org/10.1097/MCO.000000000000226) PMID: [26418823](https://pubmed.ncbi.nlm.nih.gov/26418823/)
45. Alastair F, Emma G, Emma P. Nutrition in Inflammatory Bowel Disease. *Journal of Parenteral and Enteral Nutrition*. 2011 September 1, 2011; 35(5):571–80. doi: [10.1177/0148607111413599](https://doi.org/10.1177/0148607111413599) PMID: [21825089](https://pubmed.ncbi.nlm.nih.gov/21825089/)
46. Ardesia M, Ferlazzo G, Fries W. Vitamin d and inflammatory bowel disease. *Biomed Res Int*. 2015; 2015:470805. doi: [10.1155/2015/470805](https://doi.org/10.1155/2015/470805) PMID: [26000293](https://pubmed.ncbi.nlm.nih.gov/26000293/)
47. Chen J, Bruce D, Cantorna MT. Vitamin D receptor expression controls proliferation of naive CD8+ T cells and development of CD8 mediated gastrointestinal inflammation. *BMC Immunol*. 2014; 15:6. doi: [10.1186/1471-2172-15-6](https://doi.org/10.1186/1471-2172-15-6) PMID: [24502291](https://pubmed.ncbi.nlm.nih.gov/24502291/)
48. Wang TT, Dabbas B, Laperriere D, Bitton AJ, Soualhine H, Tavera-Mendoza LE, et al. Direct and indirect induction by 1,25-dihydroxyvitamin D3 of the NOD2/CARD15-defensin beta2 innate immune pathway defective in Crohn disease. *J Biol Chem*. 2010 Jan 22; 285(4):2227–31. doi: [10.1074/jbc.C109.071225](https://doi.org/10.1074/jbc.C109.071225) PMID: [19948723](https://pubmed.ncbi.nlm.nih.gov/19948723/)
49. Jeffery LE, Burke F, Mura M, Zheng Y, Qureshi OS, Hewison M, et al. 1,25-Dihydroxyvitamin D3 and IL-2 combine to inhibit T cell production of inflammatory cytokines and promote development of regulatory T cells expressing CTLA-4 and FoxP3. *J Immunol*. 2009 Nov 1; 183(9):5458–67. doi: [10.4049/jimmunol.0803217](https://doi.org/10.4049/jimmunol.0803217) PMID: [19843932](https://pubmed.ncbi.nlm.nih.gov/19843932/)
50. Wu S, Zhang YG, Lu R, Xia Y, Zhou D, Petrof EO, et al. Intestinal epithelial vitamin D receptor deletion leads to defective autophagy in colitis. *Gut*. 2015 Jul; 64(7):1082–94. doi: [10.1136/gutjnl-2014-307436](https://doi.org/10.1136/gutjnl-2014-307436) PMID: [25080448](https://pubmed.ncbi.nlm.nih.gov/25080448/)

51. Pittet V, Juillerat P, Mottet C, Felley C, Ballabeni P, Burnand B, et al. Cohort profile: the Swiss Inflammatory Bowel Disease Cohort Study (SIBDCS). *Int J Epidemiol*. 2009 Aug; 38(4):922–31. doi: [10.1093/ije/dyn180](https://doi.org/10.1093/ije/dyn180) PMID: [18782896](https://pubmed.ncbi.nlm.nih.gov/18782896/)
52. Silverberg MS, Satsangi J, Ahmad T, Arnott ID, Bernstein CN, Brant SR, et al. Toward an integrated clinical, molecular and serological classification of inflammatory bowel disease: report of a Working Party of the 2005 Montreal World Congress of Gastroenterology. *Can J Gastroenterol*. 2005 Sep; 19 Suppl A:5A–36A. PMID: [16151544](https://pubmed.ncbi.nlm.nih.gov/16151544/)
53. Nguyen GC, Munsell M, Harris ML. Nationwide prevalence and prognostic significance of clinically diagnosable protein-calorie malnutrition in hospitalized inflammatory bowel disease patients. *Inflamm Bowel Dis*. 2008 Aug; 14(8):1105–11. doi: [10.1002/ibd.20429](https://doi.org/10.1002/ibd.20429) PMID: [18302272](https://pubmed.ncbi.nlm.nih.gov/18302272/)