

# 1 **Genome structures resolve the early diversification of** 2 **teleost fishes**

3 Elise Parey<sup>1,2</sup>, Alexandra Louis<sup>1</sup>, Jerome Montfort<sup>2</sup>, Olivier Bouchez<sup>3</sup>, Céline Roques<sup>3</sup>, Carole  
4 Iampietro<sup>3</sup>, Jerome Lluch<sup>3</sup>, Adrien Castinel<sup>3</sup>, Cécile Donnadiou<sup>3</sup>, Thomas Desvignes<sup>4</sup>, Christabel Floi  
5 Bucao<sup>5,6</sup>, Elodie Jouanno<sup>2</sup>, Ming Wen<sup>2,7</sup>, Sahar Mejri<sup>8</sup>, Ron Dirks<sup>9</sup>, Hans Jansen<sup>9</sup>, Christiaan  
6 Henkel<sup>10,11</sup>, Wei-Jen Chen<sup>12</sup>, Margot Zahm<sup>13</sup>, Cédric Cabau<sup>13</sup>, Christophe Klopp<sup>14</sup>, Andrew W.  
7 Thompson<sup>15,16</sup>, Marc Robinson-Rechavi<sup>5,6</sup>, Ingo Braasch<sup>15,16</sup>, Guillaume Lecointre<sup>17</sup>, Julien Bobe<sup>2</sup>,  
8 John H. Postlethwait<sup>4</sup>, Camille Berthelot<sup>1,18\*†</sup>, Hugues Roest Crollius<sup>1\*†</sup>, Yann Guiguen<sup>2\*†</sup>

9 <sup>1</sup> Institut de Biologie de l'ENS (IBENS), Département de Biologie, École Normale Supérieure, CNRS,  
10 INSERM, Université PSL, Paris, France.

11 <sup>2</sup> INRAE, LPGP, Rennes, France.

12 <sup>3</sup> INRAE, GeT-PlaGe, Genotoul, 31326 Castanet-Tolosan, France.

13 <sup>4</sup> Institute of Neuroscience, University of Oregon, Eugene, United States.

14 <sup>5</sup> Department of Ecology and Evolution, University of Lausanne, Lausanne, Switzerland.

15 <sup>6</sup> SIB Swiss Institute of Bioinformatics, Lausanne, Switzerland.

16 <sup>7</sup> State Key Laboratory of Developmental Biology of Freshwater Fish, College of Life Science, Hunan  
17 Normal University, Changsha, China.

18 <sup>8</sup> Florida Atlantic University, Harbor Branch Oceanographic Institute, Fort Pierce, Florida, USA.

19 <sup>9</sup> Future Genomics Technologies, Leiden, The Netherlands.

20 <sup>10</sup> Institute of Biology, University of Leiden, Leiden, The Netherlands.

21 <sup>11</sup> Faculty of Veterinary Medicine, Norwegian University of Life Sciences, Ås, Norway.

22 <sup>12</sup> Institute of Oceanography, National Taiwan University, Taipei 10617, Taiwan.

23 <sup>13</sup> Sigénae, GenPhySE, INRAE, ENVT, Université de Toulouse, Castanet Tolosan, France.

24 <sup>14</sup> Sigénae, Genotoul Bioinfo, MIAT UR875, INRAE, Castanet Tolosan, France.

25 <sup>15</sup> Department of Integrative Biology, Michigan State University, East Lansing, MI, USA.

26 <sup>16</sup> Ecology, Evolution & Behavior Program, Michigan State University, East Lansing, MI, USA.

27 <sup>17</sup> Institut Systématique, Evolution, Biodiversité (ISYEB), Muséum national d'Histoire naturelle,  
28 CNRS, SU, EPHE, UA, Paris, France.

29 <sup>18</sup> Institut Pasteur, Université de Paris, CNRS UMR 3525, INSERM UA12, Comparative Functional  
30 Genomics group, F-75015 Paris, France.

31

32 \* Corresponding authors. Emails: [camille.berthelot@pasteur.fr](mailto:camille.berthelot@pasteur.fr), [hrc@bio.ens.psl.eu](mailto:hrc@bio.ens.psl.eu),  
33 [yann.guiguen@inrae.fr](mailto:yann.guiguen@inrae.fr)

34 †These authors contributed equally to this work

35 **Abstract:** Accurate species phylogenies are a prerequisite for evolutionary research. Teleosts are by  
36 far the largest and the most diversified group of extant vertebrates, but relationships among the three  
37 oldest lineages of extant teleosts remain unresolved. Based on seven high-quality new genome  
38 assemblies in Elopomorpha (tarpons, eels), we revisited the topology of the deepest branches of  
39 the teleost phylogeny using independent gene sequence and chromosomal rearrangement  
40 phylogenomic approaches. These analyses converged to a single scenario that unambiguously places  
41 the Elopomorpha and Osteoglossomorpha (bony-tongues) in a monophyletic group sister to all other  
42 teleosts, i.e., the Clupeocephala lineage. This finding resolves over 50 years of controversy on the  
43 evolutionary relationships of these lineages and highlights the power of combining different levels of  
44 genome-wide information to solve complex phylogenies.

45

46 **One-Sentence Summary:** Whole-genome analyses place Elopomorpha (tarpons, eels) and  
47 Osteoglossomorpha (bony-tongues) as sister groups at the deepest branching of crown teleosts.

48

49

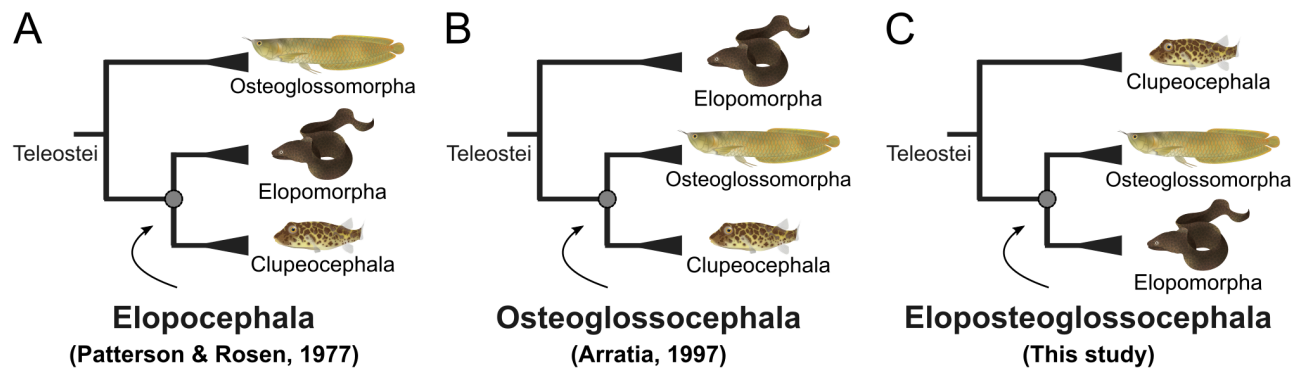
## 50 **Main Text**

51 Species phylogenies retrace sister-group relationships resulting from evolutionary histories and  
52 pathways from common ancestors to descendant species (1). Accurate species phylogenies are  
53 important for our understanding and representation of the evolution of life on earth, but they are also  
54 a fundamental prerequisite for evolutionary analyses at the developmental, anatomical, genetic, and  
55 species levels.

56 With more than 30,000 species, teleost fishes are by far the largest and the most diversified clade of  
57 extant vertebrates (2). Understanding their phylogeny has been and is still subject to many disputes  
58 at different taxonomic levels (2, 3). Among these debates, a long-standing and unresolved question  
59 concerns the topology of the earliest-branching clades of crown teleosts, i.e., the Elopomorpha  
60 (named after “*Elops*-like” and including tarpon, bonefish and eels) and the Osteoglossomorpha  
61 (named after “bony-tongues” and including goldeye, arapaima, and elephantnose fish) relative to all  
62 the other extant teleosts in the Clupeocephala lineage (including for instance zebrafish, a major  
63 biomedical species) (3, 4).

64 Based on anatomical and morphological characters, Elopiformes (tarpons and ladyfishes) were first  
65 suggested to be the most “primitive living teleosts” nearly 100 years ago ((5) cited in (4)). Since then,  
66 Elopomorpha and Osteoglossomorpha have been alternatively placed as the earliest branching clade  
67 of teleosts. A first scenario, proposed in 1977 (6), placed the Osteoglossomorpha as the earliest teleost  
68 crown group and outgroup to the Elopocephala consisting of Elopomorpha and Clupeocephala. (Fig.  
69 1A). This scenario was later challenged in 1997 (7, 8) by the placement of Elopomorpha as the earliest  
70 branching clade of teleosts, with Osteoglossomorpha and Clupeocephala composing the  
71 Osteoglossocephala clade (Fig. 1B). These early controversies were based on morphological evidence  
72 and remained largely unsolved, but the most recent authoritative view still considers the Elopomorpha  
73 as the earliest branching clade of crown teleosts (2).

74



75

76 **Figure 1: Alternative phylogenetic hypotheses for the earliest-branching teleost clades.** The Elopocephala (6) (A),  
77 Osteoglossocephala (7) (B) and Eloposteoglossocephala (C) hypotheses respectively, propose Clupeocephala and  
78 Elopomorpha, Clupeocephala and Osteoglossomorpha or Osteoglossomorpha and Elopomorpha as sister groups (see text  
79 for details).

80

81

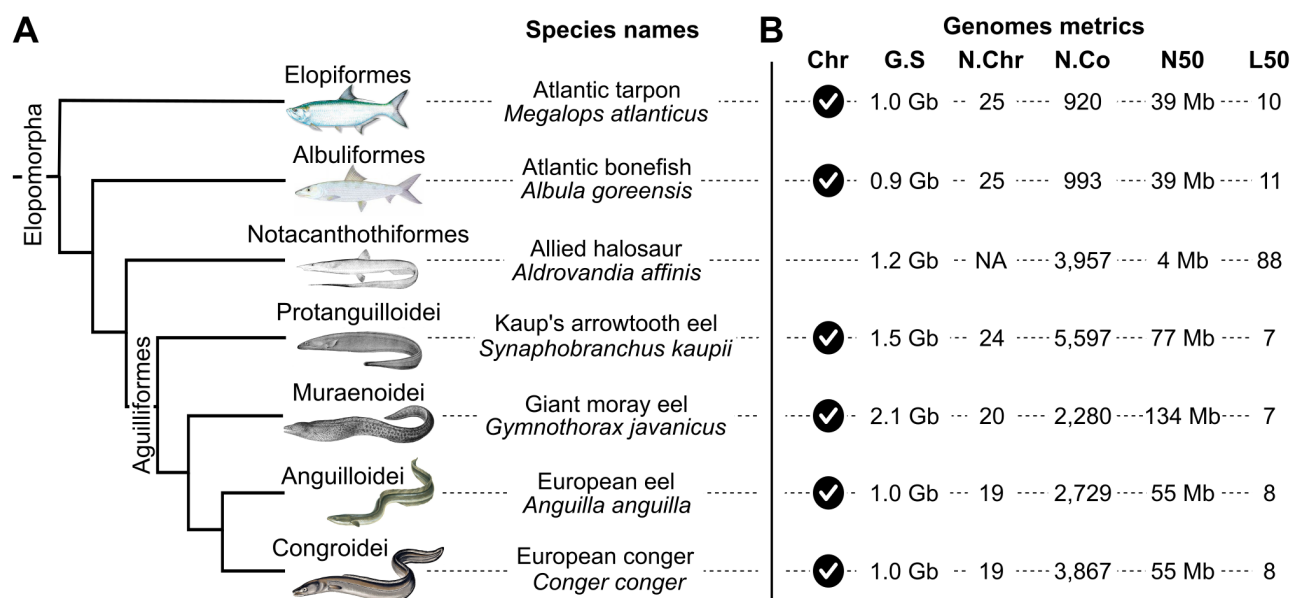
82 With the emergence of molecular phylogenetic approaches in the nineties, this question was  
83 extensively revisited using gene sequence phylogeny reconstructions (reviewed in (3, 9)). Despite  
84 extensive efforts, including several large-scale multi-locus approaches (10–12), no consensus has,  
85 however, been reached in favor of neither the Elopocephala nor the Osteoglossocephala hypothesis.  
86 In addition, a third topology placing Elopomorpha and Osteoglossomorpha as sister groups (Fig. 1C)  
87 was even suggested in the early nineties (13) and since then supported by a few more recent studies  
88 (11, 14–18). This clade, which we tentatively named the Eloposteoglossocephala (Fig. 1C), was never  
89 formally retained, probably because this topology was not supported by any morphological evidence  
90 (3). The prevailing hypothesis, confirmed by a recent meta-analysis of gene sequence phylogeny  
91 studies (9), thus remains the Osteoglossocephala hypothesis that places Elopomorpha as the earliest  
92 branching clade of extant teleosts (12)). However, the precise phylogenetic relationships of these  
93 major teleost lineages are still debated and have even been recently reviewed by Dornburg and Near  
94 (3) as one of the major unresolved questions of the twenty-first century regarding the evolution of  
95 actinopterygian fishes. To promote a reexamination of this problem, they provocatively proposed to  
96 retain “the unconventional and intriguing possibility of an osteoglossomorph and elopomorph sister  
97 group relationship” (3).

98 To resolve the phylogenetic relationships of these early-branching teleost clades, we first sequenced,  
99 assembled, and annotated high-quality reference genome sequences of seven species that represent

100 major Elopomorpha orders or families (Fig. 2 and table S1) for which chromosome-level whole-  
 101 genome resources were lacking. We combined genome information from these seven Elopomorpha  
 102 species with 18 additional publicly available genome assemblies including four Osteoglossomorpha,  
 103 10 Clupeocephala and four vertebrate outgroups, including the spotted gar and bowfin non-teleost  
 104 fishes, to perform phylogenomic analyses.

105

106



107

108 **Figure 2: Phylogeny of representative Elopomorpha species.** (A) This tree topology is based on ref. (19) and  
 109 includes each species for which we provide novel whole-genome assemblies. (B) Six of these seven genomes were  
 110 assembled at chromosome scale (Chr) with high-quality genome assembly metrics. G.S = genome size, N.Chr =  
 111 haploid chromosome number, N.Co = number of contigs, N50 = sequence length at which half of the genome  
 112 assembly is covered by longer sequences, L50 = smallest number of scaffolds needed to sum to half of the predicted  
 113 genome size.

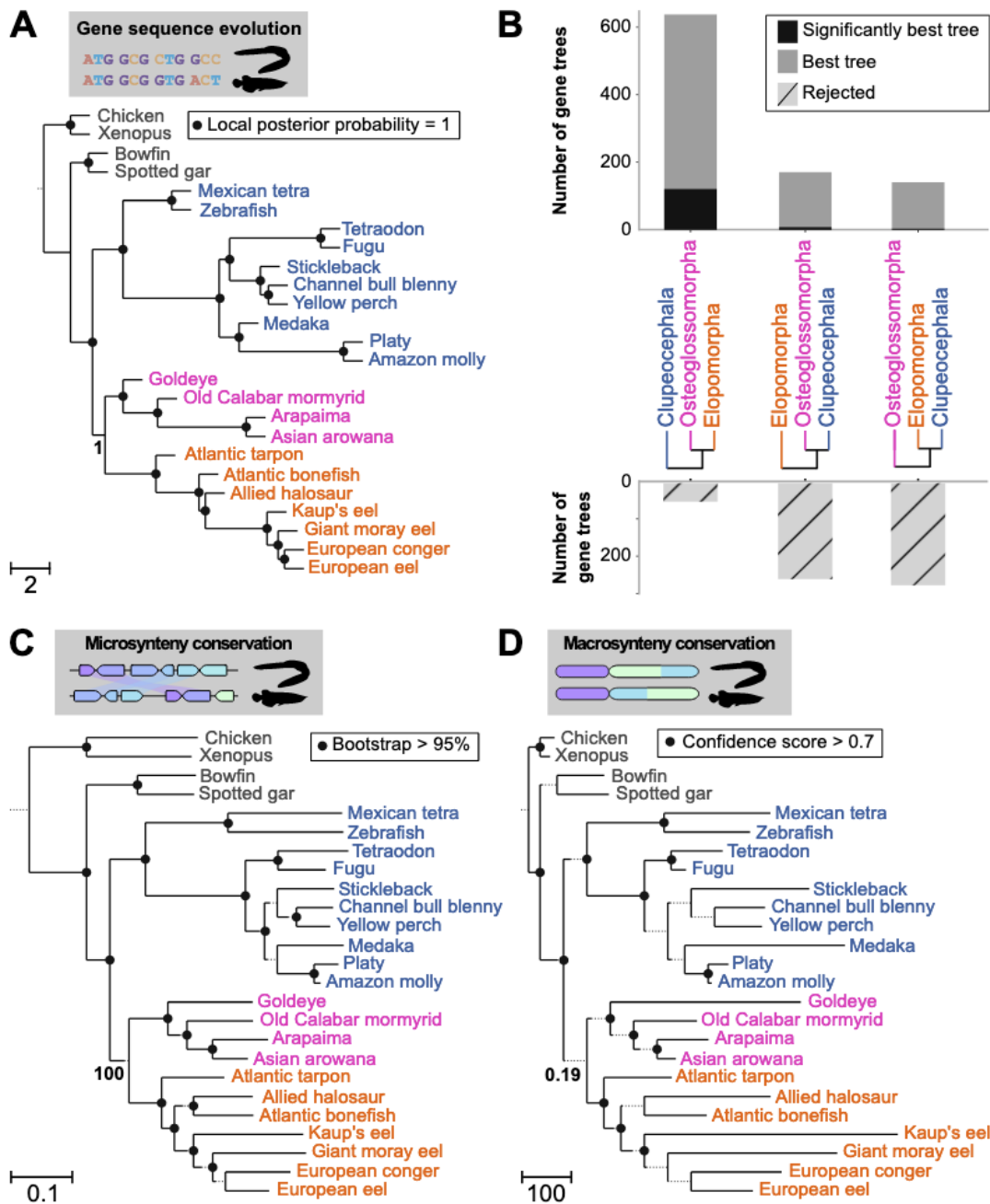
114

115

116 A major challenge for achieving accurate phylogenetic analysis of teleost genomes is their high  
 117 number of duplicate (paralogous) gene copies. Many of these paralogs are inherited from a whole  
 118 genome duplication (WGD) in their last common ancestor (20), and are known to mislead  
 119 phylogenetic reconstructions (11). To mitigate the effect of paralog inclusion, we applied a WGD-  
 120 tailored pipeline leveraging gene sequences and synteny conservation (supplementary materials,

121 Methods section) to select 955 high-confidence 1-to-1 orthologous genes across all the 25 genomes  
122 we analyzed. This list represents by far the largest molecular dataset considered for teleost phylogeny  
123 reconstruction, both in terms of included Elopomorpha genomes and of total alignment size (see fig.  
124 S1). We then performed phylogenetic reconstructions of these 955 individual gene trees using  
125 summary analyses with ASTRAL (Fig. 3A, and fig. S2 for protein trees), as well as Maximum  
126 Likelihood analyses of their concatenated sequences both at the nucleotide and amino-acid levels (fig.  
127 S3 and S4). These analyses all provided highly significant support for the Eloposteoglossocephala  
128 hypothesis that places Osteoglossomorpha and Elopomorpha as sister groups. Additionally, this  
129 Eloposteoglossocephala clade was further supported by gene-genealogy interrogation, which directly  
130 compares the likelihood of each of the three evolutionary scenarios based on individual gene sequence  
131 alignment (Fig. 3B).

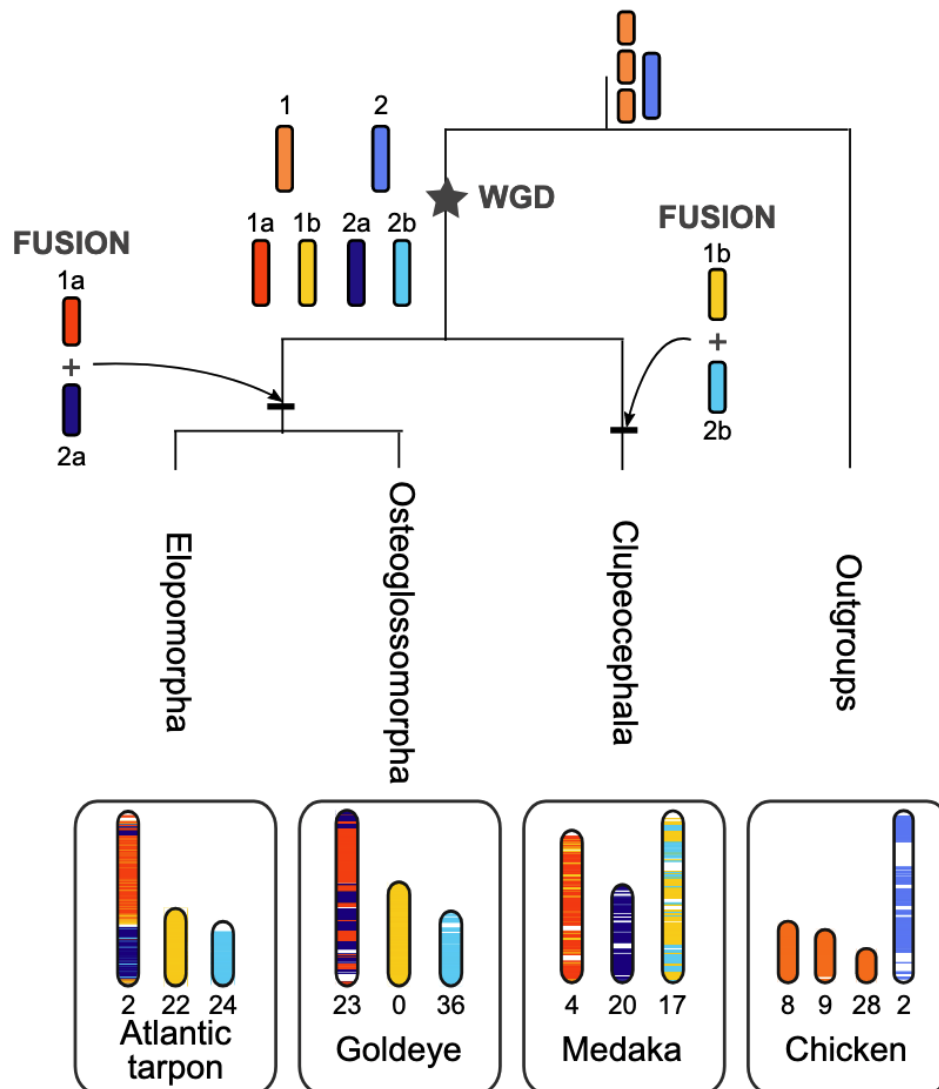
132 However, because previous sequence-based studies have yielded opposing results to resolve the three  
133 early diverging teleost branches (9, 11), we also used two novel genome-wide methods to infer  
134 species trees based on conservation of genome structures. First, we analyzed the conservation of gene  
135 adjacencies between 3,041 orthologous marker genes covering 57-98% of each teleost genome, and  
136 inferred a Neighbor-Joining species tree from local microsyntenic conservation (21). Second, we  
137 analyzed macrosyntenic evolution by measuring the fraction of shared chromosomal breakpoints  
138 between species with PhyChro (22). These two complementary approaches (Figs. 3C-3D, fig. S5)  
139 also provided convergent and robust support for the Eloposteoglossocephala scenario, confirming the  
140 results from gene sequence phylogenies.



141

142 **Figure 3: Different phylogenomic reconstruction methods converge to support Osteoglossomorpha and**  
 143 **Elopomorpha as sister groups.** (A) Species tree inferred under the multispecies coalescent model (23) from 955  
 144 single-copy gene trees. This method measures internal branch lengths in coalescent units, while lengths at terminal  
 145 branches are represented as equal and arbitrary. Node support is calculated as a local posterior probability. (B)  
 146 Molecular gene genealogy interrogation: number of gene trees supporting each hypothesis (top, gray bars),  
 147 significantly supporting each hypothesis (top, black bars), and number of significantly rejected gene trees (bottom,  
 148 dashed bar). (C) Neighbor-Joining species tree estimated from gene adjacencies between 3,041 marker genes.  
 149 Branch lengths represent the proportion of broken adjacencies and nodes are supported by bootstrap values. (D)  
 150 Best-supported species tree based on shared synteny breakpoints. Branch lengths are an estimate of the number of  
 151 breakpoints. Node supports correspond to the fraction of informative breakpoints that support the clade (22). (A-D)  
 152 Broken lines have been added to very short branches, for visualization purposes.

153 Finally, by looking at all potential chromosomal macro-rearrangements we identified a single  
 154 chromosomal fusion exclusively shared between karyotypes of Osteoglossomorpha and  
 155 Elopomorpha species (Fig. 4, fig. S6). Together with the absence of other rearrangements that would  
 156 be consistent with alternative groupings, this identical chromosomal macro-rearrangement further  
 157 supports that the two groups descend from a common ancestor, strengthening the phylogenomics  
 158 evidence for the Elopsteoglossocephala clade.  
 159



160

161 **Figure 4: Chromosomal rearrangement shared within the Elopsteoglossocephala clade.** Inferred  
 162 rearrangement scenario for two pairs of duplicated chromosomes (1a-1b and 2a-2b) in teleosts. Osteoglossomorpha  
 163 and Elopomorpha share the 1a-2a chromosomal fusion, while Clupeocephala experienced an independent fusion  
 164 between 1b and 2b.



165

166 Using a combination of new whole-genome resources for Elopomorpha and an array of  
167 complementary phylogenomic reconstruction methods, we unambiguously resolved the long-  
168 standing question of the topology of the deepest branches in the phylogeny of extant teleost fishes.  
169 This achievement highlights the power of genome-wide methods to resolve complex and ancient  
170 phylogenies, especially when these methods consider a variety of informative evolutionary characters  
171 in complement to sequence information. Chromosome rearrangements, in particular, are fixed at a  
172 low rate and thus are less prone to mutational saturation and character reversal, which can occur in  
173 sequence-based phylogenies (24).

174 Our results resolve over 50 years of controversy and demonstrate that Elopomorpha and  
175 Osteoglossomorpha constitute a clade for which we propose the name Eloposteoglossocephala  
176 (supplementary materials, section 1). This conclusion raises questions about the paucity of anatomical  
177 evidence in favor of this hypothesis, despite more than 70 years of extensive research (3, 4). We  
178 carefully reexamined the available literature on these anatomical characters in light of our results and  
179 we were not able to find a morphological character exclusively and unambiguously shared by  
180 Elopomorpha and Osteoglossomorpha (supplementary materials, section 2). However, the fusion of  
181 the retroarticular with the angular and/or the articular, a derived character previously considered a  
182 synapomorphy of the Elopomorpha (25, 26), has been shown to be shared with at least mormyrids  
183 among bony-tongues (26, 27). Even if this character is described as either present (27) or ambiguous  
184 in goldeye *Hiodon alosoides*, and absent in two other Osteoglossomorpha (26), we propose this  
185 derived state as a morphological synapomorphy of the Eloposteoglossocephala, which was  
186 secondarily lost in some Osteoglossomorpha. We anticipate that based on our results, more character  
187 mapping and new targeted anatomical and morphological searches will soon provide novel and non-  
188 ambiguous synapomorphies shared by the Eloposteoglossocephala.

189

## 190 **References**

- 191 1. D. A. Baum, S. D. Smith, S. S. S. Donovan, The Tree-Thinking Challenge. *Science*. **310**, 979–  
192 980 (2005).
- 193 2. J. S. Nelson, T. C. Grande, M. V. H. Wilson, *Fishes of the World* (John Wiley & Sons, 2016).
- 194 3. A. Dornburg, T. J. Near, The Emerging Phylogenetic Perspective on the Evolution of  
195 Actinopterygian Fishes. *Annu. Rev. Ecol. Evol. Syst.* **52**, 427–452 (2021).
- 196 4. G. Arratia, Phylogenetic relationships of Teleostei. Past and present. *Estud. Oceanol.* **19**, 19–

- 197 51 (2000).
- 198 5. C. T. Regan, The Skeleton of *Lepidosteus*, with remarks on the origin and evolution of the  
199 lower Neopterygian Fishes. *Proceedings of the Zoological Society of London*. **1923**, 445–461 (1923).
- 200 6. C. Patterson, D. E. Rosen, Review of ichthyodectiform and other Mesozoic teleost fishes, and  
201 the theory and practice of classifying fossils. *Bulletin of the AMNH*. **158** (1977) (available at  
202 <https://digitallibrary.amnh.org/handle/2246/1224>).
- 203 7. G. Arratia, Basal teleosts and teleostean phylogeny. *Paleoichthyologica* **168**, 1-168 (Verlag Dr.  
204 Friedrich Pfeil, München, 1997).
- 205 8. G. Arratia, Basal Teleosts and Teleostean Phylogeny: Response to C. Patterson. *Copeia*. **1998**,  
206 1109–1113 (1998).
- 207 9. N. Takezaki, Resolving the Early Divergence Pattern of Teleost Fish Using Genome-Scale  
208 Data. *Genome Biol Evol*. **13**, evab052 (2021).
- 209 10. T. J. Near, R. I. Eytan, A. Dornburg, K. L. Kuhn, J. A. Moore, M. P. Davis, P. C. Wainwright,  
210 M. Friedman, W. L. Smith, Resolution of ray-finned fish phylogeny and timing of diversification.  
211 *Proc Natl Acad Sci U S A*. **109**, 13698–13703 (2012).
- 212 11. L. C. Hughes, G. Ortí, Y. Huang, Y. Sun, C. C. Baldwin, A. W. Thompson, D. Arcila, R.  
213 Betancur-R, C. Li, L. Becker, N. Bellora, X. Zhao, X. Li, M. Wang, C. Fang, B. Xie, Z. Zhou, H.  
214 Huang, S. Chen, B. Venkatesh, Q. Shi, Comprehensive phylogeny of ray-finned fishes  
215 (Actinopterygii) based on transcriptomic and genomic data. *Proc Natl Acad Sci U S A*. **115**, 6249–  
216 6254 (2018).
- 217 12. R. Betancur-R, E. O. Wiley, G. Arratia, A. Acero, N. Bailly, M. Miya, G. Lecointre, G. Ortí,  
218 Phylogenetic classification of bony fishes. *BMC Evol Biol*. **17**, 162 (2017).
- 219 13. H. L. Le, G. Lecointre, R. Perasso, A 28S rRNA-based phylogeny of the gnathostomes: first  
220 steps in the analysis of conflict and congruence with morphologically based cladograms. *Mol*  
221 *Phylogenet Evol*. **2**, 31–51 (1993).
- 222 14. I. A. Hurley, R. L. Mueller, K. A. Dunn, E. J. Schmidt, M. Friedman, R. K. Ho, V. E. Prince,  
223 Z. Yang, M. G. Thomas, M. I. Coates, A new time-scale for ray-finned fish evolution. *Proc Biol Sci*.  
224 **274**, 489–498 (2007).
- 225 15. R. Broughton, in *Origin and phylogenetic interrelationships of teleosts* (Verlag Dr. Friedrich  
226 Pfeil, Munchen (Germany), J. S. Nelson, H.-P. Schultze&M. V. H. Wilson., 2010), pp. 61–76.
- 227 16. M.-Y. Chen, D. Liang, P. Zhang, Selecting Question-Specific Genes to Reduce Incongruence  
228 in Phylogenomics: A Case Study of Jawed Vertebrate Backbone Phylogeny. *Syst Biol*. **64**, 1104–  
229 1120 (2015).
- 230 17. C. Bian, Y. Hu, V. Ravi, I. S. Kuznetsova, X. Shen, X. Mu, Y. Sun, X. You, J. Li, X. Li, Y.  
231 Qiu, B.-H. Tay, N. M. Thevasagayam, A. S. Komissarov, V. Trifonov, M. Kabilov, A. Tupikin, J.  
232 Luo, Y. Liu, H. Song, C. Liu, X. Wang, D. Gu, Y. Yang, W. Li, G. Polgar, G. Fan, P. Zeng, H. Zhang,  
233 Z. Xiong, Z. Tang, C. Peng, Z. Ruan, H. Yu, J. Chen, M. Fan, Y. Huang, M. Wang, X. Zhao, G. Hu,  
234 H. Yang, J. Wang, J. Wang, X. Xu, L. Song, G. Xu, P. Xu, J. Xu, S. J. O'Brien, L. Orbán, B.  
235 Venkatesh, Q. Shi, The Asian arowana (*Scleropages formosus*) genome provides new insights into  
236 the evolution of an early lineage of teleosts. *Sci Rep*. **6**, 24501 (2016).
- 237 18. R. A. Vialle, J. E. S. de Souza, K. de P. Lopes, D. G. Teixeira, P. de A. Alves Sobrinho, A.  
238 M. Ribeiro-Dos-Santos, C. Furtado, T. Sakamoto, F. A. Oliveira Silva, E. Herculano Corrêa de  
239 Oliveira, I. G. Hamoy, P. P. Assumpção, Â. Ribeiro-Dos-Santos, J. P. M. Santos Lima, H. N. Seuánez,  
240 S. J. de Souza, S. Santos, Whole Genome Sequencing of the Pirarucu (*Arapaima gigas*) Supports  
241 Independent Emergence of Major Teleost Clades. *Genome Biol Evol*. **10**, 2366–2379 (2018).
- 242 19. J.-N. Chen, J. A. López, S. Lavoué, M. Miya, W.-J. Chen, Phylogeny of the Elopomorpha

- 243 (Teleostei): evidence from six nuclear and mitochondrial markers. *Mol Phylogenet Evol.* **70**, 152–  
244 161 (2014).
- 245 20. I. Braasch, J. H. Postlethwait, in *Polyploidy and Genome Evolution*, P. S. Soltis, D. E. Soltis,  
246 Eds. (Springer, Berlin, Heidelberg, 2012; [https://doi.org/10.1007/978-3-642-31442-1\\_17](https://doi.org/10.1007/978-3-642-31442-1_17)), pp. 341–  
247 383.
- 248 21. A. W. Thompson, M. B. Hawkins, E. Parey, D. J. Weisel, T. Ota, K. Kawasaki, E. Funk, M.  
249 Losilla, O. E. Fitch, Q. Pan, R. Feron, A. Louis, J. Montfort, M. Milhes, B. L. Racicot, K. L. Childs,  
250 Q. Fontenot, A. Ferrara, S. R. David, A. R. McCune, A. Dornburg, J. A. Yoder, Y. Guiguen, H. Roest  
251 Crollius, C. Berthelot, M. P. Harris, I. Braasch, The bowfin genome illuminates the developmental  
252 evolution of ray-finned fishes. *Nat Genet.* **53**, 1373–1384 (2021).
- 253 22. G. Drillon, R. Champeimont, F. Oteri, G. Fischer, A. Carbone, Phylogenetic Reconstruction  
254 Based on Synteny Block and Gene Adjacencies. *Mol Biol Evol.* **37**, 2747–2762 (2020).
- 255 23. C. Zhang, M. Rabiee, E. Sayyari, S. Mirarab, ASTRAL-III: polynomial time species tree  
256 reconstruction from partially resolved gene trees. *BMC Bioinformatics.* **19**, 153 (2018).
- 257 24. A. Rokas, P. W. H. Holland, Rare genomic changes as a tool for phylogenetics. *Trends in*  
258 *Ecology & Evolution.* **15**, 454–459 (2000).
- 259 25. C. Patterson, in *Major Patterns in Vertebrate Evolution*, M. K. Hecht, P. C. Goody, B. M.  
260 Hecht, Eds. (Springer US, Boston, MA, 1977; [https://doi.org/10.1007/978-1-4684-8851-7\\_21](https://doi.org/10.1007/978-1-4684-8851-7_21)),  
261 *NATO Advanced Study Institutes Series*, pp. 579–643.
- 262 26. R. Diogo, I. Doadrio, P. Vandewalle, Teleostean Phylogeny Based on Osteological and  
263 Myological Characters. *Int. J. Morphol.* **26** (2008), doi:10.4067/S0717-95022008000300001.
- 264 27. E. J. Hilton, Comparative osteology and phylogenetic systematics of fossil and living bony-  
265 tongue fishes (Actinopterygii, Teleostei, Osteoglossomorpha). *Zoological Journal of the Linnean*  
266 *Society.* **137**, 1–100 (2003).

267

268 **Acknowledgments:** We thank Yoann Guilloux, Fabien Quendo and Aaron J. Adams for their help  
269 in providing fish samples. We would also like to thank the leaders of the oceanography cruises and  
270 the crew of the RV Atalante, France and ORI, Taiwan in organizing the survey and helping to collect  
271 the deep-sea fish samples under the TDSB-TFDeepEvo joint Program.

272

273 **Funding:** This work was supported by the Agence Nationale de la Recherche, France (ANR) on the  
274 GenoFish project, 2016-2021 (grant No. ANR-16-CE12-003) to HRC., CB., JB., JHP., M.R.C and  
275 YG, and by France Génomique National infrastructure, funded as part of “Investissement d’avenir”  
276 program managed by ANR (grant No. ANR-10-INBS-09) to CD. Part of the fellowship to E.P. was  
277 supported by funds from the European Union Horizon 2020 research and innovation program under  
278 Grant Agreement No 817923 (AQUA-FAANG). M.R.-R. was supported by the Swiss National  
279 Science Foundation grant 31003A\_173048. J.H.P was supported by the National Institute of Health  
280 under grant agreement No R01OD011116.

281

282 **Author contributions:**

283       Conceptualization: CB, JB, IB, YG, CK, AL, JHP, MRR, HRC

284       Software: CB, AL, EP, HRC

285       Formal analysis: CB, AL, EP, HRC

286       Investigation: CB, OB, CC, AC, CD, RD, CFB, YG, CI, HJ, EJ, CK, GL, JL, AL, JM, EP,

287       HRC, CR, MW, MZ

288       Resources: JB, IB, WJC, RD, YG, CH, HJ, SM, JHP, AT, MW

289       Data curation: CB, CC, YG, CK, AL, JM, MZ

290       Visualization: CB, YG, AL, EP

291       Funding acquisition: CB, JB, CD, YG, JHP, MRR, HRC

292       Project administration: YG

293       Supervision: CB, YG, HRC

294       Writing – original draft: CB, YG, EP, HRC

295       Writing – review & editing: CB, JB, IB, CC, TD, YG, GL, EP, HRC, CR, MRR

296

297 **Competing interests:** Authors declare that they have no competing interests.

298 **Data and materials availability:** The Whole Genome Shotgun projects for the seven

299 Elopomorpha species are available in the Sequence Read Archive (SRA), under the following

300 BioProject references PRJNA702045 (*Conger conger*), PRJNA692825 (*Albula goreensis*),

301 PRJNA743502 (*Aldrovandia affinis*), PRJNA690086 (*Megalops atlanticus*), PRJNA693699

302 (*Anguilla anguilla*), PRJNA743503 (*Synaphobranchus kaupii*), PRJNA702255 (*Gymnothorax*

303 *javanicus*). All genome assemblies plus their annotations are also available in the omics Dataverse

304 (Open source research data repository) server (<https://doi.org/10.15454/GWL0GP>). All input data

305 (sets of orthologous marker genes, CDS codons alignments, gene coordinates files) and the

306 generated reconstructed species phylogenies have been deposited in Zenodo (doi:

307 10.5281/zenodo.6414307), along with all scripts and environments to reproduce the analyses.