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# THE ROYAL SOCIETY

# Natural alleles at the *Doa* locus underpin evolutionary changes in *Drosophila* lifespan and fecundity

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'Evolve and resequence' (E&R) studies in Drosophila melanogaster have identified many candidate loci underlying the evolution of ageing and life history, but experiments that validate the effects of such candidates remain rare. In a recent E&R study we have identified several alleles of the LAMMER kinase Darkener of apricot (Doa) as candidates for evolutionary changes in lifespan and fecundity. Here, we use two complementary approaches to confirm a functional role of *Doa* in life-history evolution. First, we used transgenic RNAi to study the effects of Doa at the whole-gene level. Ubiquitous silencing of expression in adult flies reduced both lifespan and fecundity, indicating pleiotropic effects. Second, to characterize segregating variation at Doa, we examined four candidate single nucleotide polymorphisms (SNPs; Doa-1, -2, -3, -4) using a genetic association approach. Three candidate SNPs had effects that were qualitatively consistent with expectations based on our E&R study: Doa-2 pleiotropically affected both lifespan and late-life fecundity; Doa-1 affected lifespan (but not fecundity); and Doa-4 affected late-life fecundity (but not lifespan). Finally, the last candidate allele (Doa-3) also affected lifespan, but in the opposite direction from predicted.

# 1. Background

Evolve & resequence' (E&R) studies, combining experimental evolution experiments with whole-genome sequencing, have emerged as a powerful method for identifying the genetic basis of evolutionary change [1–5]. In the *Drosophila melanogaster* model, for example, E&R studies have been successfully used to identify candidate loci underlying thermal adaptation [6,7], the evolution of developmental rate [8], body size [9], hypoxia tolerance [10,11], courtship song [12], lifespan and late-life fertility [13–15], dietary metabolism [16], pathogen resistance [17,18], egg size [19], desiccation resistance [20], starvation resistance [21] and factorial selection on multiple life-history traits [22], among others.

Despite the identification of many putatively adaptive loci in such E&R studies, experimental assays that validate the presumed functional effects of such candidates are rare, which remains a major challenge for current tests of adaptation at the genetic level [1–3,23]. Here, we examine the putative life-history effects of a candidate locus, *Darkener of apricot (Doa)*, and associated candidate alleles that we have previously identified in an E&R experiment on longevity and late-life fertility in *D. melanogaster* [22]. *Doa*, a member of the LAMMER kinases, is known to phosphorylate a wide range of substrates and to be involved in many biological functions, such as embryonic development, oocyte formation, somatic sex determination, courtship behaviour and oxidative stress resistance (see Supplementary file 1 for additional information) [24–30]. Interestingly, *Doa* has been identified as a promising life-history candidate locus in several independent E&R experiments and genome-wide association studies (GWAS) on lifespan and late-life fertility, egg volume and ovariole number [15,19,22,31–33]. These

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findings make *Doa* a prime candidate locus for further study. Importantly, while several gene functions of *Doa* have been well established in molecular genetics studies (see above), the putative effects upon fitness components of naturally occurring polymorphisms at *Doa* have not yet been assessed.

In our experiments, we used two complementary approaches to investigate the functional role of Doa in affecting two major fitness components that had evolved in our E&R study: lifespan and fecundity [22,34]. First, we used ubiquitous adult-specific RNAi silencing to examine the overall effects of Doa on lifespan and fecundity. Similar to using null mutants (amorphic mutations), this type of functional test is aimed at understanding function at the level of the whole gene; it can potentially reveal the complete phenotypic effects of a candidate gene, including any pleiotropic (and potentially deleterious) functions that it might have (e.g. [35,36]). However, given that specific alleles or mutations of a pleiotropic gene may differ in the functions they affect (see [35-37]), we next investigated segregating variation at Doa by examining four candidate single nucleotide polymorphisms (SNPs) using a genetic association approach based on 'Mendelian randomization'. Beyond confirming a role of the Doa locus in life-history adaptation, our results illustrate key differences in the effects of the four investigated SNPs on fecundity and longevity.

## 2. Material and methods

#### (a) Transgenic RNAi

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Transgenic in vivo RNAi was performed using the mifepristone (RU486)-inducible GeneSwitch(GS)-GAL4 system in combination with Doa UAS-RNAi constructs [38]. The GS system allows us to drive the expression of Doa UAS-RNAi during the adult stage only, thereby avoiding potential developmental carry-over effects. Importantly, this system makes it possible to compare the effects of RNAi (i.e. the application of the drug, resulting in RNAi-mediated knockdown) with negative control (i.e. no application of the drug) within the same transgenic genotype, thus providing the most robust control possible with regard to genotype. We used the ubiquitously expressing daughterless (da)-GeneSwitch(GS)-GAL4 construct [39] (courtesy of Véronique Monnier, Paris) to drive the expression of two independent Doa UAS-RNAi lines, obtained from the Vienna Drosophila RNAi Center (VDRC) (#19066 [D19]; #102520 [D10]), with construct insertions on chromosomes 3 and 2, respectively, thus controlling for potentially confounding effects of insertion position. These constructs target the catalytic domain of Doa that is shared by all isoforms.

All lines were kept and assays were performed at 25°C, 65% humidity and 12h:12 h light:dark cycle, on a commeal-yeast–sucrose–agar medium (per 1 L of food: 7 g agar, 50 g sucrose, 50 g cornmeal, 50 g yeast, 6 ml propionic acid, 10 ml of a 20% nipagin stock solution). After emergence, flies were kept on medium containing either 100 or 200  $\mu g \ ml^{-1}$  mifepristone (233 or 466  $\mu M$ , respectively) dissolved in ethanol, or on control medium (i.e. containing ethanol without mifepristone). Note that increasing levels of mifepristone can induce higher levels of gene knockdown, in a dose-dependent fashion. We confirmed that both RNAi lines resulted in a significant knockdown of Doa expression using quantitative real-time PCR (qRT-PCR) (see electronic supplementary material, File S2 for details).

Mifepristone concentrations of up to 200  $\mu g \, ml^{-1}$  have previously been used without detrimental effects on survival of adult flies [15,39,40]. To confirm this, we tested the effect of 100 and 200  $\mu g \, ml^{-1}$  mifepristone on lifespan and fecundity of F1 flies of a cross of *da*-GS-GAL4 females with males of the

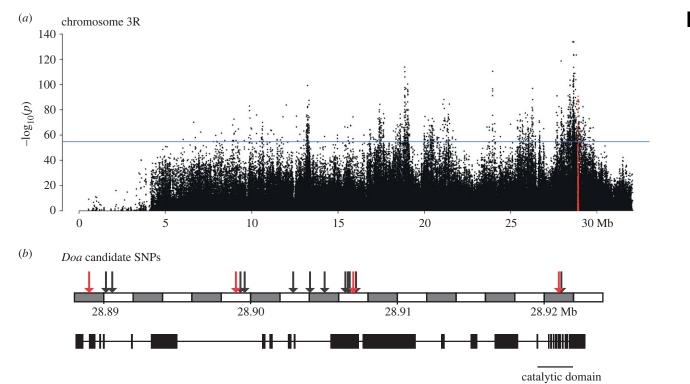
isogenic progenitor strain for the VDRC (GD) RNAi library strains,  $w^{1118}$  (#60000). We did not observe any confounding deleterious effects of these concentrations on the phenotypes of interest (electronic supplementary material, File S3).

To assess the effect of RNAi directed against on Doa on lifespan, cohorts of F1 offspring between crosses of da-GS-GAL4 virgin females and males carrying one of the two UAS-RNAi constructs or the isogenic control strain were collected within a 24 h window. Flies were sexed under mild CO<sub>2</sub> exposure and transferred to 1L demography cages with food vials (with 0, 100 or 200 µg ml<sup>-1</sup> mifepristone) attached to the cages. For each genotype and mifepristone concentration, we set up three replicate cages, each containing 75 flies per sex. Dead flies were scored and fresh food was provided every two days. Differences in lifespan between mifepristone-induced RNAi and uninduced controls were analysed in R (v. 3.3.1) using mixedeffects Cox (proportional hazards) regression with mifepristone concentration, sex and their interaction as fixed effects and with 'replicate cage' as a random effect, using the R package coxme (v. 2.2-5).

The effect of Doa RNAi on fecundity was assessed by measuring daily egg production of females. Virgin females were collected from the F1 offspring of da-GS-GAL4 females and males carrying one of the two UAS-RNAi constructs or the isogenic control strain. After 24 h, two virgin females and two  $w^{111\bar{8}}$  males were placed together in vials with either 0 (i.e. control) or 200 µg ml<sup>-1</sup> mifepristone. Ten replicate vials were prepared per genotype and mifepristone concentration. Flies were left for 48 h to ensure mating and consumption of mifepristone before the start of the experiment; after this period, they were transferred to fresh vials with food (with 0 or 200 µg ml<sup>-1</sup> mifepristone, respectively) to lay eggs for 24 h. The numbers of eggs laid by each pair of females were counted under a dissecting microscope; daily egg production was measured for nine consecutive days. We calculated average fecundity per female over 3 days in order to average out day-today variation in egg laying. Fecundity data were analysed in R (v. 3.3.1) using generalized linear mixed models with a Poisson distribution and with mifepristone concentration, with day and their interaction as fixed effects and with 'replicate vial' as a random effect using the R package lme4 (v. 1.1-13). Exposure to mifepristone of the control line did not cause adverse (and thus confounding) effects on lifespan and fecundity (see electronic supplementary material, File S3)

#### (b) SNP association study

To examine whether the four experimentally candidate SNPs at Doa affect lifespan and fecundity (see below for details of SNP identification), we performed a genetic (SNP) association study, based on 'Mendelian randomization' [41-43]. Mendelian randomization (MR) approaches aim to identify putative causal effects of candidate loci by testing the alternative allelic states in a genetically diverse background to limit the impact of potentially confounding (epistatic) factors in the genetic background. A critical factor for the reliability of MR approaches is the lack of linkage disequilibrium (LD) between the focal locus and other loci in the genetic background. Here, we used strains from the Drosophila Genetic Reference Panel (DGRP [44]; obtained from the Bloomington Drosophila Stock Center [BDSC]), which provides ample natural genetic variation for MR. As LD typically decays very rapidly in D. melanogaster, within a few hundred base pairs or so [44], the MR approach is expected to provide information on the functional impact of an individual candidate SNP, with little or no confounding effects of other (Doa) SNPs. To confirm this, we analysed LD (measured by pairwise  $r^2$ ) among all polymorphic *Doa* SNPs (minor allele frequency  $\geq$  0.1) in the complete panel of DGRP lines, as done by [43] before (see electronic supplementary material, File S4).



**Figure 1.** Overview of the experimentally evolved *Doa* SNPs. (*a*) The Manhattan plot demonstrates the position of *Doa* on chromosome 3R in the Evolve and resequence study by Hoedjes *et al.* [22]. *Doa* (all SNPs within the gene are highlighted) is located within a relatively large region at the end of chromosome 3 that had been under selection. The horizontal line (at  $-\log_{10}(54.9)$ ) indicates the threshold for significant candidate SNPs (false discovery rate (FDR) = 0.0005). (*b*) The gene structure of *Doa* with the 16 significant candidate SNPs (at FDR < 0.0005; indicated with arrows) that were identified as candidates for ageing and reproduction. Arrows that have been highlighted indicate the four SNPs that have been investigated functionally in this study. Coordinates according to reference genome v6.

For each of the four candidate nucleotide positions at Doa, we randomly selected 20 distinct DGRP lines that were fixed for the SNP allele that was previously identified as being the major allele in the short-lived, early reproduction experimental evolution lines (control lines; see [22]). Similarly, we randomly selected 20 lines fixed for the SNP allele that was found to be the major allele in the long-lived, late-reproduction experimental evolution lines (see electronic supplementary material, table S3 for details of the crosses). SNP genotypes were confirmed by sequencing a small genomic region surrounding each SNP using Sanger sequencing. For each allelic state ('short-lived' versus 'long-lived') and nucleotide position, we generated 10 unique F1 crosses, each cross being made from a different pair of distinct DGRP lines sharing the same SNP state (i.e. virgin females of one strain crossed to males from the other strain); because the DGRP lines are inbred, this was done to minimize potentially confounding homozygous effects at non-candidate loci in the genomic background. Thus, for each of the four candidate nucleotide positions and for each alternative allele ('short-lived' versus 'long-lived') we had a 10-fold replicated panel of independent F1 genotypes fixed for a given SNP allele but maximally heterozygous at other genomic positions (for details see the overview in electronic supplementary material, table S3). In the end, due to the low viability of some DGRP lines and F1 crosses, we phenotyped between 8–10 F1 crosses per candidate SNP and allelic state. To evaluate whether any other positions in the genome, besides the candidate SNP allele, were highly differentiated between the four pairs of panels, we calculated SNP-wise  $F_{\rm ST}$ based on the method of Weir and Cockerham [45], using the pooled genome sequence information per panel [42] (electronic supplementary material, File S5).

F1 crosses were reared and assays performed at 25°C, 65% humidity and 12 h:12 h light:dark cycle, on a commeal-yeast-sucrose-agar medium, as described above. Lifespan was measured using demography cages, as above. Flies that had emerged within a 24 h window were collected, and for each F1

cross 75 males and 75 females were placed in a single demography cage. Differences in lifespan between the two allelic states of each SNP were analysed in R (v. 3.3.1) using mixed-effects Cox (proportional hazards) regression with allele, sex and their interaction as fixed effects and with 'F1 cross' as a random effect using the R package coxme (v. 2.2-5).

Fecundity was measured over a period of 30 days after eclosion in order to provide insight into early (peak) and late (post-peak) fecundity. Flies that had eclosed within a 24-h window were collected for crosses; for each F1 cross, two females and two males were placed in a vial containing regular medium, with three replicate vials per F1 cross. Every third day (i.e. on days 3, 6, 9, 12, 15, 18, 21, 24, 27 and 30), the number of eggs laid by each pair of females during a 24-h period was determined using a dissecting microscope. Fecundity was analysed using generalized linear mixed models with a Poisson distribution in R (v. 3.3.1), with allelic state, day and their interaction as fixed effects and 'replicate vial' as a random effect using the R package Ime4 (v. 1.1-13).

#### 3. Results and discussion

# (a) Evolutionary changes at *Doa* might underpin life-history evolution

We previously identified *Doa* as a life-history candidate locus in an E&R experiment in which fruit flies were selected for late-life fecundity and where a longer lifespan evolved as a correlated response [22]. In that study, *Doa* was identified as one of multiple candidate loci under selection (figure 1a). Genetic tests of candidate genes and SNPs are critical to distinguish the putative causal loci from false positive signals and to determine if they have an effect on either one or

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**Table 1.** Details on the *Doa* candidate SNPs. The genomic location of the four *Doa* SNPs that were investigated functionally is shown. For each SNP, a significant difference in allele frequencies was observed between lines selected for early age-at-reproduction (control) versus lines selected for late-life (postponed) reproduction (which associated with an evolutionary increase in lifespan: 'long-lived').

			allele frequency			
name	location	feature	control	long-lived	control allele	'long-lived' allele
Doa-1	3R:28'888'916	intronic	0.18	0.52	С	T
Doa-2	3R:28'898'997	intronic	0.43	0.80	Α	G
Doa-3	3R:28′907′179	missense	0.55	0.90	A	G
Doa-4	3R:28'921'024	synonymous	0.40	0.78	С	T

**Table 2.** Statistical tests of the effects of *Doa* RNAi on lifespan and fecundity (egg-laying rate). Two independent RNAi constructs (D19 and D10) that both target the catalytic domain of *Doa* were used in combination with the mifepristone-inducible GeneSwitch-GAL4 > UAS system. A strain with the same genetic background as the two constructs ( $w^{1178}$ ) was used as control for adverse effects of mifepristone application. Significant effects are indicated in bold.

longevity			fecundity			
	χ²	p		χ²	p	
w <sup>1118</sup> (control)			w1118 (control)			
mifepristone	8.64	0.071	mifepristone	7.31	0.063	
sex	95.68	$< 2.2 \times 10^{-16}$	day	50.83	$2.4 \times 10^{-10}$	
interaction	3.63	0.163	interaction	3.59	0.166	
D10			D10			
mifepristone	51.53	$1.7 \times 10^{-10}$	mifepristone	1.64	0.6512	
sex	83.56	$< 2.2 \times 10^{-16}$	day	34.12	$7.0 \times 10^{-7}$	
interaction	37.29	$8.0 \times 10^{-9}$	interaction	1.61	0.446	
D19			D19			
mifepristone	317.24	<2.2 × 10 <sup>-16</sup>	mifepristone	320.54	$< 2.2 \times 10^{-16}$	
sex	383.21	<2.2 × 10 <sup>-16</sup>	day	508.72	$< 2.2 \times 10^{-16}$	
interaction	284.46	<2.2 × 10 <sup>-16</sup>	interaction	307.51	$< 2.2 \times 10^{-16}$	

both of the evolved traits. *Doa* is a very large gene, spanning 34.7 kb, and encodes at least 6 protein isoforms that are expressed in an age- and tissue-specific manner from alternative promoters and which have different, non-redundant functions [25,26]. SNPs located at different positions within the *Doa* gene might thus have different functions.

In total, we identified 16 biallelic SNPs (either intronic or in the coding region) having highly significant allele frequency differentiation (FDR < 0.0005) between the early-reproduction and late-reproduction (and hence increased lifespan) selection regimes in our E&R study [22] (figure 1b, electronic supplementary material, table S1). We chose four of these SNPs, based on their allele frequencies' differentiation between the selection regimes and their distribution across the length of the gene, for functional assays (figure 1, red arrows, and table 1; see below). As these SNPs were identified in an E&R study performed in a controlled laboratory setting, we first compared their allele frequencies to those in natural populations, using data from the DEST database [46] (see electronic supplementary material, File S6 for approach). These analyses showed that

all four SNPs are polymorphic in European populations and that the allele frequencies observed in the E&R study fall within the range of frequencies in natural populations (Doa-1: 0.16-0.57, Doa-2: 0.34-0.72, Doa-3: 0.65-0.98, Doa-4: 0.51-0.95, see electronic supplementary material, File S6 and table S2). Moreover, there was a significant latitudinal cline in allele frequency at Doa-4 ( $\chi^2 = 174.3$ ,  $P_{\text{corrected}} =$ 0.0069) and a significant longitudinal cline at Doa-3 ( $\chi^2$  = 222.9,  $P_{\text{corrected}} = 0.037$ ) across European populations (electronic supplementary material, table S4). This is notable because numerous life-history traits in D. melanogaster, including fecundity and lifespan, exhibit a clinal distribution, presumably due to spatially varying selection [3,42,43,47-50]. These population genetic observations thus lend further support to the idea that Doa potentially represents a target of selection on life-history traits.

#### (b) Doa has pleiotropic life-history effects

To functionally validate the role of *Doa* in longevity and fecundity at the level of the whole gene we knocked down

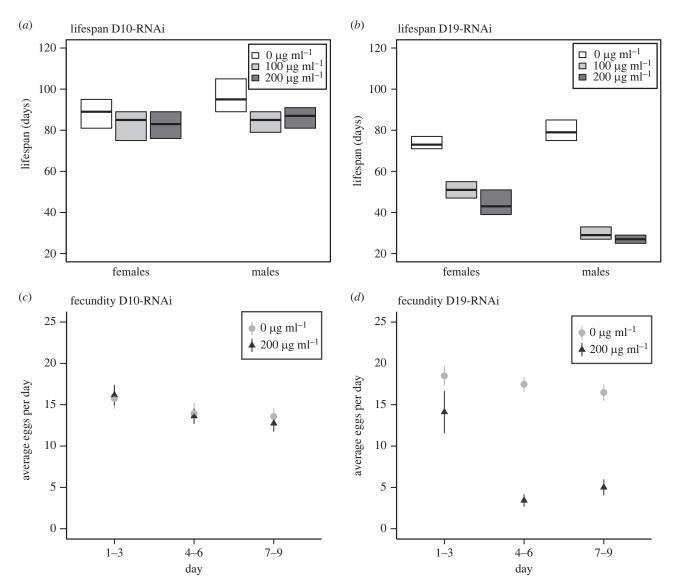


Figure 2. Knockdown of Doa using transgenic RNAi demonstrates significant effects on lifespan and fecundity. Two independent RNAi constructs (D19 and D10) that both target the catalytic domain of Doa were used in combination with the mifepristone-inducible GeneSwitch-GAL4 > UAS system. Concentrations refer to the concentrations of mifepristone (RU-486) used to induce RNAi. A significant reduction of adult lifespan was observed with both constructs ((a) construct D10, (b) construct D19), whereas a reduction in female fecundity was observed for construct D19 only ((c) construct D10, (d) construct D19; see table 2 for statistics).

all transcript variants by targeting the common, catalytic domain of Doa with ubiquitous (i.e. non-tissue-specific) transgenic RNAi. We employed two different RNAi constructs (i.e. two independent chromosomal insertions) to control for confounding effects of insertion position. To exclude potentially detrimental developmental carry-over effects of Doa knockdown on adult fitness, we reduced Doa expression levels in the adult stage only, by driving RNAi with the mifepristone-inducible GeneSwitch-GAL4 system [38]. For both Doa-RNAi constructs, we observed a significant reduction in lifespan in both sexes with increasing levels of mifepristone (figure 1a,b, table 2, electronic supplementary material, S2). Overall, the effect on lifespan was strongest for the D19 construct, which also achieved a stronger knockdown of Doa as determined by qRT-PCR as compared to construct D10. For both constructs, there was a significant interaction between sex and mifepristone concentration, which reflects the overall stronger effects of Doa RNAi on male than female lifespan. These findings agree with the observation of Huang et al. [31] that weak, constitutive knockdown of Doa (also using construct D10, VDRC #102520) affects lifespan in males, but

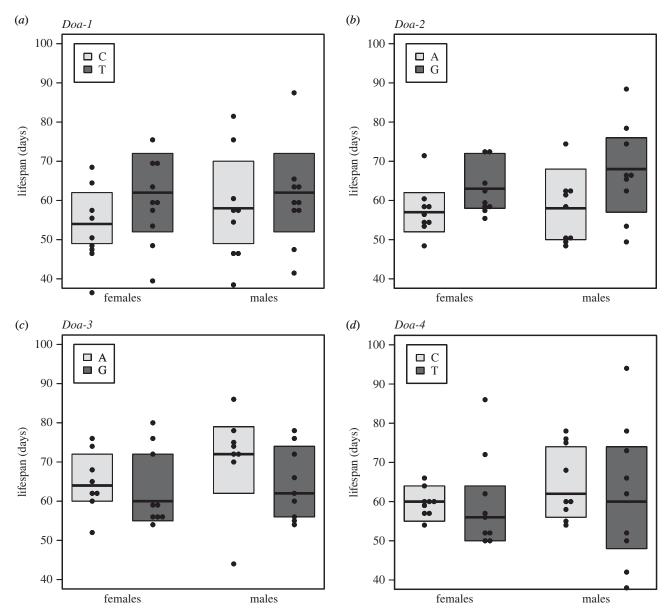
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not females; however, in their study, the direction of the effect depended on assay temperature. Sexual dimorphism in longevity is not uncommon among organisms, inlcuding fruit flies [51,52]. In terms of fecundity, we observed a strong, significant reduction in egg-laying rate for construct D19 but not for D10 (figure 2c,d, table 2).

These findings show that the Doa gene has pleiotropic effects on lifespan and reproduction [22] and demonstrate that modifying expression in the adult stage is sufficient to mediate these effects. Moreover, the magnitude and the direction of these effects depend on the strength of the knockdown (also see [31]). To obtain a better understanding of the role of Doa in evolving populations we next studied the effects of the four candidate SNPs identified in the E&R study on fecundity and longevity (see above and [22]).

# (c) *Doa* natural alleles have pleiotropic and nonpleiotropic effects

The four candidate SNPs that were functionally characterized were located both in coding and non-coding regions; Doa-4 is



**Figure 3.** Association between candidate SNPs at *Doa* and lifespan. F1 crosses of the genetically diverse *Drosophila* Genetic Reference Panel with different allelic states were assessed to test the association of the candidate SNPs with lifespan. The graphs show the average lifespans of the F1 crosses for each allelic state at the four candidate *Doa* SNP positions investigated. The allele that is predominantly found in long-lived, late-reproducing EE populations, as identified by Hoedjes *et al.* [22], is indicated by the darker shade. Significant differences in lifespan between the two allelic states were observed for *Doa-1*, *Doa-2* and *Doa-3* (see table 3 for statistics).

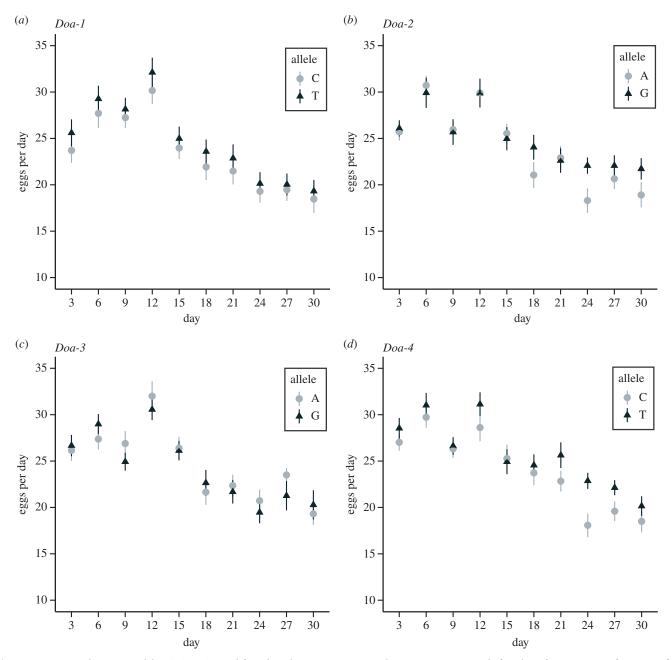
a synonymous SNP located in the catalytic domain that is shared by all isoforms; *Doa-1* and *Doa-2* are intronic SNPs and may have regulatory functions; and *Doa-3* is a missense SNP located in exon N8 (N-terminal variable region), which encodes part of the 227 kD protein isoform [26]. To study the effects of these SNPs on fitness components we used a SNP association approach based on Mendelian randomization using lines of the *Drosophila* Genetic Reference Panel (DGRP) (see above, §2) (figure 2).

For three of the *Doa* SNPs we found a significant correlation between allelic state and median lifespan (figure 3, table 3). For *Doa-1* and *Doa-2*, the correlation was in the predicted direction (i.e. increased median longevity of lines carrying the allele that was enriched in the long-lived E&R populations). Interestingly, there was also a significant correlation between lifespan and allelic state for *Doa-3*, but the direction was opposite to what we had hypothesized: although the 'G' variant was the major allele in the long-

lived experimental evolution lines [22], this allele was associated with lower median lifespan in our functional assays (figure 3, table 3).

In terms of effects on fecundity, we observed a significant correlation between allelic state and egg-laying rates for two of the *Doa* SNPs, *Doa-2* and *Doa-4*. Moreover, in both cases, there was also a significant interaction between allelic state and age, indicating that the effect on fecundity was age-specific (figure 4, table 3). The difference in egg-laying rate became visible starting from 18–24 days after eclosion, with a higher fecundity of the crosses carrying the alleles associated with selection for postponed reproduction in the E&R study [22].

To rule out potentially confounding effects of linked causal SNPs in the genetic background, we analysed LD (measured by pairwise  $r^2$ ) among Doa SNPs in the DGRP, which indicated very low levels of LD across the gene, as well as among the four candidate SNPs (see electronic supplementary material, File



**Figure 4.** Association between candidate SNPs at *Doa* and fecundity. Shown are average egg-laying rates over a period of 30 days after emergence of F1 crosses for each allelic state at the four *Doa* candidate SNP positions investigated. The allele that is pre-dominantly found in long-lived, late-reproducing EE populations, as identified by Hoedjes *et al.* [22], is indicated by the darker shade and triangles. Significant differences in lifespan between the two allelic states were observed for *Doa-2* and *Doa-4* (see table 3 for statistics).

S4). In addition, analyses of genetic differentiation (SNP-wise  $F_{\rm ST}$ ) among lines with alternative allelic states demonstrated that only the focal SNP was fixed ( $F_{\rm ST}$  = 1) for each of the four panels of lines (electronic supplementary material, File S5). None of the other SNPs, both within Doa or elsewhere in the genome, were fixed between two panels of lines, and the number of strongly differentiated SNPs ( $F_{\rm ST}$  > 0.5) was very low as well. These analyses strongly suggest that our findings are very unlikely to be confounded by LD or due to effects in the genetic background of the candidate SNPs tested.

#### 4. Conclusion

Our study provides strong support for a role of *Doa* in the evolution of lifespan and fecundity in the fruit fly, as expected

based on our previous E&R study [22]. Ubiquitous gene silencing of Doa using transgenic RNAi in adult flies reduced both lifespan and fecundity, indicating positive pleiotropy. In addition, each of the four candidate Doa SNPs tested had a significant effect on either lifespan and/or fecundity. The exact effects depended on the specific SNP, however, indicating that functional characterization of individual polymorphisms is essential for identifying the loci underlying adaptation. Three polymorphisms had effects on lifespan and/or fecundity that agree qualitatively well with predictions [22]. One of these SNPs, Doa-2, affected both lifespan and late-life fecundity, illustrating that even single nucleotide changes can have pleiotropic effects on complex traits (for other examples see [37,42,43]). However, one of the four SNPs, Doa-3, affected lifespan in the opposite direction than predicted. A possible explanation for this surprising result

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**Table 3.** Statistical tests of the effects of alternative alleles at the four *Doa* SNP positions on lifespan and fecundity (egg-laying rate). F1 crosses of the genetically diverse *Drosophila* Genetic Reference Panel with different allelic states were set up and tested for lifespan and fecundity in order to assess the association of the candidate *Doa* SNPs with lifespan. Asterisks indicate significant results: \*\*\*p < 0.001, \*\*0.001 < p < 0.01, \*0.01 < p < 0.05.

lifespan			fecundity			
	χ²	p		χ²	p	
Doa-1			Doa-1			
allele	43.27	$4.02 \times 10^{-10} ****$	allele	0.83	0.66	
sex	137.99	<2.2 × 10 <sup>-16</sup> ***	day	477.8	<2.2 × 10 <sup>-16</sup> ***	
allele $\times$ sex	41.42	$1.23 \times 10^{-10} ***$	allele $ imes$ day	0.24	0.6224	
Doa-2			Doa-2			
allele	7.62	0.022 *	allele	16.23	0.0003 ***	
sex	79.22	<2.2 × 10 <sup>-16</sup> ***	day	394.66	<2.2 × 10 <sup>-16</sup> ***	
allele $\times$ sex	3.35	0.067	allele $ imes$ day	15.84	$6.90 \times 10^{-5} ***$	
Doa-3			Doa-3			
allele	12.02	0.0025 **	allele	1.20	0.55	
sex	228.45	<2.2 × 10 <sup>-16</sup> ***	day	348.77	<2.2 × 10 <sup>-16</sup> ***	
allele $\times$ sex	11.46	0.00071 ***	allele $ imes$ day	1.13	0.28	
Doa-4			Doa-4			
allele	3.78	0.15	allele	9.42	0.009 *	
sex	141.33	<2.2 × 10 <sup>-16</sup> ***	day	486.4	<2.2 × 10 <sup>-16</sup> ***	
allele × sex	3.17	0.075	allele $\times$ day	7.68	0.0056 *	

might be that this SNP, and potentially other SNPs at *Doa*, are part of functionally and evolutionarily important haplotypes subject to linkage disequilibrium and/or epistasis. Similar analyses of other candidate loci, both within *Doa* and elsewhere in the genome, could resolve these questions and provide a more comprehensive overview of the polygenic regulation of these traits. Together, our results illustrate that it is important to go beyond traditional gene knockdown or knockout analyses and to perform functional tests of putatively adaptive candidate loci in order to understand the genetic basis of evolutionary change.

Data accessibility. The raw data for this paper are available from the Dryad Digital Repository: https://doi.org/10.5061/dryad.0gb5mkm4t [53].

Supplementary material is available online [54]. This manuscript is available as a pre-print in BioRxiv at https://doi.org/10.1101/2022.10.14.512079 [55].

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administration, resources, software, supervision, validation, visualization, writing—original draft, writing—review and editing; H.K.: investigation, writing—review and editing; L.K.: conceptualization, funding acquisition, project administration, resources, supervision, writing—review and editing; T.F.: conceptualization, resources, supervision, writing—review and editing.

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#### References

- Schlötterer C, Kofler R, Versace E, Tobler R, Franssen SU. 2015 Combining experimental evolution with next-generation sequencing: a powerful tool to study adaptation from standing genetic variation. Heredity 114, 431–440. (doi:10.1038/hdy.2014.86)
- Schlötterer C, Tobler R, Kofler R, Nolte V. 2014
   Sequencing pools of individuals—mining genome-wide polymorphism data without big
- funding. *Nat. Rev. Genet.* **15**, 749–763. (doi:10. 1038/nrq3803)
- Flatt T. 2020 Life-history evolution and the genetics of fitness components in *Drosophila melanogaster*. *Genetics* 214, 1–45. (doi:10.1534/genetics.119. 300160)
- Long A, Liti G, Luptak A, Tenaillon O. 2015
   Elucidating the molecular architecture of adaptation
- via evolve and resequence experiments. *Nat. Rev. Genet.* **16**, 567–582. (doi:10.1038/nrg3937)
- Kessner D, Novembre J. 2015 Power analysis of artificial selection experiments using efficient whole genome simulation of quantitative traits. *Genetics* 199, 991. (doi:10.1534/genetics.115.175075)
- Orozco-TerWengel P, Kapun M, Nolte V, Kofler R, Flatt T, Schlötterer C. 2012 Adaptation of *Drosophila*

- to a novel laboratory environment reveals temporally heterogeneous trajectories of selected alleles. *Mol. Ecol.* **21**, 4931–4941. (doi:10.1111/j. 1365-294X.2012.05673.x)
- Tobler R, Franssen SU, Kofler R, Orozco-Terwengel P, Nolte V, Hermisson J, Schlötterer C. 2014 Massive habitat-specific genomic response in *D. melanogaster* populations during experimental evolution in hot and cold environments. *Mol. Biol. Evol.* 31, 364–375. (doi:10.1093/molbev/mst205)
- Burke MK, Dunham JP, Shahrestani P, Thornton KR, Rose MR, Long AD. 2010 Genome-wide analysis of a long-term evolution experiment with *Drosophila*. Nature 467, 587. (doi:10.1038/nature09352)
- Turner TL, Stewart AD, Fields AT, Rice WR, Tarone AM. 2011 Population-based resequencing of experimentally evolved populations reveals the genetic basis of body size variation in *Drosophila* melanogaster. PLoS Genet. 7, e1001336. (doi:10. 1371/journal.pqen.1001336)
- Zhou D et al. 2011 Experimental selection of hypoxia-tolerant Drosophila melanogaster. Proc. Natl Acad. Sci. USA 108, 2349–2354. (doi:10.1073/pnas. 1010643108)
- Jha AR, Zhou D, Brown CD, Kreitman M, Haddad GG, White KP. 2016 Shared genetic signals of hypoxia adaptation in *Drosophila* and in highaltitude human populations. *Mol. Biol. Evol.* 33, 501–517. (doi:10.1093/molbev/msv248)
- Turner TL, Miller PM. 2012 Investigating natural variation in *Drosophila* courtship song by the evolve and resequence approach. *Genetics* 191, 633. (doi:10.1534/genetics.112.139337)
- 13. Remolina SC, Chang PL, Leips J, Nuzhdin SV, Hughes KA. 2012 Genomic basis of aging and life-history evolution in *Drosophila melanogaster*. *Evolution* **66**, 3390–3403. (doi:10.1111/j.1558-5646. 2012.01710.x)
- Carnes MU et al. 2015 The genomic basis of postponed senescence in *Drosophila melanogaster*. PLoS ONE 10, e0138569. (doi:10.1371/journal.pone. 0138569)
- 15. Fabian DK *et al.* 2018 Evolution of longevity improves immunity in *Drosophila. Evol. Lett.* **2**, 567–579. (doi:10.1002/evl3.89)
- Reed LK et al. 2014 Systems genomics of metabolic phenotypes in wild-type *Drosophila melanogaster*. Genetics 197, 781–793. (doi:10.1534/genetics.114. 163857)
- Martins NE, Faria VG, Nolte V, Schlötterer C, Teixeira L, Sucena E, Magalhaes S. 2014 Host adaptation to viruses relies on few genes with different cross-resistance properties. *Proc. Natl Acad.* Sci. USA 111, 5938–5943. (doi:10.1073/pnas. 1400378111)
- Jalvingh KM, Chang PL, Nuzhdin SV, Wertheim B. 2014 Genomic changes under rapid evolution: selection for parasitoid resistance. *Proc. R. Soc. B* 281, 20132303. (doi:10.1098/rspb.2013.2303)
- Jha AR, Miles CM, Lippert NR, Brown CD, White KP, Kreitman M. 2015 Whole genome resequencing of experimental populations reveals polygenic basis of egg size variation in *Drosophila melanogaster*. Mol.

- Biol. Evol. **32**, 2616–2632. (doi:10.1093/molbev/msv136)
- Kang L, Aggarwal DD, Rashkovetsky E, Korol AB, Michalak P. 2016 Rapid genomic changes in Drosophila melanogaster adapting to desiccation stress in an experimental evolution system. BMC Genom. 17, 233. (doi:10.1186/s12864-016-2556-y)
- Hardy CM, Burke MK, Everett LJ, Han MV, Lantz KM, Gibbs AG. 2018 Genome-wide analysis of starvationselected *Drosophila melanogaster*—a genetic model of obesity. *Mol. Biol. Evol.* 35, 50–65. (doi:10.1093/ molbev/msx254)
- Hoedjes KM, van den Heuvel J, Kapun M, Keller L, Flatt T, Zwaan BJ. 2019 Distinct genomic signals of lifespan and life history evolution in response to postponed reproduction and larval diet in *Drosophila. Evol. Lett.* 3, 598–609. (doi:10.1002/ evl3.143)
- Barrett RD, Hoekstra HE. 2011 Molecular spandrels: tests of adaptation at the genetic level. *Nat. Rev. Genet.* 12, 767–780. (doi:10.1038/nrq3015)
- Morris JZ, Navarro C, Lehmann R. 2003
   Identification and analysis of mutations in bob, Doa and eight new genes required for oocyte specification and development in Drosophila melanogaster. Genetics 164, 1435–1446. (doi:10. 1093/genetics/164.4.1435)
- Kpebe A, Rabinow L. 2008 Dissection of darkener of apricot kinase isoform functions in *Drosophila*. *Genetics* 179, 1973–1987. (doi:10.1534/genetics. 108.087858)
- Kpebe A, Rabinow L. 2008 Alternative promoter usage generates multiple evolutionarily conserved isoforms of *Drosophila* DOA kinase. *Genesis* 46, 132–143. (doi:10.1002/dvg.20374)
- Rabinow L, Samson M.-L. 2010 The role of the Drosophila LAMMER protein kinase DOA in somatic sex determination. J. Genet. 89, 271–277. (doi:10. 1007/s12041-010-0038-6)
- Fumey J, Wicker-Thomas C. 2017 Mutations at the Darkener of Apricot locus modulate pheromone production and sex behavior in Drosophila melanogaster. J. Insect. Physiol. 98, 182–187. (doi:10.1016/j.jinsphys.2017.01.005)
- Yun B, Farkas R, Lee K, Rabinow L. 1994
   The Doa locus encodes a member of a new protein kinase family and is essential for eye and embryonic development in Drosophila melanogaster. Genes Dev. 8, 1160–1173. (doi:10. 1101/gad.8.10.1160)
- James BP, Staatz WD, Wilkinson ST, Meuillet E, Powis G. 2009 Superoxide dismutase is regulated by LAMMER kinase in *Drosophila* and human cells. *Free Radical Biol. Med.* 46, 821–827. (doi:10.1016/j. freeradbiomed.2008.12.012)
- Huang W, Campbell T, Carbone MA, Jones WE, Unselt D, Anholt RR. H., Mackay TF. C. 2020 Context-dependent genetic architecture of *Drosophila* life span. *PLoS Biol.* 18, 3000645. (doi:10.1371/journal.pbio.3000645)
- Ivanov DK, Escott-Price V, Ziehm M, Magwire MM, Mackay TFC, Partridge L, Thomton JM. 2015

- Longevity GWAS using the *Drosophila* genetic reference panel. *J. Gerontol. a-Biol.* **70**, 1470–1478. (doi:10.1093/gerona/glv047)
- Pallares LF, Lea AJ, Han C, Filippova EV, Andolfatto P, Ayroles JF. 2020 Diet unmasks genetic variants that regulate lifespan in outbred *Drosophila*. bioRxiv. (doi:10.1101/2020.10.19.346312)
- May CM, van den Heuvel J, Doroszuk A, Hoedjes KM, Flatt T, Zwaan BJ. 2019 Adaptation to developmental diet influences the response to selection on age at reproduction in the fruit fly. *J. Evol. Biol.* 32, 425–437. (doi:10.1111/jeb.13425)
- 35. Stern DL. 2011 *Evolution, development, & the predictable genome*. Greenwood Village, CO: Roberts & Co. Publishers.
- Stern DL. 2000 Evolutionary developmental biology and the problem of variation. *Evolution* 54, 1079–1091. (doi:10.1111/j.0014-3820.2000. tb00544.x)
- Hoedjes KM, Kostic H, Flatt T, Keller L. 2021 A single nucleotide variant in the PPARgamma-homolog *Eip75B* affects fecundity in *Drosophila*. *bioRxiv*. (doi:10.1101/2021.12.07.471536)
- 38. Osterwalder T, Yoon KS, White BH, Keshishian H. 2001 A conditional tissue-specific transgene expression system using inducible GAL4. *Proc. Natl Acad. Sci. USA* **98**, 12 596–12 601. (doi:10.1073/pnas.221303298)
- Tricoire H, Battisti V, Trannoy S, Lasbleiz C, Pret A-M, Monnier V. 2009 The steroid hormone receptor EcR finely modulates *Drosophila* lifespan during adulthood in a sex-specific manner. *Mech. Ageing Dev.* 130, 547–552. (doi:10.1016/j.mad.2009.05. 004)
- Garschall K, Dellago H, Galikova M, Schosserer M, Flatt T, Grillari J. 2017 Ubiquitous overexpression of the DNA repair factor dPrp19 reduces DNA damage and extends *Drosophila* life span. *NPJ Aging Mech. Dis.* 3, 5. (doi:10.1038/s41514-017-0005-z)
- Smith GD, Ebrahim S. 2003 'Mendelian randomization': can genetic epidemiology contribute to understanding environmental determinants of disease? *Int. J. Epidemiol.* 32, 1–22. (doi:10.1093/ije/dyg070)
- Betancourt NJ, Rajpurohit S, Durmaz E, Fabian DK, Kapun M, Flatt T, Schmidt P. 2021 Allelic polymorphism at *foxo* contributes to local adaptation in *Drosophila melanogaster*. *Mol. Ecol.* 30, 2817–2830. (doi:10.1111/mec.15939)
- Durmaz E, Rajpurohit S, Betancourt N, Fabian DK, Kapun M, Schmidt P, Flatt T. 2019 A clinal polymorphism in the insulin signaling transcription factor foxo contributes to life-history adaptation in *Drosophila. Evolution* 73, 1774–1792. (doi:10.1111/ evo.13759)
- Mackay TFC et al. 2012 The Drosophila melanogaster genetic reference panel. Nature 482, 173–178. (doi:10.1038/nature10811)
- Weir BS, Cockerham CC. 1984 Estimating F-statistics for the analysis of population structure. Evolution 38, 1358–1370. (doi:10.1111/j.1558-5646.1984. tb05657.x)

- 46. Kapun M *et al.* 2021 *Drosophila* evolution over space and time (DEST): a new population genomics resource. *Mol. Biol. Evol.* **39**, 5782–5805. (doi:10. 1093/molbev/msab259)
- Durmaz E, Benson C, Kapun M, Schmidt P, Flatt T. 2018 An inversion supergene in *Drosophila* underpins latitudinal clines in survival traits. *J. Evol. Biol.* 31, 1354–1364. (doi:10.1111/jeb.13310)
- Fabian DK, Kapun M, Nolte V, Kofler R, Schmidt PS, Schlötterer C, Flatt T. 2012 Genome-wide patterns of latitudinal differentiation among populations of *Drosophila melanogaster* from North America. *Mol. Ecol.* 21, 4748–4769. (doi:10.1111/j.1365-294X. 2012.05731.x)

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- Kapun M et al. 2020 Genomic analysis of European Drosophila melanogaster populations reveals longitudinal structure, continent-wide selection, and previously unknown DNA viruses. Mol. Biol. Evol. 37, 2661–2678. (doi:10.1093/molbev/msaa120)
- Schmidt PS, Matzkin L, Ippolito M, Eanes WF. 2005 Geographic variation in diapause incidence, life-history traits, and climatic adaptation in *Drosophila melanogaster*. *Evolution* 59, 1721–1732. (doi:10.1554/05-115.1)
- 51. Lehtovaara A, Schielzeth H, Flis I, Friberg U. 2013 Heritability of life span is largely sex limited in *Drosophila*. *Am. Nat.* **182**, 653–665. (doi:10.1086/673296)
- Regan JC, Khericha M, Dobson AJ, Bolukbasi E, Rattanavirotkul N, Partridge L. 2016 Sex difference in pathology of the ageing gut mediates the greater

- response of female lifespan to dietary restriction. *Elife* **5**, e10956. (doi:10.7554/eLife.10956)
- Hoedjes KM, Kostic H, Keller L, Flatt T. 2022 Data from: Natural alleles at the *Doa* locus underpin evolutionary changes in *Drosophila* lifespan and fecundity. Dryad Digital Repository. (doi:10.5061/ dryad.0qb5mkm4t)
- 54. Hoedjes KM, Kostic H, Keller L, Flatt T. 2022 Natural alleles at the *Doa* locus underpin evolutionary changes in *Drosophila* lifespan and fecundity. Figshare. (doi:10.6084/m9.figshare.c.6261914)
- Hoedjes KM, Kostic H, Keller L, Flatt T. 2022 Natural alleles at the *Doa* locus underpin evolutionary changes in *Drosophila* lifespan and fecundity. *bioRxiv*. (doi:10.1101/2022.10.14.512079)