

Chromosome Size Differences May Affect Meiosis and Genome Size

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Transmission distortion has been identified by deviations from the equal representation of parental alleles in the progeny (1). Weak transmission bias and violations of the independent assortment of chromosomes are harder to detect, however, because large broods and a careful exam-

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X chromosome and of the shorter *wt* chromosome with the X. We refer to this phenomenon as skew. To exclude the possibility that skew for *mIs10* was specific to this genetic construct or chromosome V, we investigated segregation patterns for large insertion transgenes on the other four autosomes. All

Skew may affect genome evolution and genome size. Given that hermaphrodites tend to inherit the shorter chromosome and because new populations of *C. elegans* are frequently initiated by hermaphrodites (3), genome size reduction may occur over evolutionary time. Simulations of hermaphroditic mating, hermaphroditic selfing-only, and gonochoristic mating systems (2) (fig. S3) showed that skew led to a significant reduction in genome size in the hermaphroditic mating system ($P = 2.5 \times 10^{-13}$, one-sample *t* test) but not in the gonochoristic or hermaphroditic selfing-only systems (both $P > 0.27$, one-sample *t* test).

These observations of insertion disjoining from the X, causing genome size reduction in male/hermaphrodite relative to gonochoristic mating systems, is consistent with genome sizes in the genus *Caenorhabditis*. Hermaphroditism within the genus has evolved independently at least twice (4). All three male/female species tested have genome sizes of >130 Mb, whereas both sequenced hermaphroditic species have genome sizes of ~100 Mb (5–7). Thus, in at least two cases, the evolution of hermaphroditism was associated with a convergent reduction in genome size relative to the ancestral male/female species.

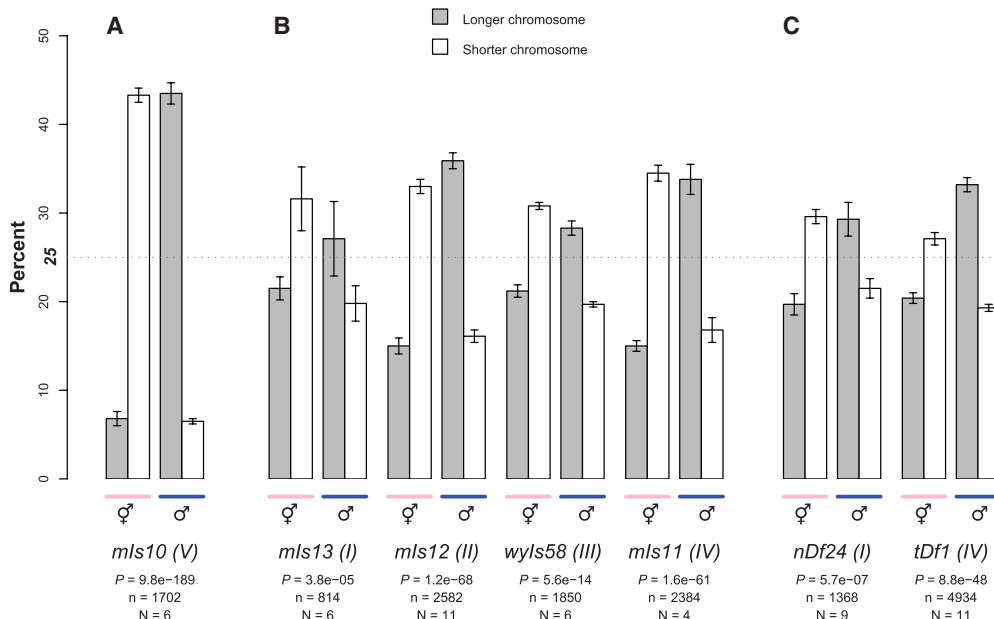


Fig. 1. Differential segregation of chromosomes with large insertions or deletions in hermaphrodite and male offspring. (A) *mIs10* transgene, (B) transgenes on chromosomes I to IV, and (C) deficiencies. The 25% line indicates expected values with no segregation bias. Error bars are SEM. Transgene or deficiency names and their chromosomal locations (roman numerals) are below respective bar plots. *P* values, χ^2 tests assuming random segregation of the two alternate chromosomes by sex. *N*, number of males tested; *n*, total individuals scored.

four insertion transgenes also preferentially segregated away from the X chromosome (Fig. 1B) with significantly differing magnitudes of skew among the insertion lines ($P < 2.2 \times 10^{-16}$, χ^2 test) and with transmission bias ratios ranging from 1.42 (*mIs13*) to 6.55 (*mIs10*). Skew also occurred in males heterozygous for large deletions (Fig. 1C, *nDf24* I and *tDf1* IV).

Because these insertions and deletions (indels) are relatively large (range ~615 kb to ~7.3 Mb, table S1), we tested whether smaller indels also exhibited skew by using a small insertion (*ruIs38*, ~33 kb) and three single-gene deletions (*unc-30*, *unc-47*, and *unc-63*, range from ~1 to ~1.5 kb). In the four cases, the shorter chromosome also segregated with the X chromosome, the skew being significant for *ruIs38*, *unc-47*, and *unc-63* (fig. S1). The magnitude of bias for these four indels (1.02 to 1.21) was smaller than for the larger indels. We also observed no difference regarding the parent of origin of an indel on skew (2). Overall, there was a significant positive correlation between indel size and transmission bias ratio (fig. S2, $P = 0.006$, Spearman's rank correlation test).

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References and Notes

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Supporting Online Material

www.sciencemag.org/cgi/content/full/329/5989/293/DC1
Materials and Methods
SOM Text
Figs. S1 to S3
Table S1
References

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