

Genome-wide enrichment analysis between endometriosis and obesity-related traits reveals novel susceptibility loci

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Endometriosis is a chronic inflammatory condition in women that results in pelvic pain and subfertility, and has been associated with decreased body mass index (BMI). Genetic variants contributing to the heritable component have started to emerge from genome-wide association studies (GWAS), although the majority remain unknown. Unexpectedly, we observed an intergenic locus on 7p15.2 that was genome-wide significantly associated with both endometriosis and fat distribution (waist-to-hip ratio adjusted for BMI; WHRadjBMI) in an independent meta-GWAS of European ancestry individuals. This led us to investigate the potential overlap in genetic variants underlying the aetiology of endometriosis, WHRadjBMI and BMI using GWAS data. Our analyses demonstrated significant enrichment of common variants between fat distribution and endometriosis ($P = 3.7 \times 10^{-3}$), which was stronger when we restricted the investigation to more severe (Stage B) cases ($P = 4.5 \times 10^{-4}$). However, no genetic enrichment was observed between endometriosis and BMI ($P = 0.79$). In addition to 7p15.2, we identify four more variants with statistically significant evidence of involvement in both endometriosis and WHRadjBMI (in/near *KIFAP3*, *CAB39L*, *WNT4*, *GRB14*); two of these, *KIFAP3* and *CAB39L*, are novel associations for both traits. *KIFAP3*, *WNT4* and 7p15.2 are associated with the *WNT* signalling pathway; formal pathway analysis confirmed a statistically significant ($P = 6.41 \times 10^{-4}$) overrepresentation of shared associations in developmental processes/*WNT* signalling between the two traits. Our results

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demonstrate an example of potential biological pleiotropy that was hitherto unknown, and represent an opportunity for functional follow-up of loci and further cross-phenotype comparisons to assess how fat distribution and endometriosis pathogenesis research fields can inform each other.

INTRODUCTION

Endometriosis is a common condition in premenopausal women characterized by chronic pelvic inflammation causing pain and subfertility (1), and has an estimated heritability of 51% (2). The International Endogene Consortium (IEC) performed the largest endometriosis GWAS to date in 3194 surgically confirmed cases (including 1364 moderate–severe—Stage B—cases) and 7060 controls of European ancestry, with replication in a further 2392 cases and 2271 controls (3). One genome-wide significant locus was observed in an intergenic region on chromosome 7p15.2 (rs12700667), primarily associated with Stage B disease ($P = 1.5 \times 10^{-9}$, OR = 1.38, 95% CI 1.24–1.53). A second locus near *WNT4* (rs7521902) was found after meta-analysis with published results from a Japanese GWAS of 1423 cases and 1318 controls (4); a genome-wide meta-analysis confirmed the two loci and found a further five (5).

Rs12700667 on 7p15.2 also marked 1 of 16 reported genome-wide significant loci associated with waist-to-hip ratio adjusted for BMI (WHRadjBMI) in an independent GWAS meta-analysis by the GIANT Consortium involving 77 167 individuals of European ancestry with replication in a further 113 636 individuals (rs1055144: discovery $P = 1.5 \times 10^{-8}$; meta-analysis $P = 1.0 \times 10^{-24}$; $r^2 = 0.5$ with rs12700667 in 1000G pilot CEU data) (6,7). This was surprising, as prospective epidemiological studies have suggested consistently that reduced BMI—a measure of overall adiposity—is associated with increased risk of endometriosis, but there is relatively limited evidence for an association with WHRadjBMI—a measure of fat distribution (8,9). We conducted a logistic regression analysis in the IEC dataset of rs1055144 on endometriosis disease status, conditioning on rs12700667, which demonstrated that the SNPs reflected the same association signal (unpublished data; conditional $P = 0.65$).

The epidemiological evidence of an association between endometriosis and BMI, together with the observed GWAS locus in common between endometriosis and WHRadjBMI, led us to conduct a systematic investigation of overlap in association signals between the IEC endometriosis GWAS and GIANT Consortium WHRadjBMI ($N = 77\ 167$) (6,7) and BMI ($N = 123\ 865$) (7,10) meta-GWAS datasets through genetic enrichment analyses.

RESULTS

Genetic enrichment analysis of endometriosis with overall adiposity and fat distribution

Using independent, imputed (1000 Genomes pilot reference panel) GWAS datasets of endometriosis (IEC; 3194 cases including 1364 Stage B cases, 7060 controls), BMI (GIANT; 123 865 individuals) and WHRadjBMI (GIANT: 77 167 individuals), we first considered loci genome-wide significantly

associated with endometriosis, BMI or WHRadjBMI in each of the individual GWAS. The two genome-wide significant endometriosis loci (intergenic 7p15.2 and *WNT4*) had significantly lower P -values of association than expected by chance in the WHRadjBMI GWAS (Table 1: rs12700667, $P = 4.4 \times 10^{-5}$ and rs7521902, $P = 1.3 \times 10^{-3}$; binomial $P = 1.0 \times 10^{-4}$), while 2 of the 16 genome-wide significant WHRadjBMI loci (intergenic 7p15.2 and *GRB14*) had $P < 0.01$ in the endometriosis GWAS (binomial $P = 0.011$). No enrichment between genome-wide significantly associated loci was observed for endometriosis versus BMI (Supplementary Material, Table S1: rs12700667, $P = 0.27$ and rs7521902, $P = 0.92$).

To investigate whether statistical enrichment extended beyond genome-wide significant loci, we investigated the most significant ($P < 1 \times 10^{-3}$) independent ($r^2 < 0.2$) endometriosis GWAS signals for enrichment of WHRadjBMI or BMI signals with $P < 0.05$ and vice versa (number of lookup SNPs per dataset: $n = 717$ to 748; see Supplementary Material, Methods). We observed statistically significant enrichment between variants associated with endometriosis (particularly Stage B) and WHRadjBMI (all endometriosis versus WHRadjBMI: $P = 3.7 \times 10^{-3}$; Stage B endometriosis versus WHRadjBMI: $P = 4.5 \times 10^{-4}$), but not between endometriosis and BMI (all endometriosis versus BMI: $P = 0.79$; Stage B endometriosis versus BMI: $P = 0.85$) (Fig. 1; Supplementary Material, Table S2). Results were similar when using female-limited WHRadjBMI ($N = 42\ 969$ women) and BMI ($N = 73\ 137$ women) GWAS summary statistics (7); to optimize power, in the remainder of the paper we therefore focus on sex-combined WHRadjBMI and BMI datasets (Supplementary Material, Fig. S1). Empirical testing of statistical enrichment through permutation (see Supplementary Material, Methods) provided near-identical results (Fig. 1; Supplementary Material, Fig. S1).

The choice of significance thresholds in the discovery and lookup datasets was based on a balance between applying a sufficiently stringent significance threshold in the discovery dataset that would minimize the proportion of false-positive association signals, while still having sufficient numbers of loci in each of the phenotypic association strata to investigate statistical enrichment, and allow the pursuit of meaningful biological pathway analyses subsequently. We considered the effect of different significance thresholds, for both discovery and lookup, which confirmed results showing enrichment of association signals between endometriosis and WHRadjBMI (Supplementary Material, Table S3), but no enrichment between endometriosis and BMI (Supplementary Material, Table S4).

To investigate potential genome-wide sharing of loci between endometriosis and WHRadjBMI or BMI, we performed polygenic prediction analyses (11) evaluating whether the aggregate effect of many variants of small effect in the WHRadjBMI and BMI GWAS could predict endometriosis status in the IEC GWAS (see Supplementary Material, Methods). There was no significant association between the WHRadjBMI- or BMI-

Table 1. Association results of published IEC genome-wide significant endometriosis loci (3) in the GIANT WHRadjBMI GWAS, and of WHRadjBMI loci (6,7) in endometriosis GWAS (lookup results are shown in bold)

GWAS	SNP (proxy; r^2)	Ch	Location (B36)	RAF (allele)	Status	Endometriosis all cases		Endometriosis Stage B only		Overall WHRadjBMI		Female-limited WHRadjBMI		Nearest gene
						P -value ^c	OR (95% CI)	P -value ^c	OR (95% CI)	P -value ^d	Effect (SE)	P -value ^c	Effect (SE)	
Endometriosis	rs12700667	7	25 868 164	0.74 (A)	G	5.1×10^{-7}	1.21 (1.12–1.31)	3.3×10^{-8}	1.36 (1.23–1.50)	4.4×10^{-5}	-0.023 (0.005)	3.3×10^{-8}	-0.023 (0.005)	Intergenic
Endometriosis	rs7521902	1	22 363 311	0.25 (A)	G	8.9×10^{-5}	1.16 (1.08–1.25)	7.5×10^{-5}	1.26 (1.14–1.39)	1.3×10^{-3}	-0.020 (0.006)	6.1×10^{-3}	-0.023 (0.009)	<i>WNT4</i>
WHRadjBMI	rs1055144 ^a	7	25 837 634	0.19 (T)	G	3.7×10^{-5}	0.84 (0.77–0.91)	3.1×10^{-4}	0.78 (0.70–0.88)	1.5×10^{-8}	0.034 (0.006)	2.3×10^{-6}	0.039 (0.008)	Intergenic
WHRadjBMI	rs10195252	2	165 221 337	0.41 (C)	G	9.8×10^{-3}	0.92 (0.85–0.98)	0.56	0.92 (0.84–1.00)	3.2×10^{-10}	-0.031 (0.005)	6.3×10^{-15}	-0.053 (0.007)	<i>GRB14</i>
Female WHRadjBMI	rs4684854	3	12 463 882	0.43 (C)	I (0.98)	0.07	1.06 (0.99–1.14)	0.14	1.07 (0.98–1.17)	1.0×10^{-4}	0.019 (0.005)	2.3×10^{-8}	0.039 (0.007)	<i>PPARG</i>
WHRadjBMI	rs718314	12	26 344 550	0.24 (G)	G	0.11	1.06 (0.99–1.15)	0.054	1.10 (0.99–1.22)	2.4×10^{-8}	0.031 (0.005)	8.2×10^{-10}	0.047 (0.008)	<i>ITPR2-SSPN</i>
WHRadjBMI	rs6861681	5	173 362 458	0.32 (A)	I (0.96)	0.15	0.95 (0.86–1.04)	0.11	0.93 (0.85–1.00)	1.4×10^{-6}	0.026 (0.005)	2.1×10^{-4}	0.027 (0.007)	<i>CPEB4</i>
WHRadjBMI	rs6795735	3	64 680 405	0.41 (T)	G	0.21	1.04 (0.98–1.12)	0.32	1.04 (0.96–1.14)	2.5×10^{-7}	-0.025 (0.005)	7.8×10^{-7}	-0.033 (0.007)	<i>ADAMTS9</i>
WHRadjBMI	rs2820446	1	21 974 881	0.71 (C)	I (0.99)	0.31	1.04 (0.97–1.12)	0.22	1.06 (0.97–1.17)	5.1×10^{-12}	0.037 (0.005)	8.5×10^{-18}	0.064 (0.007)	<i>LYPLAL1</i>
WHRadjBMI	rs498778 (rs4846567, $r^2 = 1$) ^b	3	52 453 893	0.93 (T)	I (0.95)	0.32	1.08 (0.93–1.24)	0.25	1.06 (0.89–1.27)	4.6×10^{-5}	0.055 (0.010)	1.1×10^{-3}	0.063 (0.019)	<i>NISCH-STAB1</i>
WHRadjBMI	rs1294421 (rs6784615, $r^2 = 1$) ^b	6	6 743 149	0.39 (T)	I (0.96)	0.37	1.03 (0.94–1.10)	0.28	1.03 (0.94–1.13)	6.3×10^{-9}	-0.029 (0.005)	3.4×10^{-8}	-0.038 (0.007)	<i>LY86</i>
WHRadjBMI	rs9491696	6	127 452 639	0.51 (C)	I (0.99)	0.43	0.97 (0.91–1.03)	0.64	0.98 (0.90–1.06)	2.1×10^{-14}	-0.037 (0.005)	3.4×10^{-8}	-0.038 (0.007)	<i>RSPO3</i>
WHRadjBMI	rs1443512	12	52 628 951	0.22 (A)	G	0.62	1.02 (0.94–1.10)	0.63	0.97 (0.88–1.08)	3.3×10^{-8}	0.031 (0.005)	1.4×10^{-9}	0.048 (0.008)	<i>HOXC13</i>
WHRadjBMI	rs984222	1	119 305 366	0.39 (C)	I (0.99)	0.69	0.99 (0.93–1.05)	0.31	0.95 (0.87–1.04)	3.8×10^{-14}	-0.037 (0.005)	1.2×10^{-7}	-0.036 (0.007)	<i>TBX15-WARS2</i>
WHRadjBMI	rs4823006	22	29 451 671	0.57 (A)	I (0.97)	0.72	1.01 (0.95–1.08)	0.82	1.01 (0.92–1.11)	4.7×10^{-10}	0.030 (0.005)	6.9×10^{-8}	0.037 (0.007)	<i>ZNF3</i>
Female WHRadjBMI	rs10478424	5	118 816 619	0.79 (A)	I (0.97)	0.80	1.01 (0.93–1.10)	0.56	1.03 (0.93–1.15)	1.6×10^{-4}	0.023 (0.006)	1.0×10^{-5}	0.037 (0.009)	<i>HSD17B4</i>
WHRadjBMI	rs1011731	1	170 613 171	0.44 (G)	G	0.81	0.99 (0.93–1.05)	0.77	1.01 (0.93–1.11)	1.7×10^{-10}	0.031 (0.005)	2.1×10^{-5}	0.028 (0.007)	<i>DNM3-PIGC</i>
WHRadjBMI	rs6905288	6	43 866 851	0.56 (A)	I (0.80)	0.66	0.98 (0.91–1.05)	0.66	0.99 (0.90–1.08)	4.2×10^{-10}	0.033 (0.005)	7.7×10^{-13}	0.052 (0.007)	<i>VEGFA</i>

^aLogistic regression analysis in the IEC GWAS shows that rs1055144 marks the same locus as rs12700667 (conditional $P = 0.65$; $r^2 = 0.8$).

^bSNP was not genotyped in the endometriosis GWAS dataset; result shown is of proxy SNP.

^cResults are based on an updated GWAS performed using genotype data imputed up to 1000 Genomes pilot reference panel (B36, June 2010).

^dResults are from the GIANT WHRadjBMI discovery GWAS dataset ($N = 77\ 167$); 3 of the 14 WHRadjBMI loci have $P > 5.0 \times 10^{-8}$, however, they reached genome-wide significance combined with replication analyses in up to a further 113 636 individuals (6).

^eResults from the GIANT WHRadjBMI discovery female-limited GWAS dataset ($N = 42\ 969$); one of the two female-limited WHRadjBMI loci have $P > 5.0 \times 10^{-8}$, however, they reached genome-wide significance combined with replication analyses in up to a further 71 295 individuals (7).

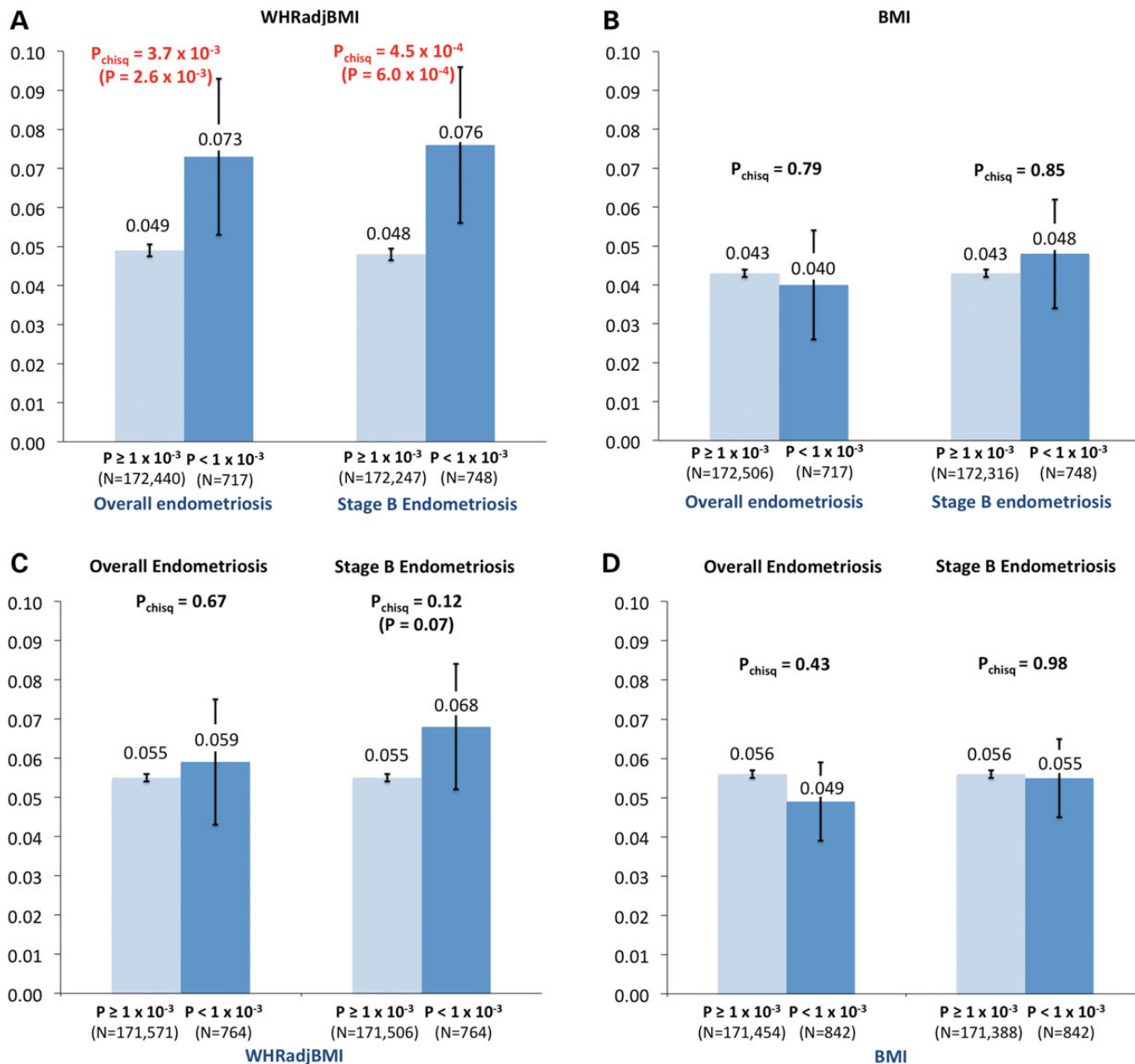


Figure 1. Genetic enrichment analyses between endometriosis, BMI and WHRadjBMI GWAS datasets, using independent ($r^2 < 0.2$) SNPs. The panels show (i) The proportion of SNPs nominally associated ($P < 0.05$) with WHRadjBMI (A) or BMI (B) by significance of overall and Stage B endometriosis association ($P < 1.0 \times 10^{-3}$ versus $P \geq 1 \times 10^{-3}$); (ii) The proportion of SNPs nominally associated ($P < 0.05$) with overall and Stage B endometriosis by significance of WHRadjBMI (C) and BMI (D) association ($P < 1.0 \times 10^{-3}$ versus $P \geq 1 \times 10^{-3}$). P -values of χ^2 tests assessing statistical difference between proportions are shown above each set of bars, and 95% confidence intervals of the proportions are given on each bar. For differences with $P_{\text{chisq}} < 0.2$, empirical P -values are given in brackets (see Supplementary Material, Methods).

derived profile scores (overall or female limited) and all/Stage B endometriosis (Supplementary Material, Tables S5–S8), suggesting no evidence for a directionally consistent *en masse*, genome-wide, shared common genetic component.

We next investigated the variants with most significant evidence for association with both endometriosis ($P < 1 \times 10^{-3}$) and WHRadjBMI ($P < 0.05$) for predominance in direction of phenotypic effects (Supplementary Material, Tables S9 and S10 and Fig. S2). No statistically significant directional

consistency was observed for these variants ($P > 0.47$), nor for the 17 loci (Table 1) that were genome-wide significantly associated with either trait (Fig. 2, $P > 0.44$). Intergenic 7p15.2 and *WNT4* showed discordant directions of effect, while the effect of *GRB14* was concordant (Fig. 2). This could suggest the presence of multiple biological pathways through which the variants influence the two phenotypes. We next set out to investigate the common biology suggested by genetic variants associated with both endometriosis and WHRadjBMI.

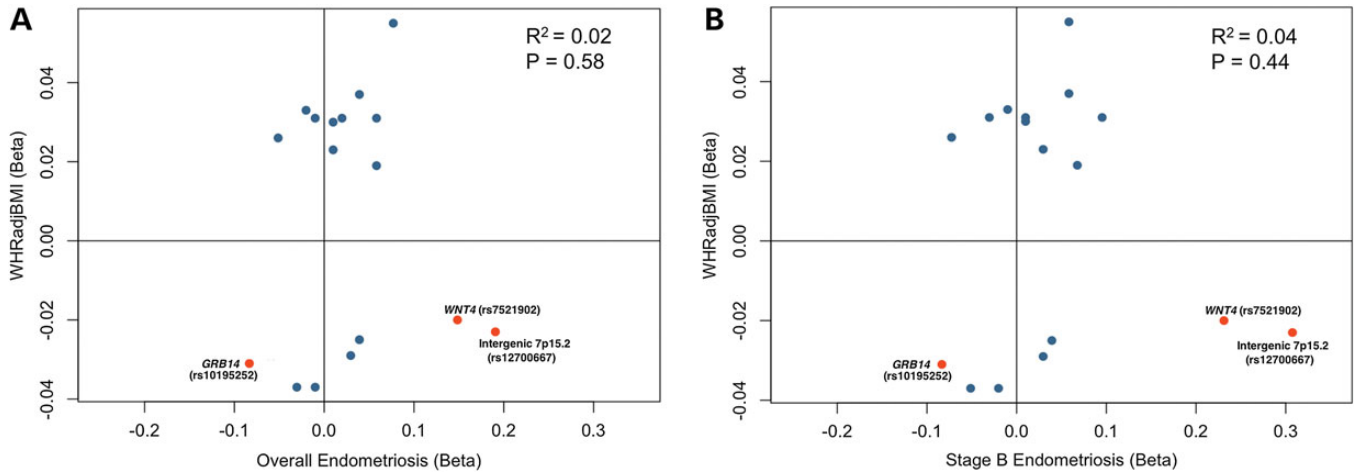


Figure 2. Directions of effect of 17 independent SNPs genome-wide significantly associated with all (A) or Stage B (B) endometriosis, or WHRadjBMI. Intergenic 7p15.2, *WNT4*, and *GRB14* are shown in red. Linear regression R^2 and P -values used to test for significant directionality of effects (35) are shown.

Biology of the loci shared between endometriosis and fat distribution

Our analysis showing significant enrichment between SNPs associated with all or Stage B endometriosis ($P < 1 \times 10^{-3}$) and WHRadjBMI ($P < 0.05$) shown in Figure 1 involved 1284 independent ($r^2 > 0.2$) loci. We explored the biological function of the loci most strongly associated with WHRadjBMI, at nominal $P < 0.005$ ($n = 16$, Table 2; see Supplementary Material, Tables S11 and S12 for all variants associated at $P < 0.05$). Two novel loci, rs560584 near *KIFAP3* (all endometriosis) and rs11619804 in *CAB39L* (Stage B endometriosis), were significantly associated with WHRadjBMI after Bonferroni correction allowing for 1284 independent tests ($P < 3.89 \times 10^{-5}$).

The endometriosis risk allele T of rs560584 (OR = 1.14 (1.07–1.22), $P = 1.42 \times 10^{-4}$) was associated with lower WHRadjBMI ($\beta = -0.021$, $P = 1.47 \times 10^{-5}$), and located in an intergenic region 46 kb downstream of *KIFAP3* (*Kinesin-associated protein 3*). Together with *KIF3A* and *KIF3B*, *KIFAP3* forms a kinesin motor complex, KIF3, that mediates cellular transport of N-cadherin and β -catenins (12), which are involved in cell adhesion, the *Wnt* canonical pathway and cell cycle progression (13). The *Wnt*/ β -catenin signalling pathway acts as a molecular switch for adipogenesis (14) and has multiple suggested roles in endometriosis through sex hormone homeostasis regulation (15), its role in development of female reproductive organs (16), molecular mechanisms of infertility (17) and mediation of fibrogenesis (18).

The Stage B endometriosis risk allele C of rs11619804 (OR = 1.17 (1.07–1.28); $P = 4.88 \times 10^{-4}$), located in *CAB39L* (Calcium-Binding Protein 39-Like), was associated with increased WHRadjBMI ($\beta = 0.022$, $P = 1.06 \times 10^{-5}$; Table 2). The function of this gene is not well characterized but the encoded protein interacts with a serine threonine kinase (*STK11*) that functions as a tumour suppressor (19).

Rs12700667 in the intergenic region 7p15.2 remained among the strongest associated shared signals, with the endometriosis risk allele A associated with reduced WHRadjBMI ($\beta = -0.023$, $P = 4.4 \times 10^{-5}$). The association maps to an intergenic high LD region of 48 kb ($r^2 > 0.8$) of unknown functionality.

Further interesting nearby loci include the miRNA *hsa-mir-148a*, with a purported role in *Wnt*/ β -catenin signalling (14); *NFE2L3* (nuclear factor (erythroid-derived 2)-like 3), a transcription factor suggested to be involved in cell differentiation, inflammation and carcinogenesis (20). The *WNT* signalling pathway was further highlighted by the nominal association of two independent ($r^2 = 0.06$) endometriosis risk variants near *WNT4* (wingless-type MMTV integration site family), rs3820282 (genic) and rs2807357 (22.4 kb downstream), with reduced WHRadjBMI ($\beta = -0.019$, $P = 5.0 \times 10^{-3}$; $\beta = -0.015$, $P = 3.7 \times 10^{-3}$; Table 2). Of note is that all shared variants implicated in *WNT* signalling (in/near intergenic 7p15.2, *WNT4*, *KIFAP3*) showed consistent—discordant—phenotypic directions of effect.

Risk variant rs10195252, 34.6 kb downstream of *GRB14* (growth factor receptor-bound protein 14) was the third locus with significant evidence for association with both overall (not Stage B) endometriosis and WHRadjBMI (Table 1). *GRB14* has an inhibitory effect on insulin receptor signalling (21), may have a role in signalling pathways that regulate growth and metabolism and has been shown to interact with fibroblast growth factor receptors (22). This shared variant is also in high LD ($r^2 = 0.93$ and $= 0.87$, respectively) with a type 2 diabetes risk variant rs13389219 (23) and fasting insulin risk variant rs6717858 (24).

Other loci of interest include rs2921188 in *PPARG* and rs6556301 near *FGFR4* (Table 2). The endometriosis risk allele A of rs2921188 (OR = 1.13, 95% CI: 1.05–1.21), $P = 5.9 \times 10^{-4}$) in *PPARG* (peroxisome proliferator-activated receptor gamma) is associated with increased WHRadjBMI ($\beta = 0.017$; $P = 1.1 \times 10^{-3}$). *PPARG* is a nuclear hormone receptor that regulates fatty acid storage and glucose metabolism. Synthetic ligands, such as insulin sensitizing drugs, target *PPARG* in treatment of diabetes to lower serum glucose levels (25) and are also documented to have anti-inflammatory, anti-angiogenic and anti-proliferative effects on endometrium, with baboon models suggesting a role in targeting endometriotic disease (26). Stage B endometriosis risk allele G of rs6556301 near *FGFR4* (*fibroblast growth factor receptor*, OR = 1.17 [1.07–1.28], $P = 7.4 \times 10^{-4}$) is associated with reduced WHRadjBMI ($\beta = -0.021$, $P = 1.9 \times 10^{-4}$). *FGFR4* interacts

Table 2. Results of the top all/Stage B endometriosis loci ($P < 1 \times 10^{-3}$) associated with WHRadjBMI at $P < 0.005$

SNP	Chr	Position (B36)	RAF (allele)	Endometriosis		Overall WHRadjBMI			Female-limited WHRadjBMI			Nearest loci (distance)
				P-value	OR (95% CI)	P-value	Effect	SE	P-value	Effect	SE	
All cases												
rs560584	1	168 357 136	0.41 (T)	1.4×10^{-4}	1.14 (1.07–1.22)	1.4×10^{-5}	-0.021	0.005	1.1×10^{-3}	-0.022	0.677	<i>KIFAP3</i> (46 632)
rs12700667	7	25 868 164	0.74 (A)	5.1×10^{-7}	1.22 (1.13–1.32)	4.4×10^{-5}	-0.023	0.005	3.4×10^{-4}	-0.028	0.284	<i>NFE2L3</i> (2 90 221)
rs2921188	3	12 387 115	0.64 (A)	5.9×10^{-4}	1.13 (1.05–1.21)	1.1×10^{-3}	0.017	0.005	1.8×10^{-4}	0.026	0.054	<i>PPARG</i> (0)
rs1250248	2	215 995 338	0.27 (A)	1.6×10^{-5}	1.17 (1.09–1.26)	1.0×10^{-3}	0.018	0.005	9.9×10^{-4}	0.025	0.242	<i>FNI</i> (0)
rs2630787	3	21 847 339	0.52 (C)	9.2×10^{-4}	1.12 (1.05–1.19)	1.9×10^{-3}	-0.015	0.004	0.38	-0.006	0.030	<i>ZNF659</i> (79 518)
rs1430788	2	67 721 916	0.31 (C)	9.3×10^{-5}	1.15 (1.07–1.23)	2.7×10^{-3}	0.016	0.005	3.1×10^{-3}	0.022	0.330	<i>ETAA1</i> (230 878)
rs906721	3	184 687 691	0.41 (A)	6.1×10^{-5}	1.16 (1.08–1.24)	4.2×10^{-3}	0.015	0.005	1.7×10^{-3}	0.023	0.140	<i>KLHL6</i> (322)
rs1868894	4	187 606 728	0.80 (C)	2.3×10^{-4}	1.16 (1.07–1.26)	4.9×10^{-3}	-0.018	0.006	0.13	-0.013	0.524	<i>MTNRIA</i> (85 075)
rs3820282	1	22 340 802	0.16 (T)	3.3×10^{-7}	1.26 (1.15–1.37)	5.0×10^{-3}	-0.019	0.007	0.09	-0.016	0.749	<i>WNT4</i> (0)
Stage B cases												
rs11619804	13	49 888 131	0.53 (C)	4.8×10^{-4}	1.17 (1.07–1.28)	1.1×10^{-5}	0.022	0.005	2.2×10^{-2}	0.016	0.022	<i>CAB39L</i> (0)
rs12700667	7	25 868 164	0.74 (A)	3.3×10^{-9}	1.36 (1.23–1.50)	4.4×10^{-5}	-0.023	0.005	3.4×10^{-4}	-0.028	0.284	<i>NFE2L3</i> (290 221)
rs2782659	6	45 794 768	0.33 (G)	4.2×10^{-4}	1.18 (1.08–1.30)	9.2×10^{-5}	0.020	0.005	1.7×10^{-4}	0.027	0.108	<i>RUNX2</i> (167 970)
rs6556301	5	176 460 183	0.63 (G)	7.4×10^{-4}	1.17 (1.07–1.28)	1.9×10^{-4}	-0.021	0.005	7.8×10^{-3}	-0.021	0.845	<i>FGFR4</i> (2450)
rs1250248	2	215 995 338	0.27 (A)	2.9×10^{-8}	1.32 (1.19–1.45)	1.2×10^{-3}	0.018	0.005	9.9×10^{-4}	0.025	0.242	<i>FNI</i> (0)
rs4131816	1	161 662 648	0.85 (T)	5.4×10^{-4}	1.24 (1.10–1.41)	1.5×10^{-3}	0.022	0.007	0.25	0.011	0.072	<i>NUF2</i> (70 470)
rs9912335	17	77 552 948	0.69 (T)	3.1×10^{-4}	1.19 (1.08–1.31)	3.5×10^{-3}	-0.021	0.007	0.10	-0.016	0.454	<i>ASPSCR1</i> (0)
rs10878362	12	64 703 760	0.69 (C)	4.9×10^{-4}	1.19 (1.08–1.31)	3.6×10^{-3}	0.015	0.005	3.1×10^{-3}	0.022	0.204	<i>HMG2</i> (57 421)
rs2807357	1	22 364 571	0.64 (A)	9.7×10^{-4}	1.16 (1.06–1.27)	3.7×10^{-3}	-0.015	0.005	1.0×10^{-3}	-0.024	0.081	<i>WNT4</i> (22 373)
rs906721	3	184 687 691	0.41 (A)	1.4×10^{-4}	1.20 (1.09–1.32)	4.2×10^{-3}	0.015	0.005	1.7×10^{-3}	0.023	0.140	<i>KLHL6</i> (322)
rs12267660	10	4 419 530	0.85 (G)	7.9×10^{-4}	1.24 (1.09–1.40)	4.6×10^{-3}	0.02	0.007	8.0×10^{-3}	0.030	0.133	<i>CR749391</i> (191 913)
rs11685481	2	67 590 253	0.15 (C)	8.4×10^{-4}	1.23 (1.09–1.38)	4.8×10^{-3}	0.018	0.006	1.1×10^{-2}	0.022	0.451	<i>ETAA1</i> (99 215)

with fibroblast growth factors, which have roles in angiogenesis, wound healing and cell migration (27).

Expression quantitative trait loci analysis of the shared endometriosis and fat distribution loci

We investigated the potential impact of the described 16 genes (Table 2) shared between endometriosis and WHRadjBMI on transcriptional function using three public expression data resources: (i) the Mammalian Gene Expression Uterus database (MGEx-Udb) (28) containing published information on transcriptional activity of specific genes in human endometrial tissue from individuals with and without endometriosis; (ii) the MuTHER study which collected expression and eQTL data from 776 abdominal fat tissues (29); and (iii) the MOLOBB dataset of differential expression levels between abdominal and gluteal fat from 49 individuals (30). Based on the limited available evidence in the MGEx-Udb database, two genes are transcribed in endometrial tissue of women with endometriosis but dormant in those without endometriosis: *PPARG* and *FGFR4* (Supplementary Material, Table S13). Of the 16 genes, 15 had probes present within 1 Mb either side of the SNP in the MuTHER database; however, none showed significant association with nearby transcripts in abdominal fat tissue (Supplementary Material, Table S14). The MOLOBB study data showed *cis*-eQTL evidence for differential expression of two genes; *KIFAP3* (rs560584; fold change = 0.14, adjusted $P = 0.04$) (Supplementary Material, Table S15). Additional transcriptional evidence relevant to the intergenic 7p15.2 locus includes the presence of an expression QTL associated with a transcript of unknown function, *AA553656*, in subcutaneous abdominal fat tissue (6), and the differential expression of nearby *hsa-miR-148a* between gluteal and abdominal fat tissue samples (31).

Pathway analysis

To identify potential common biological pathways involved in the aetiology of endometriosis and the variability of fat distribution, we conducted pathway analyses using genes with evidence for enrichment between the traits using (i) the PANTHER database (32) and (ii) GRAIL (33). For the PANTHER analysis, we selected the 91 and 108 genes located in a 1 Mb interval surrounding each independent SNP associated with all endometriosis ($P < 1.0 \times 10^{-3}$) and WHRadjBMI ($P < 0.05$), and Stage B endometriosis ($P < 1.0 \times 10^{-3}$) and WHRadjBMI ($P < 0.05$), respectively (see Supplementary Material, Methods). This excluded intergenic loci without a gene within 1 Mb, such as our top shared locus at 7p15.2. We tested whether the two sets of genes showed significant overrepresentation of a particular pathway, for each of 176 curated pathways and 241 biological processes. The top enriched pathways were ‘developmental processes’ (all endometriosis: $P = 1.2 \times 10^{-3}$; Stage B: $P = 1.25 \times 10^{-4}$), ‘WNT signalling’ (all endometriosis: $P = 1.07 \times 10^{-4}$), ‘gonadotropin-releasing hormone receptor’ (all endometriosis: $P = 1.48 \times 10^{-3}$), ‘cadherin signalling’ (Stage B: $P = 6.42 \times 10^{-4}$), ‘FGF signalling’ (Stage B: $P = 2.96 \times 10^{-3}$) and ‘TGF-beta signalling’ (Stage B: $P = 1.48 \times 10^{-3}$) pathways (Supplementary Material, Tables S16 and S17). Bonferroni correction for the number of pathways

tested (see Supplementary Material, Methods) rendered ‘WNT signalling’, ‘developmental processes’, ‘cellular processes’ and ‘cell communication’ significantly enriched; however, this adjustment is conservative, as exemplified by ‘cadherin signalling’ genes being a subset of those in the ‘WNT signalling’ pathway. Sensitivity analyses exploring the effect of different endometriosis association thresholds on pathway analyses showed very consistent results for threshold $P < 1.0 \times 10^{-2}$, with the same top three enriched pathways—WNT signalling, Cadherin signalling and Gonadotropin-releasing hormone receptor pathway. No meaningful pathway analyses could be conducted on the limited number of genes passing association threshold $P < 1 \times 10^{-4}$ (Supplementary Material, Table S18).

We used GRAIL (33) to search for connectivity between the 91 and 108 genes all/Stage B endometriosis and WHRadjBMI-associated genes and specific keywords from the published literature that describe potential functional connections. We identified 17 genes with nominal significance ($P < 0.05$) for potential functional connectivity for ‘all’ endometriosis and WHRadjBMI and six genes for Stage B endometriosis and WHRadjBMI (Supplementary Material, Fig. S3 and Tables S19 and S20). The keywords associated with these connections included ‘cadherin’, ‘differentiation’, ‘development’ and ‘insulin’ for ‘all’ endo, and ‘development’ and ‘embryos’ for Stage B endometriosis, marking again developmental processes and cadherin signalling as biological pathways shared in the origins of endometriosis and fat distribution.

DISCUSSION

In this study, we have investigated the overlap in genetic association signals from the largest GWA studies to date of endometriosis, overall adiposity (BMI) and fat distribution (WHRadjBMI). Our results demonstrated that there is a shared genetic basis between endometriosis and fat distribution that extends over and above the single genome-wide significant locus that has been reported in GWAS of the separate traits. Our analyses highlight novel loci in/near *KIFAP3* and *CAB39L*, which together with intergenic 7p15.2, *WNT4* and *GRB14*, showed significant evidence of trait association sharing. The strength of evidence of enrichment was similar for overall versus female-limited WHRadjBMI loci, which may be unexpected, given that endometriosis is a female condition. However, the lack of a stronger enrichment between female-specific WHRadjBMI GWAS results and endometriosis, compared with all WHRadjBMI results should be considered against the effects of a reduced sample size used for female-specific WHRadjBMI analyses on power of association detection.

The enrichment of associated variants was generally stronger when the endometriosis cases were restricted to moderate–severe (Stage B) disease, despite the smaller sample size. Indeed, the association of the top intergenic GWAS locus on 7p15.2, also genome-wide significantly associated with WHRadjBMI, is limited to Stage B endometriosis. Stage B—or ASRM Stages III/IV disease (34)—is typically characterized by ovarian (endometrioma) or deep infiltrating (rectovaginal) lesions, which were shown to have a substantially greater underlying genetic contribution than milder, peritoneal disease (ASRM Stage I/II) (3). The particular enrichment between

WHRadjBMI and Stages III/IV endometriosis is intriguing, and another reason for further functional work to concentrate on this endometriosis sub-type. There are, however, specific loci that show enrichment of association with WHRadjBMI and overall endometriosis, the analysis of which therefore remains of interest. An example is *GRB14*, which did not show significant association with Stage B disease, displayed a concordant direction of effect between endometriosis and WHRadjBMI, and the biological function of which also seems to suggest an entirely different contribution to the origins of both phenotypes than the 7p15.2 and *WNT4* loci.

The limited available eQTL data showed significant evidence for differential expression of *KIFAP3* between different fat depots. The variants with most evidence for enrichment between the traits, in/near intergenic 7p15.2, *KIFAP3* and *WNT4*, were all implicated in *WNT* signalling and had consistent—discordant—directions of effect, with endometriosis risk alleles associated with a decreased WHRadjBMI. Indeed, biological pathway analyses showed significant evidence for the involvement of developmental processes and *WNT* signalling in endometriosis aetiology and regulation of fat distribution, a potential pleiotropic connection that has not been reported to date.

The relatively limited epidemiological evidence of phenotypic correlation between endometriosis and WHRadjBMI (8,9) is consistent with the absence of strong directional consistency of phenotypic effects of genetic variants underlying both traits at a genome-wide level. Most studies of genetic pleiotropy between traits to date have focused on genome-wide directional consistency between epidemiologically or clinically (postulated) correlated traits, such as different metabolic traits (6,35) or psychiatric conditions (36). However, genome-wide consistency in directionality of phenotypic effects would most likely apply to traits that share a large proportion of causality, and that epidemiologically lie on the same causal pathway(s) and are thus more likely to be examples of mediated (genetic variants influencing one phenotype indirectly through association with a second phenotype) rather than biological (genetic variants exerting a direct biological influence on more than one phenotype) pleiotropy (37). Thus, our results of genetic enrichment between endometriosis and WHRadjBMI demonstrate an example of the biological complexity of aetiological associations between complex traits, and suggest that the underlying shared loci are potentially biologically pleiotropic, given the absence of phenotypic correlation between endometriosis and WHRadjBMI and absence of *en masse* directional consistency of shared genetic variants on the phenotypes (37,38). It also demonstrates more generally how potential perturbation of a causal pathway through, for example, drug treatment targeting one trait could have unexpected effects on another, even when there is no clear evidence that these traits are associated clinically or epidemiologically—a problem often encountered in drug development. Systematic exploration of biological pleiotropy of genetic variants marking potential drug targets may help in highlighting the potential of such unwanted or unexpected effects.

While the observed genetic enrichment between endometriosis and WHRadjBMI presents new avenues for exploring common biology, the total absence of any genetic enrichment between endometriosis and BMI (within the limits of power presented by these large datasets) is intriguing given the consistent, prospective, observational epidemiological evidence of

phenotypic association between reduced BMI and endometriosis risk (8). Our analyses represent an adaptation of Mendelian randomization analyses (39,40), in which genetic variants robustly associated with BMI in the largest GWAS analyses to date (10) are investigated for association with endometriosis. The total lack of genetic enrichment suggests that reduced BMI is not causally related to endometriosis risk. Rather, it suggests that the observed phenotypic association (8) is either driven by shared environmental factors, or is due to confounding factors related to BMI affecting, for example, diagnostic opportunity for endometriosis.

These novel findings present an entirely new opportunity for functional targeted follow-up of pleiotropic loci between endometriosis and WHRadjBMI in relevant disease tissues such as endometrium and fat tissue, cellular systems, animal models and further cross-trait comparisons, to uncover their biological functions and to assess how studies in the fat distribution research field can inform research into endometriosis pathogenesis, biomarker identification and drug target discovery and validation.

MATERIALS AND METHODS

Genome-wide association studies

IEC endometriosis GWAS

This GWAS included 3194 surgically confirmed endometriosis cases and 7060 controls from Australia and the UK. Disease severity of the endometriosis cases was assessed retrospectively from surgical records using the rAFS classification system and grouped into two phenotypes: Stage A (Stage I or II disease or some ovarian disease with a few adhesions; $n = 1686$) or Stage B (Stage III or IV disease; $n = 1364$). We previously showed an increased genetic loading among 1364 cases with Stage B endometriosis compared with 1666 with Stage A disease (3), which led to two GWA analyses, including (i) 3194 ‘all’ endometriosis case and (ii) 1364 Stage B cases (Table 3). The genotyped data were imputed up to 1000 Genomes pilot reference panel (B36, June 2010) and the GWAS was performed again, using a missing data likelihood in a logistic regression model including

Table 3. Summary description of the GWAS used in the genetic enrichment analysis

GWAS	Consortium	Sample size	No. of SNPs (million)	References
Endometriosis— all cases	IEC	3194 cases, 7060 controls	~12.5	Painter <i>et al.</i> (3)
Endometriosis— Stage B cases	IEC	1363 cases, 7060 controls	~12.5	Painter <i>et al.</i> (3)
WHRadjBMI	GIANT	77 167	~2.85	Heid <i>et al.</i> (6)
Female-limited WHRadjBMI	GIANT	42 969	~2.85	Randall <i>et al.</i> (7)
BMI	GIANT	123 865	~2.85	Speliotes <i>et al.</i> (10)
Female-limited BMI	GIANT	73 137	~2.85	Randall <i>et al.</i> (7)

IEC, International Endogene Consortium; GIANT, Genetic Investigation of Anthropometric Traits Consortium; BMI, body mass index adjusted for age; WHRadjBMI, waist to hip ratio adjusted for BMI and age.

a covariate representing the Australian and the UK strata, with the imputed data ($N = 12.5$ million SNPs). The enrichment analysis we present is from this set of results.

GIANT Consortium

WHR GWAS. A total of 77 167 subjects of European ancestry informative of body fat distribution measurement WHR from 32 GWAS were included (6). The genotype data were imputed up to HapMap 2 CEU reference panel. The associations of 2.85 million SNPs with WHR were examined in a fixed-effects meta-analysis, after inverse normal transformation of WHR and adjusting for BMI and age within each study in an additive genetic model; analyses were conducted for males and females combined (6) and limited to females only (7) (Table 3).

BMI GWAS. A total of 123 865 subjects with overall adiposity measurement BMI from 46 GWAS were included (10). The genotype data were imputed up to HapMap two CEU reference panels. The associations of 2.85 million SNPs with BMI were tested in an inverse-variance meta-analysis, after inverse normally transformation of BMI and adjusting for age and other appropriate covariates in an additive genetic model within each study; analyses were conducted for males and females combined (10) and limited to females only (7) (Table 3).

Genetic enrichment analysis

With one test of association conducted for each SNP, the GWAS analyses produced a genome-wide distribution of P -values of individual SNP associations. Prior to testing enrichment: (i) the overlap of SNPs present in endometriosis GWAS versus WHRadjBMI and BMI GWAS was taken, (ii) all SNPs with $MAF \leq 0.01$ were removed, (iii) all SNPs with A/T and C/G base pairs were removed, (iv) correlated SNPs ($r^2 > 0.2$) were removed as previously reported (41) by taking the most significantly associated SNP and eliminating all SNPs that have a HapMap CEU pairwise correlation coefficient (r^2) > 0.2 with that SNP, then processing to the next strongly associated SNP remaining. This resulted in 173 157 independent SNPs in endometriosis versus WHRadjBMI and 173 223 in endometriosis versus BMI enrichment analyses.

The independent SNPs in the tails ($P < 1 \times 10^{-3}$) of the association results distribution of the two endometriosis GWAS (all endometriosis and 'Stage B' cases) were investigated for enrichment of WHRadjBMI or BMI low P -value ($P < 0.05$) association signals; in reversal, SNPs in the tails of WHRadjBMI and BMI GWAS ($P < 1 \times 10^{-3}$) were investigated for evidence of nominal association ($P < 0.05$) in the two endometriosis GWAS. The threshold of $P < 1 \times 10^{-3}$ corresponded to the point at which endometriosis GWAS results started to deviate from the null distribution (evidence for association) in the overall and Stage B endometriosis Q–Q plots (Supplementary Material, Fig. S4). Enrichment was assessed in R by means of Pearson's χ^2 tests with Yates' continuity correction, testing for the difference in proportion of SNPs with association $P < 0.05$ in the lookup dataset according to association in the discovery dataset ($P < 1 \times 10^{-3}$ versus $P \geq 1 \times 10^{-3}$). To test for consistency in directionality of phenotypic effects of the SNPs with evidence of enrichment, linear regression analysis was performed on the effect (β) of each SNP for WHRadjBMI as

predictor variable and the effect (β) of endometriosis risk as the outcome variable (35). In addition, a two-sided binomial test was performed with null hypothesis $P = 0.50$.

Permutation-based enrichment analysis

For those results that showed nominally significant ($P < 0.10$) evidence for enrichment in χ^2 tests of contingency tables, we performed permutation-based analyses to obtain empirical estimates of significance of enrichment. We (i) randomly picked the same number of independent SNPs 'associated' with the discovery trait at $P < 1 \times 10^{-3}$ (e.g. the number of SNPs associated with all endometriosis at $P < 1 \times 10^{-3}$ was $n = 717$) from the WHRadjBMI dataset; (ii) counted how many of the randomly selected SNPs had P -values of association with WHRadjBMI < 0.05 ; (iii) repeated Steps (i) and (ii) 10 000 times; (iv) determined the number of instances among the 10 000 draws in which the number of SNPs associated at $P < 0.05$ with WHRadjBMI was greater or equal to the number we observed in our original analysis (e.g. $\geq 52/717$). For example, for overall endometriosis and overall WHRadjBMI, we observed this in 26/10 000 instances, corresponding to a P -value of 2.6×10^{-3} , which was very similar to the P -value obtained from the χ^2 test ($P = 3.7 \times 10^{-3}$).

Polygenic prediction analysis

The independent SNPs in both WHRadjBMI and endometriosis datasets were used to conduct a polygenic prediction analysis (11). The aim of this analysis was to evaluate the aggregate effects of many SNPs of small effect and assess whether subsets of SNPs selected in this manner from one disease/trait GWAS predict disease/trait status in another, thus providing a measure of a common polygenic component with concordant directions of effect underlying the traits. Briefly, subsets of SNPs were selected from the WHRadjBMI GWAS data based on their association with WHRadjBMI using increasingly liberal thresholds, that is, $P < 0.01$, $P < 0.05$, $P < 0.1$, $P < 0.2$, $P < 0.3$, $P < 0.4$, $P < 0.5$ and $P < 0.75$. Using these thresholds, we defined sets of allele-specific scores in the WHRadjBMI dataset to generate risk profile scores for individuals in the endometriosis dataset. For each individual in the endometriosis dataset, we calculated the number of score alleles they possessed, each weighted by their effect size (β -value) of association in the WHRadjBMI dataset. To assess whether the aggregate scores were associated with endometriosis risk, we tested for a higher mean score in cases compared with controls. Logistic regression was used to assess the relationship between endometriosis disease status and aggregate risk score.

Expression analyses

MGEx-Udb

The mammalian gene expression uterus database (MGEx-Udb) is a manually curated uterus-specific database created using a meta-analysis approach from published papers (28) that provides lists of transcribed and dormant genes for various normal, pathological (e.g. endometriosis, cervical cancer and endometrial cancer) and experimental (e.g. treatment and

knockout) conditions. Each gene's expression status is indicated by a reliability score, derived based on the consensus across multiple samples and studies which highly variable (<http://resource.ibab.ac.in/MGEx-Udb/>).

MuTHER

The MuTHER resource includes LCLs, skin and adipose tissue-derived simultaneously from a subset of well-phenotyped healthy female twins (29). Whole-genome expression profiling of the samples, each with either two or three technical replicates, was performed using the Illumina Human HT-12 V3 BeadChips (Illumina, Inc.) according to the protocol supplied by the manufacturer. Log₂ transformed expression signals were normalized separately per tissue as follows: quantile normalization was performed across technical replicates of each individual followed by quantile normalization across all individuals.

Genotyping was conducted using a combination of Illumina arrays (HumanHap300, HumanHap610Q, 1M-Duo and 1.2MDuo 1 M). Untyped HapMap2 SNPs were imputed using the IMPUTE software package (v2). In total, there were 776 samples with genotypes and expression values in adipose tissue. Association between all SNPs (MAF > 5%, IMPUTE info score > 0.8) within a gene or within 1 Mb of the gene transcription start or end site, and normalized expression values, were performed with the GenABEL/ProbABEL packages (42) using polygenic linear models incorporating a kinship matrix (GenABEL) followed by the mm score test with imputed genotypes (ProbABEL). Age and experimental batch were included as cofactors in the analysis. Benjamini Hochberg corrected *P*-values are reported.

MolOBB

We performed differential *cis*-eQTL analysis to compare the expression levels in gluteal and abdominal fat tissue from 49 individuals in the MolOBB dataset (24 with and 25 without metabolic syndrome—MetSyn) (30). We first checked for the presence of the SNP in the MolOBB genotype data and, in the case of absence, selected any proxies ($r^2 > 0.8$) available. We then searched for nearby genes (± 500 kb) covered by the expression data using the bioconductor R package GenomicRanges (43) and tested for association at each pair using a linear model with the expression level as an outcome and the SNP allelic dosage as a predictor, adjusting for age, gender and MetSyn case–control status. This analysis was carried out for both abdominal and gluteal subcutaneous adipose tissue. To investigate whether genes were differentially expressed between the two tissues, we applied a linear mixed model with tissue, MetSyn case–control status, gender and plate were as fixed effects, and subject as a random effect using MAANOVA (44), as previously described in Min *et al.* (30). We report the uncorrected and genome-wide FDR corrected *F*s test *P*-values (30).

Biological pathway analysis

PANTHER

We conducted analyses using the PANTHER 8.1 database containing pathway information on 20 000 genes (*Homo sapiens*) (32). We selected independent SNPs, which had association *P*-values < 1×10^{-3} in the endometriosis datasets and an association *P*-value of < 0.05 in the WHRadjBMI dataset, resulting

in (i) 91 SNPs for all endometriosis and WHRadjBMI and (ii) 108 SNPs for Stage B endometriosis and WHRadjBMI. Each SNP was mapped to the closest gene within 1 Mb; 88 of 91 and 103 of 108 genes were present in the PANTHER database, and these subsets were tested for correlation with 241 biological processes and 176 pathways classified in the database (32). For each biological process/pathway, the difference between the observed fraction of genes in that pathway and the number expected by chance was tested using Fisher exact test. A Bonferroni correction was used as a conservative method for adjusting for the maximum number of biological processes ($n = 278$; $P = 1.80 \times 10^{-4}$) and pathways ($n = 78$; $P = 6.41 \times 10^{-4}$) tested.

SUPPLEMENTARY MATERIAL

Supplementary Material is available at *HMG* online.

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REFERENCES

- Giudice, L.C. and Kao, L.C. (2004) Endometriosis. *Lancet*, **364**, 1789–1799.
- Treloar, S.A., O'Connor, D.T., O'Connor, V.M. and Martin, N.G. (1999) Genetic influences on endometriosis in an Australian twin sample. *sueT@qimr.edu.au. Fertil. Steril.*, **71**, 701–710.
- Painter, J.N., Anderson, C.A., Nyholt, D.R., Macgregor, S., Lin, J., Lee, S.H., Lambert, A., Zhao, Z.Z., Roseman, F., Guo, Q. *et al.* (2011) Genome-wide association study identifies a locus at 7p15.2 associated with endometriosis. *Nat. Genet.*, **43**, 51–54.
- Uno, S., Zembutsu, H., Hirasawa, A., Takahashi, A., Kubo, M., Akahane, T., Aoki, D., Kamatani, N., Hirata, K. and Nakamura, Y. (2010) A genome-wide association study identifies genetic variants in the CDKN2BAS locus associated with endometriosis in Japanese. *Nat. Genet.*, **42**, 707–710.
- Nyholt, D.R., Low, S.K., Anderson, C.A., Painter, J.N., Uno, S., Morris, A.P., MacGregor, S., Gordon, S.D., Henders, A.K., Martin, N.G. *et al.* (2012) Genome-wide association meta-analysis identifies new endometriosis risk loci. *Nat. Genet.*, **44**, 1355–1359.
- Heid, I.M., Jackson, A.U., Randall, J.C., Winkler, T.W., Qi, L., Steinthorsdottir, V., Thorleifsson, G., Zillikens, M.C., Speliotes, E.K., Magi, R. *et al.* (2010) Meta-analysis identifies 13 new loci associated with waist-hip ratio and reveals sexual dimorphism in the genetic basis of fat distribution. *Nat. Genet.*, **42**, 949–960.
- Randall, J.C., Winkler, T.W., Kutalik, Z., Berndt, S.I., Jackson, A.U., Monda, K.L., Kilpelainen, T.O., Esko, T., Magi, R., Li, S. *et al.* (2013) Sex-stratified genome-wide association studies including 270,000 individuals show sexual dimorphism in genetic loci for anthropometric traits. *PLoS Genet.*, **9**, e1003500.
- Shah, D.K., Correia, K.F., Vitonis, A.F. and Missmer, S.A. (2013) Body size and endometriosis: results from 20 years of follow-up within the Nurses' Health Study II prospective cohort. *Hum. Reprod.*, **28**, 1783–1792.
- McCann, S.E., Freudenheim, J.L., Darrow, S.L., Batt, R.E. and Zielezny, M.A. (1993) Endometriosis and body fat distribution. *Obstet. Gynecol.*, **82**, 545–549.
- Speliotes, E.K., Willer, C.J., Berndt, S.I., Monda, K.L., Thorleifsson, G., Jackson, A.U., Lango Allen, H., Lindgren, C.M., Luan, J., Magi, R. *et al.* (2010) Association analyses of 249,796 individuals reveal 18 new loci associated with body mass index. *Nat. Genet.*, **42**, 937–948.
- International Schizophrenia, C., Purcell, S.M., Wray, N.R., Stone, J.L., Visscher, P.M., O'Donovan, M.C., Sullivan, P.F. and Sklar, P. (2009) Common polygenic variation contributes to risk of schizophrenia and bipolar disorder. *Nature*, **460**, 748–752.
- Teng, J., Rai, T., Tanaka, Y., Takei, Y., Nakata, T., Hirasawa, M., Kulkarni, A.B. and Hirokawa, N. (2005) The KIF3 motor transports N-cadherin and organizes the developing neuroepithelium. *Nat. Cell. Biol.*, **7**, 474–482.
- Nelson, W.J. and Nusse, R. (2004) Convergence of Wnt, beta-catenin, and cadherin pathways. *Science*, **303**, 1483–1487.
- Qin, L., Chen, Y., Niu, Y., Chen, W., Wang, Q., Xiao, S., Li, A., Xie, Y., Li, J., Zhao, X. *et al.* (2010) A deep investigation into the adipogenesis mechanism: profile of microRNAs regulating adipogenesis by modulating the canonical Wnt/beta-catenin signaling pathway. *BMC Genomics*, **11**, 320.
- Wang, Y., van der Zee, M., Fodde, R. and Blok, L.J. (2010) Wnt/beta-catenin and sex hormone signaling in endometrial homeostasis and cancer. *Oncotarget*, **1**, 674–684.
- Vainio, S., Heikkila, M., Kispert, A., Chin, N. and McMahon, A.P. (1999) Female development in mammals is regulated by Wnt-4 signalling. *Nature*, **397**, 405–409.
- Matsuzaki, S., Darcha, C., Maleysson, E., Canis, M. and Mage, G. (2010) Impaired down-regulation of E-cadherin and beta-catenin protein expression in endometrial epithelial cells in the mid-secretory endometrium of infertile patients with endometriosis. *J. Clin. Endocrinol. Metab.*, **95**, 3437–3445.
- Matsuzaki, S. and Darcha, C. (2013) Involvement of the Wnt/beta-catenin signaling pathway in the cellular and molecular mechanisms of fibrosis in endometriosis. *PLoS ONE*, **8**, e76808.
- Boudeau, J., Baas, A.F., Deak, M., Morrice, N.A., Kieloch, A., Schutkowski, M., Prescott, A.R., Clevers, H.C. and Alessi, D.R. (2003) MO25alpha/beta interact with STRADalpha/beta enhancing their ability to bind, activate and localize LKB1 in the cytoplasm. *EMBO J.*, **22**, 5102–5114.
- Chevillard, G. and Blank, V. (2011) NFE2L3 (NRF3): the Cinderella of the Cap'n'Collar transcription factors. *Cell. Mol. Life Sci.*, **68**, 3337–3348.
- Berezat, V., Kasus-Jacobi, A., Perdereau, D., Cariou, B., Girard, J. and Burnol, A.F. (2002) Inhibition of insulin receptor catalytic activity by the molecular adapter Grb14. *J. Biol. Chem.*, **277**, 4845–4852.
- Reilly, J.F., Mickey, G. and Maher, P.A. (2000) Association of fibroblast growth factor receptor 1 with the adaptor protein Grb14. Characterization of a new receptor binding partner. *J. Biol. Chem.*, **275**, 7771–7778.
- Morris, A.P., Voight, B.F., Teslovich, T.M., Ferreira, T., Segre, A.V., Steinthorsdottir, V., Strawbridge, R.J., Khan, H., Grallert, H., Mahajan, A. *et al.* (2012) Large-scale association analysis provides insights into the genetic architecture and pathophysiology of type 2 diabetes. *Nat. Genet.*, **44**, 981–990.
- Scott, R.A., Lagou, V., Welch, R.P., Wheeler, E., Montasser, M.E., Luan, J., Magi, R., Strawbridge, R.J., Rehnberg, E., Gustafsson, S. *et al.* (2012) Large-scale association analyses identify new loci influencing glycemic traits and provide insight into the underlying biological pathways. *Nat. Genet.*, **44**, 991–1005.
- Ahmadian, M., Suh, J.M., Hah, N., Liddle, C., Atkins, A.R., Downes, M. and Evans, R.M. (2013) PPARgamma signaling and metabolism: the good, the bad and the future. *Nat. Med.*, **19**, 557–566.
- Lebovic, D.I., Mwenda, J.M., Chai, D.C., Santi, A., Xu, X. and D'Hooghe, T. (2010) Peroxisome proliferator-activated receptor-(gamma) receptor ligand partially prevents the development of endometrial explants in baboons: a prospective, randomized, placebo-controlled study. *Endocrinology*, **151**, 1846–1852.
- Loo, B.B., Darwish, K.K., Vainikka, S.S., Saarikettu, J.J., Vihko, P.P., Hermonen, J.J., Goldman, A.A., Alitalo, K.K. and Jalkanen, M.M. (2000) Production and characterization of the extracellular domain of recombinant human fibroblast growth factor receptor 4. *Int. J. Biochem. Cell. Biol.*, **32**, 489–497.
- Bajpai, A.K., Davuluri, S., Chandrashekar, D.S., Ilakya, S., Dinakaran, M. and Acharya, K.K. (2012) MGEx-Udb: a mammalian uterus database for expression-based cataloguing of genes across conditions, including endometriosis and cervical cancer. *PLoS ONE*, **7**, e36776.
- Grundberg, E., Small, K.S., Hedman, A.K., Nica, A.C., Buil, A., Keildson, S., Bell, J.T., Yang, T.P., Meduri, E., Barrett, A. *et al.* (2012) Mapping cis- and trans-regulatory effects across multiple tissues in twins. *Nat. Genet.*, **44**, 1084–1089.
- Min, J.L., Nicholson, G., Halgrimsdottir, I., Almstrup, K., Petri, A., Barrett, A., Travers, M., Rayner, N.W., Magi, R., Pettersson, F.H. *et al.* (2012) Coexpression network analysis in abdominal and gluteal adipose tissue reveals regulatory genetic loci for metabolic syndrome and related phenotypes. *PLoS Genet.*, **8**, e1002505.
- Rantalainen, M., Herrera, B.M., Nicholson, G., Bowden, R., Wills, Q.F., Min, J.L., Neville, M.J., Barrett, M., Allen, M., Rayner, N.W. *et al.* (2011) MicroRNA expression in abdominal and gluteal adipose tissue is associated with mRNA expression levels and partly genetically driven. *PLoS ONE*, **6**, e27338.
- Mi, H., Muruganujan, A. and Thomas, P.D. (2013) PANTHER in 2013: modeling the evolution of gene function, and other gene attributes, in the context of phylogenetic trees. *Nucleic Acids Res.*, **41**, D377–D386.
- Raychaudhuri, S., Plenge, R.M., Rossin, E.J., Ng, A.C., International Schizophrenia, C., Purcell, S.M., Sklar, P., Scolnick, E.M., Xavier, R.J., Altshuler, D. *et al.* (2009) Identifying relationships among genomic disease regions: predicting genes at pathogenic SNP associations and rare deletions. *PLoS Genet.*, **5**, e1000534.
- ASRM. (1997) Revised American Society for Reproductive Medicine classification of endometriosis. *Fertil. Steril.*, **67**, 817–821.
- Do, R., Willer, C.J., Schmidt, E.M., Sengupta, S., Gao, C., Peloso, G.M., Gustafsson, S., Kanoni, S., Ganna, A., Chen, J. *et al.* (2013) Common variants associated with plasma triglycerides and risk for coronary artery disease. *Nat. Genet.*, **45**, 1345–1352.
- Andreassen, O.A., Harbo, H.F., Wang, Y., Thompson, W.K., Schork, A.J., Mattingsdal, M., Zuber, V., Bettella, F., Ripke, S., Kelsoe, J.R. *et al.* (2014) Genetic pleiotropy between multiple sclerosis and schizophrenia but not bipolar disorder: differential involvement of immune-related gene loci. *Mol. Psychiatry*. doi:10.1038/mp.2013.195.
- Solovieff, N., Cotsapas, C., Lee, P.H., Purcell, S.M. and Smoller, J.W. (2013) Pleiotropy in complex traits: challenges and strategies. *Nat. Rev. Genet.*, **14**, 483–495.

38. Sivakumaran, S., Agakov, F., Theodoratou, E., Prendergast, J.G., Zgaga, L., Manolio, T., Rudan, I., McKeigue, P., Wilson, J.F. and Campbell, H. (2011) Abundant pleiotropy in human complex diseases and traits. *Am. J. Hum. Genet.*, **89**, 607–618.
39. Smith, G.D. and Ebrahim, S. (2003) 'Mendelian randomization': can genetic epidemiology contribute to understanding environmental determinants of disease? *Int. J. Epidemiol.*, **32**, 1–22.
40. Voight, B.F., Peloso, G.M., Orho-Melander, M., Frikke-Schmidt, R., Barbalic, M., Jensen, M.K., Hindy, G., Holm, H., Ding, E.L., Johnson, T. et al. (2012) Plasma HDL cholesterol and risk of myocardial infarction: a mendelian randomisation study. *Lancet*, **380**, 572–580.
41. Lindgren, C.M., Heid, I.M., Randall, J.C., Lamina, C., Steinthorsdottir, V., Qi, L., Speliotes, E.K., Thorleifsson, G., Willer, C.J., Herrera, B.M. et al. (2009) Genome-wide association scan meta-analysis identifies three loci influencing adiposity and fat distribution. *PLoS Genet.*, **5**, e1000508.
42. Aulchenko, Y.S., Ripke, S., Isaacs, A. and van Duijn, C.M. (2007) GenABEL: an R library for genome-wide association analysis. *Bioinformatics*, **23**, 1294–1296.
43. Aboyoun, P.H. and Lawrence, M. (2013) GenomicRanges: representation and manipulation of genomic intervals. R Package Version, 1.8.7.
44. Wu, H., Cui, X. and Churchill, G.A. (2002) MAANOVA: a software package for the analysis of spotted cDNA microarray experiments. In Parmigiani, G. and Garrett, E.S. (eds), *The Analysis of Gene Expression Data*. Springer-Verlag, New York, USA, pp. 313–341.

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