Pathologie 2023 · 44 (Suppl 3):S128–S135 https://doi.org/10.1007/s00292-023-01260-y Accepted: 19 October 2023 Published online: 4 December 2023 © The Author(s) 2023



What is new in the classification of peripheral T cell lymphomas?

Laurence de Leval 💿 · Bettina Bisig 💿

Institute of Pathology, Department of Laboratory Medicine and Pathology, Lausanne University Hospital (CHUV) and Lausanne University, Lausanne, Switzerland

Abstract

In this review focus article, we highlight the main modifications introduced in the latest 2022 International Consensus Classification and World Health Organization classification (ICC and WHO-HAEM5) of mature T (and NK) cell neoplasms (PTCLs) and consequent implications for diagnostic practice. The changes result from recent advances in the genomic and molecular characterization of PTCLs and enhanced understanding of their pathobiology. Specifically, consideration is given to the following groups of diseases: Epstein–Barr virus (EBV)-associated neoplasms; follicular helper T cell lymphoma; anaplastic large cell lymphomas; primary intestinal T and NK cell lymphomas and lymphoproliferative disorders; and PTCL, not otherwise specified.

Keywords

Follicular helper T cell lymphoma · Anaplastic large cell lymphomas · Intestinal T and NK cell lymphomas · Mutations · Cell of origin

Neoplasms derived from mature NK or T cells (peripheral T cell lymphomas, PTCLs) are rare overall, but encompass diverse clinical presentations of diseases ranging from uncommonly indolent to usually aggressive. The two classifications of lymphoid neoplasms developed in 2022, namely the International Consensus Classification (ICC) [5] and the fifth edition of the World Health Organization (WHO) classification (WHO-HAEM5) [1], represent updates of the 2017 revised fourth WHO classification (WHO-HAEM4R) (Fig. 1), and rely on a multiparametric definition of lymphoma entities. Recent advances in refining the clinicopathologic features and molecular and genomic profiling of PTCLs have translated into adjustments and changes introduced in both proposals which are largely overlapping, overall reflecting similar conceptual shifts, with slight differences.

EBV-associated T cell and NK cell neoplasms

In the group of Epstein–Barr virus (EBV)driven lymphoproliferative disorders of childhood [21], hydroa vacciniforme lymphoproliferative disorder (LPD) replaces what was previously designated hydroa vacciniforme-like LPD, because essentially all such lesions are associated with EBV infection. The ICC further recognizes two variants: a classic indolent form (limited to the skin) and a systemic aggressive form of the disease, more common in non-Caucasians. Chronic active EBV disease now replaces chronic active EBV infection to denote a pathologic disease condition, in line with the notion that pathogenic mutations indicating a neoplastic process are detected in a subset of patients.

Die Pathologie

The terminology, definition, and diagnostic criteria of extranodal NK/T cell lymphoma (ENKTCL) nasal type are unchanged in the ICC. Since this lymphoma is known to occur at various extranodal sites besides the nasal area which is involved in typical cases, "nasal type" was dropped in the WHO-HAEM5.

Cases of primary nodal EBV-positive T cell or NK cell lymphoma, formerly considered as a subtype of PTCL, not otherwise specified (NOS), are now categorized as a separate entity, namely primary nodal EBV+ T cell/NK cell lymphoma, provisional in the ICC, or nodal EBV+ T and NK cell



Scan QR code & read article online

| WHO-HAEM4R-2017 | ICC-2022 | WHO-HAEM5-2022 |
|--|---|---|
| T-cell prolymphocytic leukemia | T-cell prolymphocytic leukaemia | T-prolymphocytic leukemia |
| T-cell large granular lymphocytic leukaemia | T-cell large granular lymphocytic leukaemia | T-large granular lymphocytic leukaemia |
| Chronic lymphoproliferative | Chronic lymphoproliferative | NK-large granular lymphocytic |
| disorder of NK cells | disorder of NK cells | leukaemia |
| Adult T-cell leukemia/lymphoma | Adult T-cell leukemia/lymphoma | Adult T-cell leukemia/lymphoma |
| EBV-positive T-cell/NK-cell | EBV-positive T-cell/NK-cell | EBV-positive T- and NK-cell |
| lymphoproliferative disorders of | lymphoproliferative disorders of | lymphoid proliferations and |
| childhood | <u>childhood</u> | lymphomas of childhood |
| Hydroa vacciniforme-like | Hydroa vacciniforme | Hydroa vacciniforme |
| lymphoproliferative disorder | lymphoproliferative disorder - Classic type and systemic | lymphoproliferative disorder |
| | type | |
| Severe mosquito bite allergy | Severe mosquito bite allergy | Severe mosquito bite allergy |
| Chronic active EBV infection of T- | Chronic active EBV disease, | Systemic chronic active EBV |
| and NK-cell type, systemic form | systemic (T-cell and NK-cell phenotype) | disease |
| Systemic EBV-positive T-cell | Systemic EBV-positive T-cell | Systemic EBV-positive T-cell |
| lymphoma of childhood | lymphoma of childhood | lymphoma of childhood |
| Extranodal NK/T-cell lymphoma, nasal type | Extranodal NK/T-cell lymphoma, nasal type | Extranodal NK/T-cell lymphoma |
| Aggressive NK-cell leukemia | Aggressive NK-cell leukemia | Aggressive NK-cell leukemia |
| Not listed as an entity, subtype | Primary nodal EBV+ T-cell/NK-cell | EBV+ nodal T- and NK-cell |
| of peripheral T-cell lymphoma, | lymphoma | lymphoma |
| not otherwise specified (PTCL- | | |
| NOS) | | |
| Enteropathy-associated T-cell | Enteropathy-associated T-cell | Enteropathy-associated T-cell |
| lymphoma | lymphoma | lymphoma |
| Not listed as an entity | Type II refractory celiac disease | Not listed as an entity |
| Monomorphic epitheliotropic | Monomorphic epitheliotropic | Monomorphic epitheliotropic |
| Intestinal I-cell lymphoma | Intestinal I-cell lymphoma | Intestinal I-cell lymphoma |
| Intestinal I-cell lymphoma, NOS | Intestinal I-cell lymphoma, NOS | Intestinal I-cell lymphoma, NOS |
| Indolent I-cell | Indolent cional I-cell | Indolent 1-cell lymphoma of the |
| the aastrointestingl tract | the gestrointestinal tract | gastrointestinai tract |
| Not listed | | Indolont NK-coll |
| Not listed | lymphoproliferative disorder of | lymphoproliferative disorder of |
| | the gastrointestinal tract | the gastrointestinal tract |
| Hepatosplenic T-cell lymphoma | Hepatosplenic T-cell lymphoma | Hepatosplenic T-cell lymphoma |
| Mycosis fungoides | Mycosis fungoides | Mycosis fungoides |
| Sezarv syndrome | Sezary syndrome | Sezarv syndrome |
| Primary cutaneous CD30+ T-cell | Primary cutaneous CD30+ T-cell | Primary cutaneous CD30+ T-cell |
| lymphoproliferative disorders | lymphoproliferative disorders | lymphoproliferative disorder: |
| - Lymphomatoid papulosis | - Lymphomatoid papulosis | Lymphomatoid papulosis |
| - Primary cutaneous | - Primary cutaneous anaplastic | Primary cutaneous CD30+ T-cell |
| anaplastic large cell | large cell lymphoma | lymphoproliferative disorder: |
| lymphoma | | Primary cutaneous anaplastic large |
| | | cell lymphoma |

Fig. 1 ◄ Classification of mature T and NK cell neoplasms in the ICC [5] and WHO-HAEM5 [1] proposals (2022) with reference to the WHO-HAEM4R classification (2017). The entities are listed according to the order in which they appear in the ICC-2022. Colors denote groups of entities. Italics indicate the entities provisional in the WHO-HAEM4R and ICC-2022

| Primary cutaneous CD4+ | Primary cutaneous CD4+ | Primary cutaneous small/medium |
|---|---|---|
| small/medium T-cell | small/medium T-cell | CD4+ T-cell lymphoproliferative |
| lymphoproliferative disorder | lymphoproliferative disorder | disorder |
| Subcutaneous panniculitis-like T- | Subcutaneous panniculitis-like T- | Subcutaneous panniculitis-like T- |
| cell lymphoma | cell lymphoma | cell lymphoma |
| Primary cutaneous gamma-delta | Primary cutaneous gamma-delta | Primary cutaneous gamma-delta |
| T-cell lymphoma | T-cell lymphoma | T-cell lymphoma |
| Primary cutaneous acral CD8+ T- | Primary cutaneous acral CD8+ T- | Primary cutaneous acral CD8+ T- |
| cell lymphoma | cell lymphoproliferative disorder | cell lymphoproliferative disorder |
| Primary cutaneous CD8+ | Primary cutaneous CD8+ | Primary cutaneous CD8+ |
| aggressive epidermotropic | aggressive epidermotropic | aggressive epidermotropic |
| cytotoxic T-cell lymphoma | cytotoxic T-cell lymphoma | cytotoxic T-cell lymphoma |
| Not listed | Not listed | Primary cutaneous peripheral T- |
| Peripheral T-cell lymphoma, NOS | Peripheral T-cell lymphoma, NOS | Peripheral T-cell lymphoma, NOS |
| <u>Nodal lymphomas of T follicular</u> <u>helper origin</u> Angioimmunoblastic T-cell lymphoma | Follicular helper T-cell lymphoma Follicular helper T-cell lymphoma, angioimmunoblastic type (angioimmunoblastic T-cell lymphoma) | <u>Nodal T-follicular helper (TFH) cell</u> <u>lymphoma</u> Nodal TFH cell lymphoma, angioimmunoblastic-type |
| Follicular T-cell lymphoma | Follicular helper T-cell lymphoma, follicular type | Nodal TFH cell lymphoma, follicular-type |
| Nodal peripheral T-cell lymphoma with T follicular helper phenotype | Follicular helper T-cell lymphoma, NOS | Nodal TFH cell lymphoma, NOS |
| Anaplastic large cell lymphoma, | Anaplastic large cell lymphoma, | ALK-positive anaplastic large cell |
| ALK-positive | ALK-positive | lymphoma |
| Anaplastic large cell lymphoma, | Anaplastic large cell lymphoma, | ALK-negative anaplastic large cell |
| ALK-negative | ALK-negative | lymphoma |
| Breast implant-associated | Breast implant-associated | Breast implant-associated |
| anaplastic large cell lymphoma | anaplastic large cell lymphoma | anaplastic large cell lymphoma |

Fig. 1 ◀ (Continued)

lymphoma in the WHO-HAEM5 [21]. This rare disease, most prevalent in East Asia, involves lymph nodes and is frequently disseminated but lacks nasal involvement and tends to occur in elderly adults, in association with HIV infection or immunodeficient conditions [14]. Pathological features distinct from ENKTCL include a monomorphic large cell morphology, less frequent necrosis, negativity for CD56, positivity for CD8, and more frequent derivation from T cells than from NK cells [14, 19]. The tumor is characterized by loss of 14g11.2, upregulation of immune pathways, low genomic instability and recurrent mutations involving the epigenetic modifiers, such as TET2 and DNMT3A, and JAK-STAT pathway genes [27].

Follicular helper T cell lymphoma

In 2017, the developing concept that follicular helper derivation represents a unifying feature of a large group of nodal CD4+ T cell lymphomas was reflected by the creation of an umbrella term "nodal T cell lymphoma of T follicular helper (TFH) origin" to encompass angioimmunoblastic T cell lymphoma, follicular T cell lymphoma, and nodal PTCL with T follicular helper phenotype. Since then, this notion has been reinforced by additional evidence indicating shared molecular and genetic features [10], and, importantly, clinical data suggest that this grouping might be relevant to treatment decisions, as TFH lymphoma appear more sensitive to epigenetic therapies than non-TFH PTCLs (**Fig. 2a**; [4]). Therefore, the ICC considers one single disease entity, namely follicular helper T cell lymphoma, comprising three subtypes, angioimmunoblastic, follicular, and NOS (Fig. 2b-d; [11]). This entity by definition excludes primary cutaneous CD4+ T cell lymphoproliferations which also feature a TFH phenotype. The WHO-HAEM5 proposal is more conservative, considering a family of three related entities of nodal T follicular helper cell lymphomas. The TFH immunophenotype is defined by the expression of at least two and ideally three TFH markers out of a panel of at least five markers (CD10, BCL6, PD1, ICOS, CXCL13) that it is now recommended to test for routinely and systematically when a diagnosis of TFH lymphoma is considered or must be excluded [1, 5]. TFH lymphomas frequently carry mutations in *TET2, DNMT3A, RHOA,* and *IDH2*, which are rarely seen in combination in other PTCL entities; hence, mutational testing may be diagnostically useful [8, 12].

Anaplastic large cell lymphomas

The four entities of anaplastic large cell lymphomas (ALCLs) are identical in both proposals: ALK-positive (ALK+) and ALKnegative (ALK−) ALCL, primary cutaneous ALCL (within the spectrum of CD30-positive cutaneous T cell lymphoproliferative disorders), and breast implant-associated (BIA-)ALCL. Among ALK− ALCLs, those with *DUSP22* rearrangement (25–30% of cases; **□** Fig. 3a–d) differ from those



Fig. 2 ▲ Follicular helper T cell lymphoma. a Evolution of the classification of T cell lymphomas of follicular helper T cell (TFH cell) derivation in the successive classifications. b Follicular helper T cell lymphoma of the angioimmunoblastic type (TFHL-AI) comprising a polymorphous cellular infiltrate with prominent vessels and expansion of CD21+ follicular dendritic cells. c Follicular helper T cell lymphoma of the follicular type (TFHL-F) is exemplified here by a case showing a follicular lymphoma-like appearance, comprising nodules of CD3+ T cells also positive for several TFH markers (not shown). d Follicular helper T cell lymphoma, not otherwise specified (TFHL-NOS), consists of a diffuse lymphoproliferation of atypical CD4+ cells expressing two or more TFH markers, shown here is CXCL13 expression (b-d HE and immunoperoxidase)

devoid of this alteration, as they usually lack JAK-STAT3 activation and EMA expression, less frequently express cytotoxic molecules, harbor *MSC* mutations in about one third of cases, and have distinctive transcriptomic signature and methylation profiles [17, 18]. The clinical impact of DUSP22 rearrangement remains controversial: the initially reported markedly superior prognosis of these cases was not confirmed in subsequent studies, while data from more recent cohorts still support an intermediate prognosis of DUSP22rearranged ALK– ALCL, standing between ALK+ ALCL and *DUSP22*-non rearranged ALK– ALCL [23, 24]. Taking into account its biological and prognostic peculiarities, the ICC recognizes *DUSP22*-rearranged ALCL as a genetically defined subtype of the disease and recommends systematic FISH testing for *DUSP22* in ALK– ALCL [8].



Fig. 3 ▲ Anaplastic large cell lymphomas. a–d ALK-negative *DUSP22*-rearranged anaplastic large cell lymphoma, showing a typical morphology including kidney-shaped (*arrow*) and doughnut-shaped (*arrowhead*) tumor cells (a), which are strongly and diffusely CD30 positive (b), and negative for ALK (not shown) and granzyme B (c). Break-apart FISH reveals *DUSP22* gene rearrangement (d *red* and *green arrows* point to split signals). e Breast implant-associated anaplastic large cell lymphoma diagnosed on a Papanicolaou-stained smear of a periprosthetic effusion, displaying large tumor cells with anaplastic nuclear features, admixed with inflammatory cells

Other structural aberrations are recurrent in ALK– ALCL but less common. These include *TP63* rearrangements, associated with an adverse prognosis [23]; as well as fusion genes involving tyrosine kinases such as *JAK2*, *FRK*, *ROS1*, and *TYK2*, which may represent potential therapeutic targets [8].

BIA-ALCL (**D** Fig. 3e) is recognized as a definitive entity both in the ICC and WHO-HAEM5. While histopathologically it largely overlaps with systemic ALK– ALCL, the pathogenetic association of BIA-ALCL with the microenvironment of textured breast implants is unique. At the genetic level, a highly characteristic 20q13.13 loss has been reported in two thirds of cases [7], and mutations in epigenetic modifiers such as *KMT2C*, *KMT2D*, and *CREBBP* are also frequently detected [16]. Similar to systemic ALCL, activation of the JAK-STAT3 pathway is a constant feature of BIA-ALCL, most commonly through mutations of *STAT3* and/or *JAK1* [16]. In contrast, rearrangements of *ALK*, *DUSP22*, or *TP63* associated with systemic ALCLs are not observed. The prognosis of BIA-ALCL is generally excellent after surgical removal of the periprosthetic fibrous capsule, but is less favorable in cases of infiltration of the adjacent breast parenchyma [15].

Primary intestinal T and NK cell lymphomas and lymphoproliferative disorders

The three main aggressive types of primary intestinal T cell lymphomas (enteropathy-associated T cell lymphoma [EATL], monomorphic epitheliotropic intestinal T cell lymphoma (MEITL), and intestinal T cell lymphoma, NOS) are unchanged [9]. EATL occurs in populations with a higher prevalence of HLA haplotypes predisposing to celiac disease, as a complication of celiac disease and refractory celiac disease, or de novo in individuals with no history of malabsorption. The tumors may be multiple and present as ul-



Fig. 4 ▲ Indolent NK cell lymphoproliferative disorder of the gastrointestinal tract. This colonic biopsy shows a diffuse mucosal infiltrate of atypical lymphoid cells with clear cytoplasm, partially obliterating the crypts (a and b). The cells are positive for TIA-1 (c), CD56 (d), and CD3 (e), and were negative for EBV (not shown)



Fig. 5 ◀ Algorithm for the diagnosis of nodal peripheral T cell lymphomas (PT-CLs). (Adapted from [5])

 Table 1
 Comparison of PTCL-GATA3 and PTCL-TBX21 subtypes of peripheral T cell lymphomas, not otherwise specified (PTCL, NOS) [3, 12]

| | PTCL-GATA3 30–40% | PTCL-TBX21 50–60% |
|---------------------------------|--|---|
| Gene expression signature | Th1 like MYC overexpression High proliferation PI3K activation | Th2 like Subset cytotoxic Enrichment of NF-kappa B path- way |
| Clinical | Poorer outcome | Better outcome Cytotoxic phenotype associated with poorer outcome |
| Morphology and phenotype | Less inflammatory background GATA3+ and/or CCR4+ (>50%) | Inflammatory background TBX21+ and/or CXCR3+ (>20%) |
| Genomics and gene expression | Higher genomic complexity Genomic aberrations include dele- tions of 17p (<i>TP53</i>), 9p (<i>CDKN2A</i>), and 10p (<i>PTEN</i>) | Fewer genomic aberrations, tar- geting cytotoxic effector genes Frequent mutations in epigenetic modulators (e.g., <i>TET2</i> , <i>DNMT3A</i>) |

cers or, less commonly, masses, comprise a polymorphous infiltrate with admixed inflammation, and often pleomorphic to anaplastic lymphoma cells. The typical immunophenotype is CD3+ CD4- CD8-CD30+/- TCR-silent EBV-negative, with expression of cytotoxic molecules. MEITL presents as a tumor mass and spans a morphologic spectrum. While typical cases are monomorphic with little necrosis, other tumors exhibit pleomorphic cytology and/or other atypical features like necrosis, brisk mitotic activity, and angiocentricity [26]. In MEITL, the neoplastic cells are CD3+ CD4- CD8+ CD56+ TCRpositive (gamma-delta more commonly than alpha-beta) EBV-negative. Genomic features may be helpful in differentiating between EATL and MEITL: alterations in the JAK/STAT pathway genes target primarily STAT3 and JAK1 in EATL, and STAT5B and JAK3 in MEITL. Deleterious alterations of the SETD2 gene, translating into reduced H3K36 trimethylation, are almost constant and rather specific to MEITL [22]. Type II refractory celiac disease has been added to the list of entities in the ICC, as this represents an "in situ" neoplastic condition precursor to EATL, and recent works have shown that it often already harbors driving mutations in JAK1 and/or STAT3 similar to those present in EATL [6].

The formerly provisional "indolent T cell LPD of the gastrointestinal tract" is confirmed in the ICC with the addition of "clonal" to emphasize its neoplastic nature. Indeed a variety of somatic genetic alterations have been found in these cases, including a recurrent JAK2::STAT3 fusion in a subset of CD4+ cases [25]. In WHO-HAEM5, the name has been modified to "indolent T cell lymphoma," given the fact that transformation into a high-grade PTCL has been described in some patients. Both proposals have created a new category to classify the indolent gastrointestinal LPD of NK cells (Fig. 4), which also carry a variety of genetic mutations, including a recurrent JAK3 small in-frame deletion [28]. These T and NK LPDs of the gastrointestinal tract are in general restricted to the mucosa and represent a diagnostic challenge and should not be confused, on the one hand with inflammatory conditions, on the other hand with aggressive lymphomas, since their course is usually indolent despite possible relapses, multifocality, and chronicity, and they do not respond to chemotherapy.

Peripheral T cell lymphomas, not otherwise specified

The group of PTCLs, not otherwise specified (NOS), remains a diagnosis of exclusion (**■** Fig. 5). Cases with a TFH immunophenotype must be excluded, since lymphomas with no morphologic specification but showing a TFH immunophenotype, defined by the expression of two or ideally three TFH markers, are classified as TFH lymphoma, NOS. Moreover, caution must be applied in this scenario to exclude primary cutaneous T cell lymphomas or human T-lymphotropic virus type 1 (HTLV-1)associated adult T leukemia/lymphoma, as these entities, which are often CD4+, may show expression of TFH markers [20].

Two biological subtypes of PTCL, NOS, namely PTCL-TBX21 and PTCL-GATA3, have been identified by gene expression profiling, and are characterized by overexpression of transcription factors TBX21 or GATA3 and corresponding target genes, with different prognoses and distinct oncogenic pathways ([12, 13]; **Table 1**). An immunohistochemical algorithm using four markers applied sequentially (TBX21, CXCR3, GATA3, and CCR4) can provide surrogate information on the molecular subtypes [3], and a digital nanostringbased assay has recently been published [2]. However, it is acknowledged that there is currently too little evidence to recommend molecular subtyping of PTCL, NOS, in routine clinical use [3]. PTCL-GATA3 demonstrates high genomic complexity characterized by biallelic deletion/ mutation of TP53, CDKN2A/B, or RB1, and carries a worse prognosis compared to PTCL-TBX21, which shows low genomic complexity and few recurrent specific genetic changes.

Conclusion

In conclusion, the updated classifications of T and NK cell neoplasms confirm the diversity and complexity of these disorders. Nevertheless, the accumulating knowledge of their biology is translated into more meaningful categories, and an increasing importance of molecular testing for precision diagnosis and tailored therapy.

Corresponding address

Prof. Laurence de Leval, MD PhD Institute of Pathology, Department of Laboratory Medicine and Pathology, Lausanne University Hospital (CHUV) and Lausanne University 25 rue du Bugnon, 1011 Lausanne, Switzerland laurence.deleval@chuv.ch

Funding. Open access funding provided by University of Lausanne

Declarations

Conflict of interest. L. de Leval and B. Bisig declare that they have no competing interests.

For this article no studies with human participants or animals were performed by any of the authors.

The supplement containing this article is not sponsored by industry.

Open Access. This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

References

- Alaggio R, Amador C, Anagnostopoulos I et al (2022) The 5th edition of the world health organization classification of haematolymphoid tumours: lymphoid neoplasms. Leukemia 36:1720–1748
- Amador C, Bouska A, Wright G et al (2022) Gene expression signatures for the accurate diagnosis of peripheral T-cell Lymphoma entities in the routine clinical practice. J Clin Oncol 40:4261–4275
- Amador C, Greiner TC, Heavican TB et al (2019) Reproducing the molecular subclassification of peripheral T-cell lymphoma-NOS by immunohistochemistry. Blood 134:2159–2170
- 4. Bachy E, Camus V, Thieblemont C et al (2022) Romidepsin plus CHOP versus CHOP in patients with previously untreated peripheral T-cell Lymphoma: results of the Ro-CHOP phase III study (conducted by LYSA). J Clin Oncol 40:242–251
- Campo E, Jaffe ES, Cook JR et al (2022) The international consensus classification of mature lymphoid neoplasms: a report from the clinical advisory committee. Blood 140:1229–1253
- Cording S, Lhermitte L, Malamut G et al (2022) Oncogenetic landscape of lymphomagenesis in coeliac disease. Gut 71:497–508
- 7. de Leval L (2020) Chromosomes in breast lymphoma. Blood 136:2848–2849
- de Leval L, Alizadeh AA, Bergsagel PL et al (2022) Genomic profiling for clinical decision making in lymphoid neoplasms. Blood 140:2193–2227
- 9. de Leval L, Feldman AL, Pileri S et al (2023) Extranodal T- and NK-cell lymphomas. Virchows Arch 482:245–264
- Dobay MP, Lemonnier F, Missiaglia E et al (2017) Integrative clinicopathological and molecular analyses of angioimmunoblastic T-cell lymphoma and other nodal lymphomas of follicular helper T-cell origin. Haematologica 102:e148–e151
- Feldman AL, Laurent C, Narbaitz M et al (2023) Classification and diagnostic evaluation of nodal T- and NK-cell lymphomas. Virchows Arch 482:265–279
- 12. Heavican TB, Bouska A, Yu J et al (2019) Genetic drivers of oncogenic pathways in molecular subgroups of peripheral T-cell lymphoma. Blood 133:1664–1676
- 13. lqbal J, Wright G, Wang C et al (2014) Gene expression signatures delineate biological and prognostic

Zusammenfassung

Was gibt es Neues bei der Klassifikation der peripheren T-Zell-Lymphomen?

In dem vorliegenden Übersichtsbeitrag liegt der Schwerpunkt auf den wesentlichen Modifikationen der aktuellen, 2022 eingeführten Klassifikation reifer T- (und NK-)Zell-Neoplasien, also peripherer T-Zell-Lymphome (PTCL) gemäß International Consensus Classification (ICC) und der Weltgesundheitsorganisation-Klassifikation (5. Auflage, WHO-HAEM5) sowie auf den daraus folgenden Auswirkungen auf die praktische Diagnostik. Die Veränderungen beruhen auf aktuellen Fortschritten in der genomischen und molekularen Charakterisierung der PTCL und einem vertieften Verständnis ihrer Pathobiologie. Insbesondere werden die folgenden Krankheitsgruppen berücksichtigt: Epstein-Barr-Virus(EBV)-assoziierte Neoplasien, follikuläres T-Helfer-Zell-Lymphom, anaplastische großzellige Lymphome, primäre intestinale T- und NK-Zell-Lymphome sowie lymphoproliferative Erkrankungen, außerdem nicht anderweitig spezifizierte PTCL.

Schlüsselwörter

Follikuläres T-Helfer-Zell-Lymphom · Anaplastische großzellige Lymphome · Intestinale T- und NK-Zell-Lymphome · Mutationen · Ursprungszelle

subgroups in peripheral T-cell lymphoma. Blood 123:2915–2923

- Kato S, Yamashita D, Nakamura S (2020) Nodal EBV+ cytotoxic T-cell lymphoma: a literature review based on the 2017 WHO classification. J Clin Exp Hematop 60:30–36
- Laurent C, Delas A, Gaulard P et al (2016) Breast implant-associated anaplastic large cell lymphoma: two distinct clinicopathological variants with different outcomes. Ann Oncol 27:306–314
- Laurent C, Nicolae A, Laurent C et al (2020) Gene alterations in epigenetic modifiers and JAK-STAT signaling are frequent in breast implant-associated ALCL. Blood 135:360–370
- Luchtel RA, Dasari S, Oishi N et al (2018) Molecular profiling reveals immunogenic cues in anaplastic large cell lymphomas with DUSP22 rearrangements. Blood 132:1386–1398
- Luchtel RA, Zimmermann MT, Hu G et al (2019) Recurrent MSC (E116K) mutations in ALKnegative anaplastic large cell lymphoma. Blood 133:2776–2789
- Nicolae A, Bouilly J, Lara D et al (2022) Nodal cytotoxic peripheral T-celllymphoma occurs frequently in the clinical setting of immunodysregulation and is associated with recurrent epigenetic alterations. Mod Pathol 35:1126–1136
- Ondrejka SL, Amador C, Climent F et al (2023) Follicular helper T-cell lymphomas: disease spectrum, relationship with clonal hematopoiesis, and mimics—a report of the 2022 EA4HP/SH lymphoma workshop. Virchows Arch 483:349–365
- 21. Quintanilla-Martinez L, Swerdlow SH, Tousseyn Tet al (2023) New concepts in EBV-associated B, T, and NK cell lymphoproliferative disorders. Virchows Arch 482:227–244
- 22. Roberti A, Dobay MP, Bisig B et al (2016) Type II enteropathy-associated T-cell lymphoma features a unique genomic profile with highly recurrent SETD2 alterations. Nat Commun 7:12602
- 23. Savage KJ, Slack GW (2023) DUSP22-rearranged ALK-negative anaplastic large cell lymphoma is a pathogenetically distinct disease but can have variable clinical outcome. Haematologica 108:1463–1467
- 24. Sibon D, Bisig B, Bonnet C et al (2023) ALK-negative anaplastic large cell lymphoma with DUSP22 rear-

rangement has distinctive disease characteristics with better progression-free survival: a LYSA study. Haematologica 108:1590–1603

- Soderquist CR, Patel N, Murty VV et al (2020) Genetic and phenotypic characterization of indolent T-cell lymphoproliferative disorders of the gastrointestinal tract. Haematologica 105:1895–1906
- Veloza L, Cavalieri D, Missiaglia E et al (2023) Monomorphic epitheliotropic intestinal T-cell lymphoma comprises morphologic and genomic heterogeneity impacting outcome. Haematologica 108:181–195
- Wai CMM, Chen S, Phyu Tet al (2022) Immune pathway upregulation and lower genomic instability distinguish EBV-positive nodal T/NK-cell lymphoma from ENKTL and PTCL-NOS. Haematologica 107:1864–1879
- Xiao W, Gupta GK, Yao J et al (2019) Recurrent somatic JAK3 mutations in NK-cell enteropathy. Blood 134:986–991

Publisher's Note. Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.