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1 Title: *Comamonas kerstersii* bacteremia in a patient with diverticulosis

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21

22 **ABSTRACT**

23 We report for the first time a case of bacteremia caused by *Comamonas kerstersii* in a 65-year-old patient
24 with sign of diverticulosis. In addition, we review the isolation of *Comamonas* sp. and related organisms
25 in our hospital over 25 years.

26

27 **CASE REPORT**

28 *Comamonas kerstersii* is a non-fermenting β -Proteobacteria described in 2003 that has long been
29 considered as non-pathogenic (19). This organism has been recently associated with intra-abdominal
30 infection consecutive to perforation of the digestive tract (1). Herein, we describe a case of polymicrobial
31 bacteremia due to *Comamonas kerstersii* and *Bacteroides fragilis* in a 65-year-old diabetic man that was
32 admitted to the emergency department of the hospital due to sudden onset of fever and chills. The patient
33 reported episodes of vomiting and diarrhea and mentioned that he drank water from a small river. Stool
34 cultures did not disclose *Salmonella*, *Shigella*, *Aeromonas* or *Campylobacter* species. The detection of
35 *Clostridium difficile* toxin A and B and glutamate dehydrogenase antigen was also negative. Blood
36 cultures (two pairs of bottles) were drawn from a peripheral vein and the patient was discharged under
37 treatment with oral ciprofloxacin for a gastroenteritis of unknown origin. The Blood cultures were
38 processed into a BACTEC FX automated blood culture system (Becton Dickinson, Sparks, MD). A first
39 aerobic blood culture bottle became positive and the Gram staining revealed the presence of long
40 filamentous Gram-negative bacilli (Figure 1). The bacterial identification by MALDI-TOF (Bruker
41 Daltonics GmbH, Leipzig, Germany) analysis was performed the same day using a protocol that we
42 recently developed based on the analyses of a bacterial pellet preparation from the blood bottles (5, 16,
43 17). The strain was identified as *Comamonas kerstersii*, a Gram-negative non-fermentative bacterium, and
44 prompted the hospitalization of the patient. The patient was afebrile at that time, but palpation of the left
45 lower abdominal quadrant was painful. An abdominal CT scan revealed diverticulosis without evidence
46 of diverticulitis. Consecutively, the anaerobic blood bottles from the same pair became positive for

47 *Bacteroides fragilis*. We monitored the following MIC ($\mu\text{g.ml}$) for the *Comamonas kerstersii* strain:
48 ceftazidime, 0.75; meropenem, 0.004; minocycline, 0.38; levofloxacin, 4; co-trimoxazole, >32;
49 ciprofloxacin, 32. For the *Bacteroides fragilis* strain, the MIC were: amoxicillin-clavulanate, 1.5;
50 piperacillin-tazobac, 6; imipenem, 0.12; meropenem, 0.025; metronidazole, 0.25; clindamycin, 16;
51 ciprofloxacin, 32. A treatment of imipenem-cilastatine was given for 10 days and the patient recovered.
52 The final diagnosis was a mixed bacteremia with *Comamonas kerstersii* and *Bacteroides fragilis* in the
53 setting of a diverticulosis.

54

55 Comamonads are Gram negative, non-fermentative bacteria, oxydase- and catalase-positive, largely
56 motile due to the presence of polar flagella. The *Comamonas* genus originally contained *Comamonas*
57 *terrigena*, *Comamonas testosteroni* (previously *Pseudomonas testosteroni*) and *Comamonas acidovorans*
58 (previously *Pseudomonas acidovorans*) (6). It now contains seventeen species while *Comamonas*
59 *acidovorans* has been separated from the *Comamonas* genus on the basis of 16S rRNA and is now known
60 as *Delftia acidovorans* (20). Although ubiquitously distributed in the environment (soil and water),
61 *Comamonas* and *Delftia* sp. are rarely associated with infections in humans. However, several
62 publications have incriminated *Comamonas testosteroni* and *Delftia acidovorans* in particular in human
63 diseases, including severe invasive infections such as bacteremia and meningitis (2-4, 7, 10, 11, 20).

64 *Comamonas kerstersii*, described in 2003 (19), has recently been reported as an agent of intra-abdominal
65 infection by Almuzara and colleagues (1). The present case is the first report of *Comamonas kerstersii*
66 bacteremia. We initially performed the identification of the strain at the species level directly from the
67 positive blood bottle using MALDI-TOF, with a spectral score of 2.176 (5, 16, 17). Subsequently, it was
68 recovered both from the blood agar plate (with a spectral score of 2.26), on which the growth was
69 maximal and from the “chocolate” agar plate that is supplemented with NAD (factor V) and hemin (factor
70 X). We also proceeded to the amplification and sequencing of the 16S rRNA ribosomal gene in order to
71 confirm the MALDI-TOF identification of this strain (8). The analysis of the sequences using the BLAST

72 V2.0 software (<http://www.ncbi.nlm.nih.gov/BLAST/>) showed 100% of identity with the sequences
73 corresponding to the 16S RNA ribosomal gene of *Comamonas kerstersii* strain LMG 5323 (19). From
74 both the blood bottle and the agar plates, the strain appeared as an extremely long Gram-negative
75 filamentous bacillus which is a very unusual phenotype for bacteria of this genus (Figure 1). The
76 *Comamonas* and *Delftia* strains previously isolated in our hospital are Gram-negative short bacilli or rods
77 (Figure 1), which is the morphology described for these organisms (19, 20).

78 Translocation from the digestive tract seems to be a predominant cause of infections by *Delftia* and
79 *Comamonas* species. Recently, Hagiya and colleagues reported a *Delftia acidovorans* bacteremia in a 46-
80 year-old woman caused by translocation of the bacteria consecutive to pesticide poisoning (10). A
81 bacteremia caused by *Comamonas testosteroni* was previously reported, in a 22-year-old man with
82 perforated appendix (9). In the four cases reported by Almuzara and colleagues, the *Comamonas*
83 *kerstersii* strains were isolated from intra-abdominal collections (1). We previously identified another
84 *Comamonas kerstersii* strain in an intra-peritoneal collection of an 11-year-old child with a perforated
85 appendix (table 1 and figure 1). Herein, the digestive origin of the *Comamonas kerstersii* strain is
86 supported by the fact that: i) the patient reported abdominal pain, vomiting and diarrhea, ii) the CT scan
87 revealed evidence of diverticulosis, and iii) the enteric bacteria *Bacteroides fragilis* was isolated from the
88 blood culture in this setting. The infection could originate from the water that the patient drank in the
89 countryside.

90 *Comamonads* have been rarely associated with infection in humans despite their ubiquitous distribution in
91 the environment possibly due to the difficulty to accurately distinguish *Comamonas* species from
92 *Pseudomonas* species in the pre-MALDI-TOF area (1). Alternatively, *Comamonads* could have been
93 under-recognized due to their common occurrence in the setting of a polymicrobial infection. In our 1027-
94 bed tertiary care university hospital, thirty-three *Comamonas* sp. strains and thirty-eight *Delftia*
95 *acidovorans* strains were isolated from 1997 to 2013. They were primarily isolated from respiratory
96 samples (33%), urogenital samples (23%) and digestive samples (21%); bacteremia represented 5% (three

97 patients) of all cases (Table 1). All 3 cases were poly-microbial bacteremia. The first bacteremia case
98 was due to *Comamonas testosteroni* in association with *Streptococcus parasanguis* and *Ralstonia pickettii*
99 in a 33-year old man. A second case involved *Delftia acidovorans* in association with *Streptococcus*
100 *agalactiae* in blood cultures from a 61-year-old man. The last case is the present *Comamonas kerstersii*
101 and *Bacteroides fragilis* co-infection.

102 Like *Delftia acidovorans*, *Comamonas testosteroni* is the *Comamonas* species predominantly associated
103 with bacteremia (table 1) (7, 9, 10, 20). Translocation from the digestive tract and catheters are the
104 predominant source of infection (9, 10, 13-15). Children or patients with compromised immune systems
105 (AIDS or patients treated with chemotherapies) appear to be particularly at risk to develop *Comamonas*
106 sp. or *Delftia acidovorans* bacteremia (12, 13). Interestingly, Khan and colleagues reported a fatal
107 outcome in an 4-year old immuno-competent child presenting a *Delftia acidovorans* bacteremia (12). The
108 patient presented herein did not display any sign of immunodeficiency suggesting that such bacteremia
109 may also occur in the absence of immunosuppression. The likely high inoculum in the water that was
110 drunk and the diabetic status of the patient are two significant co-factors that may explain the occurrence
111 of a bacteremia in the setting of a gastro-intestinal infection. Similarly, a *Comamonas* species bacteremia
112 has been associated with exposure to possibly contaminated water of a fish tank (18).

113 This report reveals that *Comamonas kerstersii* and other non-fermenting related bacteria may be
114 involved in severe diseases independently of perforation of the digestive tract. Moreover, this report
115 highlights the usefulness of MALDI-TOF in redefining the epidemiology and clinical syndromes due to
116 some non-fermentative Gram negative bacteria that were difficult to identify in the pre-MALDI-TOF
117 area.

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183 **Figure legend:**

184 **Figure 1: Gram staining of *Comamonas kerstersii*, *Comamonas testosteroni* and *Delftia acidovorans***
185 **strains isolated in clinical samples from Lausanne University Hospital.** (A) The *Comamonas*
186 *kerstersii* strain of the present case report directly from the blood-cultures bottle or B after culture on
187 blood agar medium. The strain displays long filaments when compared to the other strains that appears as
188 Gram-negative short bacilli or rods. C) A *Comamonas kerstersii* strain identified in an intra-peritoneal
189 collection of an 11-year-old child with a perforated appendix. D) *Comamonas testosteroni* involved in a
190 bacteremia in a 33-year- old man. E) *Delftia acidovorans* identified in blood cultures from a 61-year-old
191 man.

Table 1: *Comamonas* sp. and *Delfia acidovorans* isolated from clinical samples in the Lausanne University Hospital from 1997 to 2013.

| | Respiratory and ENT ^a | Urogenital ^b | Intra-abdominal ^c | Skin | Blood culture | Surgical wound | Others ^d | Total patient (samples) | % of patient |
|--|----------------------------------|-------------------------|------------------------------|-------------|-----------------|----------------|---------------------|-------------------------|--------------|
| <i>Delfia acidovorans</i> | 19 (20) | 12 (13) | - | 5 | 1 (13) | - | 1 (4) | 38 (55) | 54.3 |
| <i>Comamonas testosteroni</i> | 2 (4) | 3 | 8 | 1 (4) | 1 | 4 (5) | 1 | 20 (26) | 28.57 |
| <i>Comamonas kerstersii</i> | - | - | 1 | - | 1 | - | - | 2 | 2.86 |
| <i>Comamonas aquatica</i> | - | 1 (4) | - | - | - | - | - | 1 (4) | 1.43 |
| <i>Comamonas specis</i> ^e | 2 | - (2) | 6 (8) | 1 | - | - | - (1) | 9 (14) | 12.86 |
| Number of patients (number of samples) | 23 (26) | 16 (22) | 15 (17) | 7 (10) | 3 (15) | 4 (5) | 2 (6) | 70 (101) | 100 |
| Percentage of patients (percentage of samples) | 32.86 (25.74) | 22.86 (21.78) | 21.43 (16.83) | 10 (9.9) | 4.29 (14.85) | 5.71 (4.95) | 2.85 (5.94) | 100 | |

^aEar nose and throat

^bUrine, vaginal swab and placenta

^cascitic fluid, peritoneal fluid, penrose liquid, kehr drain

^dstools (1), bone fragment (1), orifice smear (1)

^eno identification at the species level

Number represent patients, brackets represent the number of samples

