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## A Dystroglycan Mutation Associated with Limb-Girdle Muscular Dystrophy

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### SUMMARY

Dystroglycan, which serves as a major extracellular matrix receptor in muscle and the central nervous system, requires extensive O-glycosylation to function. We identified a dystroglycan missense mutation (Thr192→Met) in a woman with limb-girdle muscular dystrophy and cognitive impairment. A mouse model harboring this mutation recapitulates the immunohistochemical and neuromuscular abnormalities observed in the patient. In vitro and in vivo studies showed that the mutation impairs the receptor function of dystroglycan in skeletal muscle and brain by inhibiting the post-translational modification, mediated by the glycosyltransferase LARGE, of the phosphorylated O-mannosyl glycans on  $\alpha$ -dystroglycan that is required for high-affinity binding to laminin.

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MUSCULAR DYSTROPHIES ARE GENETIC DISEASES CHARACTERIZED BY weakness and progressive degeneration of skeletal muscle. The transmembrane protein dystroglycan, which is ultimately cleaved into an  $\alpha$  and a  $\beta$  component, is a key link between the cytoskeleton and extracellular-matrix proteins that bear laminin globular domains (e.g., laminin, agrin, and neurexin).<sup>1,2</sup> The mucin domain of  $\alpha$ -dystroglycan is modified with numerous O-linked oligosaccharides that are essential for its normal function as an extracellular-matrix receptor in various tissues, including skeletal muscle and brain.<sup>2,3</sup>

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Hypoglycosylation of  $\alpha$ -dystroglycan and a consequent reduction of  $\alpha$ -dystroglycan binding to extracellular-matrix proteins are observed in patients with the Walker–Warburg syndrome, muscle–eye–brain disease, Fukuyama-type congenital muscular dystrophy, congenital muscular dystrophy types 1C and 1D, and limb-girdle muscular dystrophy 2I.<sup>2</sup> To date, genes encoding six putative and known glycosyltransferases (*POMT1*, *POMT2*, *POMGnT1*, *LARGE*, *FKTN*, and *FKRP*) have been shown to be responsible for approximately 50% of cases of these diseases, which are collectively referred to as secondary dystroglycanopathies<sup>4–11</sup> and which often feature brain abnormalities as well as muscular dystrophy. Despite recent advances in our understanding of the molecular mechanisms underlying secondary dystroglycanopathies, it remains unclear whether dystroglycan is the only target of these enzymes or whether other substrates contribute to the pathogenesis of these diseases.

Dystroglycan is known to be modified with several classes of oligosaccharides: N-glycans, mucin-type O-glycans, and O-mannosyl glycans.<sup>2</sup> The O-mannosyl glycans are thought to be responsible for dystroglycan's ligand-binding activity because the POMT1–POMT2 protein complex catalyzes the transfer of mannose to serine or threonine residues by means of an O-linkage.<sup>2</sup> We recently reported that maturation of  $\alpha$ -dystroglycan to its laminin-binding form requires a novel biosynthetic pathway involving phosphorylation on O-mannosyl glycans and that *LARGE* is crucial for further modification of phosphorylated O-mannosyl glycans on  $\alpha$ -dystroglycan.<sup>12</sup>

Here we report a missense mutation in the dystroglycan-encoding gene, *DAG1*, in a patient with limb-girdle muscular dystrophy and cognitive impairment. Our studies revealed that this substitution interferes with *LARGE*-dependent maturation of phosphorylated O-mannosyl glycans on  $\alpha$ -dystroglycan and leads to the disease-causing defect of  $\alpha$ -dystroglycan binding to laminin.

## METHODS

### CLINICAL INFORMATION

The patient initially received a diagnosis of limb-girdle muscular dystrophy with mental retardation.<sup>13</sup> Clinical data regarding this patient have been reported elsewhere.<sup>14</sup> (See also the Supplementary Appendix, available with the full text of this article at NEJM.org.) This study was approved by the institutional review board at Hacettepe University, and written informed consent was obtained from the patient's guardians.

### BIOCHEMICAL ANALYSIS

Biochemical analyses, including Western blotting, laminin-binding assays, and protein pull-down assays, were carried out as previously described.<sup>3,15</sup> (For details about the cell-culture and biochemical methods used, see the Supplementary Appendix.)

### MOUSE MODEL

Protocols for generating a knock-in mouse model harboring a mutation equivalent to that in our patient, as well as animal care and animal procedures, were approved by the University of Iowa Animal Care and Use Committee (see the Supplementary Appendix).

## RESULTS

We studied a Turkish patient with a relatively mild form of muscular dystrophy accompanied by cognitive impairment (see Case 2 in Dinçer et al.<sup>14</sup>). Although immunofluorescence analysis showed that  $\alpha$ -dystroglycan was hypoglycosylated in this

patient, no mutation was found in genes previously implicated in the pathogenesis of secondary dystroglycanopathies.<sup>14</sup> Since the dystroglycan-encoding gene, *DAG1*, is a plausible candidate gene, we investigated whether our patient harbored a causative mutation therein. Sequencing of the coding and noncoding regions of *DAG1* revealed a homozygous C-to-T missense mutation at position c.575, causing substitution of a methionine for a threonine at amino acid residue 192 (T192→M) of the protein (Fig. 1A). Both parents were heterozygous carriers of this mutated allele, and allele segregation in the family was consistent with recessive disease inheritance (Fig. 1B). The mutated threonine residue is located in the  $\alpha$ -dystroglycan N-terminal and is highly conserved across species (Fig. 1C).

We first assessed the effect of the newly identified  $\alpha$ -dystroglycan mutation in vitro, with respect to expression, post-translational processing, or both. To this end, we transiently expressed the full-length wild-type and T192→M dystroglycan proteins in dystroglycan-null myoblasts. Western blotting with an antibody that recognizes functionally glycosylated  $\alpha$ -dystroglycan (IIIH6)<sup>3,15</sup> revealed considerably lower immunoreactivity for T192→M than for wild-type dystroglycan when similar amounts of protein were expressed (Fig. 1D). Biotinylation analysis revealed that the cell-surface expression of T192→M and wild-type dystroglycan was similar (Fig. 1A in the Supplementary Appendix). Thus, in myoblasts, the T192→M substitution affects neither protein localization nor protein stability, but it does impair post-translational modification. A laminin-binding assay confirmed that the T192→M mutation considerably reduces the effectiveness of dystroglycan as a laminin receptor (Fig. 1D, and Fig. 1B and 1C in the Supplementary Appendix).

We next assessed the effect of the T192→M mutation on functional modification of  $\alpha$ -dystroglycan in vivo by introducing a T190→M mutation (which corresponds to the human T192→M mutation) into the mouse *Dag1* gene (Fig. 2 in the Supplementary Appendix). Histologic analysis revealed hallmarks of muscular dystrophy, such as centrally nucleated fibers, in the mutant mice (Fig. 2A, and Fig. 3A and 4 in the Supplementary Appendix). Immunofluorescence and biochemical analyses showed that the mutated  $\alpha$ -dystroglycan was expressed at the skeletal-muscle sarcolemma but that both ligand-binding activity and IIIH6 reactivity at this location were considerably decreased (Fig. 2B, and Fig. 3B and 5 in the Supplementary Appendix). Analysis of the uptake in the diaphragm of the membrane-impermeable Evans blue dye revealed disruption of the skeletal-muscle sarcolemma in the T190→M mice after exercise (Fig. 2C). The mutant mice also had marked impairment of exercise performance (e.g., reduced forelimb grip strength) (Fig. 2D). Mice heterozygous for the T190→M mutation did not have obvious muscle defects at the histologic level, despite a slight reduction in functional glycosylation of  $\alpha$ -dystroglycan (Fig. 4 and 6 in the Supplementary Appendix). Thus, the T190→M mutation caused skeletal-muscle abnormalities in mice that were consistent with those observed in our patient.

In addition to its role in skeletal muscle, dystroglycan is crucial for maintaining the structure and function of the brain and neuromuscular junctions.<sup>3,16–18</sup> Western blot and laminin-overlay analyses revealed that laminin-binding activity was significantly diminished in the brains of mice with the T190→M mutation (Fig. 3A). Morphologic analysis of the neuromuscular junctions revealed that the T190→M mutation interfered with their maturation; on staining of diaphragm samples with  $\alpha$ -Bungarotoxin conjugated with Alexa Fluor 488 (Invitrogen), which tightly binds acetylcholine receptors, the neuromuscular junctions in wild-type animals formed the expected pretzel-like structure, whereas those in the T190→M mice were round and immature (Fig. 3B and 3C, and Fig. 7 in the Supplementary Appendix). Consistent with impaired neuromuscular-junction maturation was the observation that the mutant mice had marked deficiencies in assays of neuromuscular function — for example, a reduction in retention time on a rotating-rod device (Fig. 3D). Furthermore, although no structural abnormality was evident in the brains

of the T190→M mice (an observation that was consistent with the results on magnetic resonance imaging [MRI] of the patient's brain), these mice had abnormal hind-limb claspings, a phenotype common to mouse models featuring neurologic impairment (Fig. 8 in the Supplementary Appendix). Collectively, these results provide strong evidence that the mutation in the patient caused neurologic impairment as well as muscular dystrophy as a consequence of impaired  $\alpha$ -dystroglycan post-translational modification.

Dystroglycan is also expressed in cardiac muscle, where its function as an extracellular-matrix receptor is important for limiting activity-induced myocardial damage.<sup>19</sup> Given that some patients with mutations in *FKTN* and *FKRP* have dilated cardiomyopathy,<sup>20,21</sup> we analyzed cardiac tissue from T190→M mice. We observed no obvious signs of any pathological abnormality, such as fibrosis or uptake of Evans blue dye after exercise (Fig. 9 in the Supplementary Appendix). Moreover, in contrast to the observed effects in brain and skeletal muscle, laminin-binding activity was affected only minimally in the hearts of T190→M mice (Fig. 3A). Thus, it appears that normal cardiac function is maintained in T190→M knock-in mice and that the effects of the mutation on functional modification of  $\alpha$ -dystroglycan are tissue-dependent.

Mutations in *LARGE* have been identified in patients with the Walker–Warburg syndrome or congenital muscular dystrophy type 1C, as well as in mice with myodystrophy (*Large<sup>myd</sup>*).<sup>9,10,22</sup> We recently found that *LARGE* is essential for phosphorylated O-mannosyl glycans to mature into a form capable of binding laminin.<sup>12</sup> We investigated the glycosylation status of T190→M  $\alpha$ -dystroglycan by subjecting the protein to treatment with cold aqueous hydrofluoric acid, which cleaves phosphoester linkages, and to inorganic metal-affinity chromatography, which captures monoester-linked phosphorylated compounds. Hydrofluoric acid treatment clearly reduced the molecular weight of T190→M  $\alpha$ -dystroglycan, and the beads on chromatography failed to capture T190→M  $\alpha$ -dystroglycan, although they readily bound  $\alpha$ -dystroglycan in samples from *Large<sup>myd</sup>* animals (Fig. 10 in the Supplementary Appendix). These results suggest that T190→M  $\alpha$ -dystroglycan bears phosphorylated O-mannosyl glycans but that these moieties are not sufficiently mature to confer laminin-receptor function on  $\alpha$ -dystroglycan.

Our findings suggest that the T192→M mutation affects *LARGE*-dependent modification of  $\alpha$ -dystroglycan. A previous study showed that the dystroglycan N-terminal serves as a binding site for *LARGE*, an interaction that is essential for  $\alpha$ -dystroglycan's laminin-binding activity.<sup>15</sup> An *in silico* model suggested that the side chain of the methionine residue could affect the protein surface structure<sup>23</sup> (Fig. 11A in the Supplementary Appendix); accordingly, we hypothesized that the T192→M mutation abrogates the dystroglycan–*LARGE* interaction owing to a conformational change in the dystroglycan N-terminal. A pull-down assay with Fc-tagged  $\alpha$ -dystroglycan mutants showed that although the wild-type protein bound to *LARGE*, variants harboring the T192→M mutation did not (Fig. 11B and 11C in the Supplementary Appendix). These findings support the notion that the T192→M mutation in dystroglycan dramatically impairs the interaction between  $\alpha$ -dystroglycan and *LARGE*, leading to incomplete post-translational modification of  $\alpha$ -dystroglycan.

## DISCUSSION

Although genes responsible for O-glycosylation-dependent muscular dystrophy (i.e., dystroglycanopathy) have been identified, the molecular mechanisms whereby mutations therein lead to impaired skeletal-muscle and brain function remain elusive — in part because defects in dystroglycan itself have not been reported. Here we describe such a case, which represents a new disease class (i.e., primary dystroglycanopathy). The T192→M mutation in this case caused deficiencies in  $\alpha$ -dystroglycan glycosylation and a marked reduction in  $\alpha$ -

dystroglycan's ability to bind extracellular-matrix components. Furthermore, our knock-in mouse model harboring the patient's mutation recapitulates the phenotype of secondary dystroglycanopathies in humans, supporting the view that dystroglycan is the main — and probably only — protein that is subject to the glycosylation abnormalities that cause muscular dystrophy.

Patients with dystroglycanopathy have a broad spectrum of disease phenotypes, with or without brain involvement. Limb-girdle muscular dystrophy with mental retardation was diagnosed in our patient according to the classification system established by Godfrey et al.<sup>13</sup> Biochemical findings from analyses of mouse T190→M show that laminin-binding activity is considerably reduced (Fig. 1 and 2), suggesting that residual glycosylation may account for the relatively mild muscle phenotype, as compared with the findings in patients with other congenital muscular dystrophies.

Our patient had severe cognitive impairment, yet MRI revealed no gross structural abnormality in the brain.<sup>14</sup> In more severe cases of human dystroglycanopathy, as well as in mice with a brain-specific *Dagl* knock-out, structural as well as functional brain abnormalities have been documented.<sup>3–6,9,10,16</sup> Thus, it may be that residual laminin binding by human T192→M dystroglycan is sufficient for the development of the cerebral layer but not for the establishment of normal synaptic physiology. It remains unclear how dystroglycan contributes to synaptic physiology; however, reports that synaptic components possessing laminin globular domains play a fundamental role in establishing and maintaining synaptic function<sup>24,25</sup> suggest that a reduction in the affinity of dystroglycan for synaptic proteins could be responsible for the neurologic phenotypes observed in the patient described here.

Our biochemical analyses show that the T192→M mutation impairs maturation of a specific modification of phosphorylated O-mannosyl glycans on  $\alpha$ -dystroglycan (Fig. 10 in the Supplementary Appendix). In the heart, T190→M  $\alpha$ -dystroglycan maintains substantial laminin-binding activity despite a reduction in molecular weight consistent with that in skeletal muscle and brain (Fig. 2 and 3A), and this remaining activity may contribute to preserved cardiac function (Fig. 8 in the Supplementary Appendix). The tissue variability in levels of  $\alpha$ -dystroglycan's functional glycosylation remains unexplained, but there are several possible explanations (e.g., tissue-specific protein complexes might increase the affinity between dystroglycan and LARGE, or the enzymatic activity of LARGE might be enhanced in heart tissue). Further investigation will be required to delineate the tissue-specific mechanisms ensuring that phosphorylated O-mannosyl residues of  $\alpha$ -dystroglycan mature into laminin-binding glycans.

In conclusion, we found a pathogenic missense mutation in *DAG1* that selectively impairs further modification of phosphorylated O-mannosyl glycans on  $\alpha$ -dystroglycan, leading to neuromuscular abnormalities. Our findings constitute evidence for the inclusion of *DAG1* in the list of genes whose mutations cause muscular dystrophy and cognitive impairment in humans.

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

## Acknowledgments

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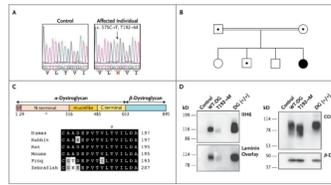
Diabetes and Digestive and Kidney Diseases (P30 DK 54759, to the University of Iowa Gene Transfer Vector Core), and the Muscular Dystrophy Campaign and Institute of Child Health Biomedical Research Centre (to Dr. Muntoni). Dr. Campbell is an investigator at the Howard Hughes Medical Institute.

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## References

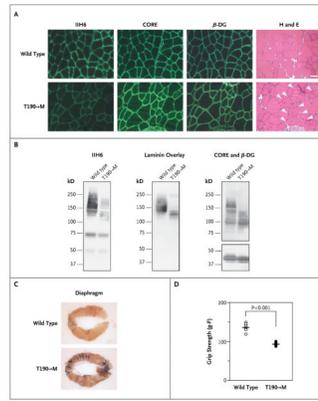
1. Ibraghimov-Beskrovnaya O, Ervasti JM, Leveille CJ, Slaughter CA, Sernett SW, Campbell KP. Primary structure of dystrophin-associated glycoproteins linking dystrophin to the extracellular matrix. *Nature*. 1992; 355:696–702. [PubMed: 1741056]
2. Barresi R, Campbell KP. Dystroglycan: from biosynthesis to pathogenesis of human disease. *J Cell Sci*. 2006; 119:199–207. [PubMed: 16410545]
3. Michele DE, Barresi R, Kanagawa M, et al. Post-translational disruption of dystroglycan-ligand interactions in congenital muscular dystrophies. *Nature*. 2002; 418:417–22. [PubMed: 12140558]
4. Beltrán-Valero de Bernabé D, Currier S, Steinbrecher A, et al. Mutations in the *O*-mannosyltransferase gene *POMT1* give rise to the severe neuronal migration disorder Walker-Warburg syndrome. *Am J Hum Genet*. 2002; 71:1033–43. [PubMed: 12369018]
5. van Reeuwijk J, Janssen M, van den Elzen C, et al. *POMT2* mutations cause  $\alpha$ -dystroglycan hypoglycosylation and Walker-Warburg syndrome. *J Med Genet*. 2005; 42:907–12. [PubMed: 15894594]
6. Yoshida A, Kobayashi K, Many H, et al. Muscular dystrophy and neuronal migration disorder caused by mutations in a glycosyltransferase, POMGnT1. *Dev Cell*. 2001; 1:717–24. [PubMed: 11709191]
7. Kobayashi K, Nakahori Y, Miyake M, et al. An ancient retrotransposal insertion causes Fukuyama-type congenital muscular dystrophy. *Nature*. 1998; 394:388–92. [PubMed: 9690476]
8. Brockington M, Blake DJ, Prandini P, et al. Mutations in the fukutin-related protein gene (*FKRP*) cause a form of congenital muscular dystrophy with secondary laminin  $\alpha 2$  deficiency and abnormal glycosylation of  $\alpha$ -dystroglycan. *Am J Hum Genet*. 2001; 69:1198–209. [PubMed: 11592034]
9. Longman C, Brockington M, Torelli S, et al. Mutations in the human *LARGE* gene cause MDC1D, a novel form of congenital muscular dystrophy with severe mental retardation and abnormal glycosylation of  $\alpha$ -dystroglycan. *Hum Mol Genet*. 2003; 12:2853–61. [PubMed: 12966029]
10. van Reeuwijk J, Grewal PK, Salih MA, et al. Intragenic deletion in the *LARGE* gene causes Walker-Warburg syndrome. *Hum Genet*. 2007; 121:685–90. [PubMed: 17436019]
11. Mercuri E, Messina S, Bruno C, et al. Congenital muscular dystrophies with defective glycosylation of dystroglycan: a population study. *Neurology*. 2009; 72:1802–9. [PubMed: 19299310]
12. Yoshida-Moriguchi T, Yu L, Stalnakier SH, et al. *O*-mannosyl phosphorylation of  $\alpha$ -dystroglycan is required for laminin binding. *Science*. 2010; 327:88–92. [PubMed: 20044576]
13. Godfrey C, Clement E, Mein R, et al. Refining genotype phenotype correlations in muscular dystrophies with defective glycosylation of dystroglycan. *Brain*. 2007; 130:2725–35. [PubMed: 17878207]
14. Dinçer P, Balci B, Yuva Y, et al. A novel form of recessive limb girdle muscular dystrophy with mental retardation and abnormal expression of  $\alpha$ -dystroglycan. *Neuromuscul Disord*. 2003; 13:771–8. [PubMed: 14678799]
15. Kanagawa M, Saito F, Kunz S, et al. Molecular recognition by LARGE is essential for expression of functional dystroglycan. *Cell*. 2004; 117:953–64. [PubMed: 15210115]
16. Moore SA, Saito F, Chen J, et al. Deletion of brain dystroglycan recapitulates aspects of congenital muscular dystrophy. *Nature*. 2002; 418:422–5. [PubMed: 12140559]

17. Côté PD, Moukhles H, Lindenbaum M, Carbonetto S. Chimaeric mice deficient in dystroglycans develop muscular dystrophy and have disrupted myoneural synapses. *Nat Genet.* 1999; 23:338–42. [PubMed: 10610181]
18. Nishimune H, Valdez G, Jarad G, et al. Laminins promote postsynaptic maturation by an autocrine mechanism at the neuromuscular junction. *J Cell Biol.* 2008; 182:1201–15. [PubMed: 18794334]
19. Michele DE, Kabaeva Z, Davis SL, Weiss RM, Campbell KP. Dystroglycan matrix receptor function in cardiac myocytes is important for limiting activity-induced myocardial damage. *Circ Res.* 2009; 105:984–93. [PubMed: 19797173]
20. Murakami T, Hayashi YK, Noguchi S, et al. Fukutin gene mutations cause dilated cardiomyopathy with minimal muscle weakness. *Ann Neurol.* 2006; 60:597–602. [PubMed: 17036286]
21. Mercuri E, Brockington M, Straub V, et al. Phenotypic spectrum associated with mutations in the fukutin-related protein gene. *Ann Neurol.* 2003; 53:537–42. [PubMed: 12666124]
22. Grewal PK, Holzfeind PJ, Bittner RE, Hewitt JE. Mutant glycosyltransferase and altered glycosylation of  $\alpha$ -dystroglycan in the myodystrophy mouse. *Nat Genet.* 2001; 28:151–4. [PubMed: 11381262]
23. Bozic D, Sciandra F, Lamba D, Brancaccio A. The structure of the N-terminal region of murine skeletal muscle  $\alpha$ -dystroglycan discloses a modular architecture. *J Biol Chem.* 2004; 279:44812–6. [PubMed: 15326183]
24. Südhof TC. Neuroligins and neuexins link synaptic function to cognitive disease. *Nature.* 2008; 455:903–11. [PubMed: 18923512]
25. Sato S, Omori Y, Katoh K, et al. Pikachurin, a dystroglycan ligand, is essential for photoreceptor ribbon synapse formation. *Nat Neurosci.* 2008; 11:923–31. [PubMed: 18641643]



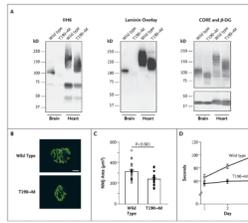
**Figure 1. Identification of a DAG1 Missense Mutation in a Woman with Limb-Girdle Muscular Dystrophy, and In Vitro Biochemical Characterization of the Encoded Protein**

In Panel A, electrochromatograms reveal sequence variation in *DAG1*. A homozygous mutation, c.575C→T (arrow), leads to an amino acid change, T192→M. The T192→M variant was not detected in any of the 100 Turkish control subjects tested (200 chromosomes). Panel B shows the family pedigree. Squares denote male family members, and circles female family members; the double line indicates a consanguineous marriage; the black circle denotes the affected woman, and dots indicate heterozygous carriers. Panel C shows a schematic representation of a dystroglycan mutant protein (top). Human  $\alpha$ -dystroglycan is composed of a signal peptide (SP) (amino acids 1 through 29), an N-terminal domain (amino acids 30 through 316), a mucinlike domain (amino acids 317 through 485), and a C-terminal domain (amino acids 486 through 653). The position of the patient's mutation (T192→M) is indicated by an asterisk. Amino acid residues are aligned flanking the codon affected by the T192→M missense mutation (bottom). Among these residues, black boxes denote identity and gray boxes similarity. Panel D shows a biochemical analysis of wild-type (WT-DG) and T192→M dystroglycan in dystroglycan-null myoblast cells. Functional modification of  $\alpha$ -dystroglycan was evaluated by Western blotting with an antibody (IIH6) that recognizes only the glycosylated form of  $\alpha$ -dystroglycan and by laminin-overlay assay. Overall expression of wild-type dystroglycan and T192→M dystroglycan was detected with the use of antibodies against the  $\alpha$ -dystroglycan protein core (CORE) and antibodies against  $\beta$ -dystroglycan ( $\beta$ -DG).



**Figure 2. Biochemical and Physiological Skeletal-Muscle Phenotypes of Disease in Gene-Targeted Mice with the Same Dystroglycan Mutation as the Patient**

Panel A shows immunofluorescence and histologic analyses of T190→M skeletal-muscle (iliopsoas) sections prepared when the animals were 21 weeks of age. Serial sections were stained with antibodies against glycosylated  $\alpha$ -dystroglycan (IHH6), the  $\alpha$ -dystroglycan core (CORE), and  $\beta$ -dystroglycan ( $\beta$ -DG). Histologic abnormalities in the sections were evaluated by means of hematoxylin and eosin (H and E) staining. White arrowheads indicate centrally nucleated fibers (scale bar, 50  $\mu$ m). Panel B shows a representative biochemical analysis of wild-type (WT) and T190→M  $\alpha$ -dystroglycan isolated from wheat-germ agglutinin-enriched homogenates of skeletal muscle. Both Western blotting and laminin-overlay assays were carried out. Functional modification of  $\alpha$ -dystroglycan was impaired by the T190→M mutation *in vivo*, as shown by the dramatic reduction in IHH6 immunoreactivity and laminin binding in the absence of major differences in CORE and  $\beta$ -DG immunoreactivity. Panel C shows the uptake of Evans blue dye in the diaphragm after exercise in wild-type and T190→M mice at 8 weeks of age. Panel D shows the whole-mouse grip-strength measurements, in gram-force (g-F), for the T190→M mice and their wild-type littermates. The P value was obtained with the use of Student's t-test.



### Figure 3. Neuromuscular Phenotypes in Mice with the Dystroglycan Mutation

Panel A shows functional modification of  $\alpha$ -dystroglycan in the brain and heart of mice with the T190→M mutation. Antibody staining and laminin-overlay assays were performed on blots of wheat-germ agglutinin-enriched homogenates from the brain and heart of T190→M mice and their wild-type (WT) littermates. Panels B and C show defects in maturation of the neuromuscular junction (NMJ) in the T190→M mice. Panel B shows representative images of the NMJ in the diaphragm; specimens were stained, with  $\alpha$ -Bungarotoxin labeled with Alexa Fluor 488 (scale bar, 10  $\mu$ m). Panel C shows individual data (circles) and mean ( $\pm$ SE) values for the NMJ area in wild-type and T190→M mice. Panel D shows the mean ( $\pm$ SE) values of five rotating-rod trials performed to assess neurologic defects in the T190→M mice. The time that the mice remained on the rotating rod was significantly longer for the wild-type mice than for the T190→M mice ( $P < 0.001$  by one-way analysis of variance).