



Use of Dyes to Detect Fingermarks

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8 **Use of stains to detect fingerprints**
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Abstract

Detection of fingermarks at a crime scene or on related items is of prime interest for forensic investigators, mainly for identification purposes. Most of the fingermarks are invisible to the naked eye, however, and application of detection techniques is required to establish visual contrast between the secretion residue and the underlying substrate. We give here a review of the field related to the concept of using stains to detect fingermarks. A distinction has been made between the physically driven classical detection techniques, the chemically driven ones, and those based on nanostructured materials, an emerging field in forensic science.

Key words: amino acid, blood, detection method, dye, fingermark, fingerprint, forensic, science, identification, lipid, luminescence, nanoparticle, powder, trace evidence

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3 In a criminal investigation, one role of the forensic scientist is to collect information that
4 permits the identification of people involved in the events. Fingerprints constitute one of the
5 most important items of physical evidence for identification. A fingerprint is defined as a
6 unique pattern present on the fingertips of each individual as well as on the palms and the
7 soles. A fingerprint is characterized by invariability from birth to death, excluding scars and
8 irreversible skin damage that could occur during one's life. In this context, we chose to make a
9 distinction between fingerprints and fingermarks. Fingerprints are physically present on the
10 individual's body and can be inked on a ten-print card to be used for comparison in an
11 identification process. Fingermarks are defined as the secretion residue that is left by a person
12 when a surface is touched with unprotected hands. The origin of this residue is in the pores
13 along the skin surface through which human sweat is excreted. The chemical composition of
14 sweat varies according to the nature of the secreting gland and its location on the body
15 (Ramotowski 2001). In addition, contaminants such as cosmetics or other exogenous
16 components can be present on the fingertips due to contact of bare hands with one's face or
17 fatty food. As a consequence, the composition of the residue that is left on a substrate
18 generally is a complex mixture of organic and inorganic materials dispersed in an aqueous
19 matrix, e.g., amino acids, fatty acids, glycerides, inorganic ions, proteins and esters
20 (Ramotowski 2001). Most importantly, such residue generally is not visible to the naked eye.
21 In such situations the fingermarks are said to be "latent" and the application of detection
22 techniques is required to permit their visualization (Champod et al. 2004). This constitutes a
23 major and continuing challenge for forensic scientists.
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52 We address here the different techniques and reagents used to visualize latent
53 (invisible) fingermarks. The key principle of fingermark detection is simple: to generate as
54 great an optical contrast as possible between the ridges constituting the fingerprint pattern and
55 the underlying substrate without disturbing the pattern formed by the trace. This is necessary
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3 to ensure efficient visualization of the latent fingerprints, their ridges and their characteristics
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5 so they can be used for identification. For white or transparent substrates (e.g., printer paper
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7 or polypropylene plastic sheets), good contrast can be obtained readily by staining the residue
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9 using dark colored dyes. When dealing with colored, patterned or complex substrates, e.g.,
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11 magazines, commercial packages and banknotes, however, dyes are not appropriate, and the
12
13 use of a luminescent stain is necessary to avoid confusion due to the background pattern. The
14
15 use of luminescence to detect fingerprints also is generally preferred for sensitivity reasons.
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18 Actual fingerprints sometimes are composed of extremely small amounts of secretions due to
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20 a lack of secretion deposited or to further alterations caused by time and environmental
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22 conditions. This makes them difficult to detect when stained with classic chromophores. The
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24 luminescence mode, however, increases the sensitivity of detection so that faint fingerprints
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26 can be recorded and processed for identification. This also explains why current research
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28 projects in the field are directed primarily toward development of new and efficient
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30 luminescent detection techniques.
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36 To obtain good contrast, it is necessary for a stain to interact specifically with the
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38 secretion residue and not with the underlying substrate (or reciprocally, but this is less often
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40 the case). Because the composition of fingerprints varies according to various parameters that
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42 forensic scientists cannot control, e.g., sex and age of the donor, storage conditions of the
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44 evidence before collection, age of the fingerprint, or nature of the substrate, it is necessary to
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46 target compounds that are most likely to be found in sufficient quantities in almost all latent
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48 fingerprints. Numerous techniques have been developed that target organic and inorganic
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50 components with more or less success. Each technique generally is limited in its application
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52 according to the type of the substrate and the nature of the secretion. For example, 1,2-
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54 indanedione and ninhydrin are applied in the case of latent fingerprints on porous surfaces,
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56 cyanoacrylate fuming followed by the application of stains is the most efficient approach for
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3 nonporous ones, physical developer and oil red O are two techniques for detecting
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fingermarks when a substrate has been wet, acid yellow 7 and acid violet 17 are specific for blood-contaminated fingermarks, just to cite a few of several situations encountered. These limitations find their origin in the intrinsic interaction mechanisms between the reagents and the secretion residue.

We do not intend to review extensively all the existing fingerprint detection techniques. We shall instead provide a broad overview of the historical and routinely used methods as well as the techniques for development. Issues concerning application protocols, formulations, or comparative studies will not be covered. In the context of this review, we shall classify the techniques according to the kind of stain that is used. The reader must appreciate that we have adopted a very broad definition of the term, "stain." We define a stain as any organic molecule, chemical reagent or nanocomposite that permits the visualization of initially latent fingermarks through the creation of a visible contrast between the ridges and the underlying substrate. Note that this view is in keeping with definitions existing in the biological staining literature, such as that of Horobin (1988). The coloration of the secretion residue can be obtained through 1) physical incorporation of stains in the residue itself, e.g., using a lysochrome, 2) chemical reaction between an initially colorless reagent and some components of the secretion residue, e.g., the amino acids, or 3) physico-chemical processes leading to specific targeting of the residue, e.g., negatively charged metal nanoparticles, and its subsequent visualization. Consequently, we begin with stains that color the fingermarks without undergoing a chemical reaction, continue with stains that chemically react with some components of the secretion residue, and finally, describe techniques based on nanostructured materials, a growing field in forensic research.

Physical detection mechanisms

By physical detection, we mean colored molecules or particles that have a physical affinity for the fingerprints and permit their visualization without modification of their molecular structure (Voss-de Haan 2006).

Before addressing the issue of stains, it is worth mentioning that it often is possible to detect fingerprints before any treatment of the evidence by using only ambient light and varying the incident angle. Different optical techniques such as episcopic coaxial illumination (Reiss 1903, Pfister 1985, Ziv and Springer 1993), filtered light (Champod et al. 2004), or reflected ultraviolet (UV) light, i.e., the RUVIS system (Saferstein and Graf 2001) can be used to highlight marks on the substrates. Alternatively, some components of the secretions may show an intrinsic luminescence that can be visualized under UV light (Saitoh and Akiba 2005, 2006) or laser illumination (Dalrymple et al. 1977, Menzel 1999). Most of the time, these detection methods suffice when fresh fingerprints, rich in secretion, are laid on nonporous substrates. If the marks are weak, poor in secretion, old, or on a colored substrate, optical techniques must be extended by using stains.

Powder dusting

Used since the beginning of the last century, powder dusting (Reiss 1911) is one of the first techniques used to detect fingerprints, and it still is widely applied at crime scenes. The principle is simple: powder is dusted onto items using a brush so that it physically adheres to the moist, sticky and greasy substances of fingerprint residue (Lee and Gaensslen 2001, Champod et al. 2004). As a result, the contrast between the substrate and the dusted ridges permits observation of the latent fingerprints. This technique is inexpensive and easy to set up, but is limited to smooth and nonporous surfaces. Moreover, only fresh fingerprints give

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3 satisfactory results; the quality decreases as the fingermarks age owing to a loss of stickiness
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5 of their residues, which can be problematic.
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8 Conventional powders generally are composed of a contrasting agent, e.g., metal
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10 oxides or chromophoric dyes, plus resinous materials, e.g., stearic acid, for adhesion (Sodhi
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12 and Kaur 2001). The first colored dusting powders were obtained by using different metal
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14 oxides, sulfides and carbonates, e.g., lead, mercury or antimony. These compounds
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16 understandably were abandoned because of toxicity issues. Among the routinely used
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18 powders currently, a black one is generally applied on light colored substrates. The black
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20 color is obtained by including graphite, powdered charcoal, lamp black or molybdenum
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22 powder in their composition. On dark backgrounds, white powders, e.g., titanium dioxide,
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24 zinc oxide or zinc sulfide, are preferred (Goode and Morris 1983). Another widely used
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26 powder, regarded as one of the most effective (James et al. 1991a), is the so called
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28 "argenteratum" (Thomas 1973, 1975) composed of flat aluminium particles and 3-5% (w/w)
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30 stearic acid. Another commonly used preparation is a gold colored powder composed of
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32 bronze flakes. As their name indicates, magnetic powders contain a magnetic carrier in
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34 addition to a stain. They require the use of a magnetic brush for dusting, thus avoiding
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36 destructive contacts between brush hairs and the secretion residue (James et al. 1991b, 1992,
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38 1993). Luminescent powders are used mainly on multicolored backgrounds, when poor
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40 contrast might be expected with conventional powders. A great variety of luminescent dyes or
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42 pigments are available, e.g., acridine yellow, coumarin 6, or rhodamine 6G (Lee and
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44 Gaensslen 2001). Cyano blue dye, Nile red and proflavine also were tested recently as
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46 fluorescent dusting components (Sodhi and Kaur 2004, 2005, Kaur 2006).
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55 An alternative way of applying powders is as suspensions in aqueous solution, a
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57 method also known as "wet powdering" or "small particle reagent." This technique will be
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59 described below.
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Camphor

Camphor dusting may be considered an unconventional dusting method, because it requires the use of soot freshly obtained by burning it (Waldoch 1993). Camphor is a flammable white crystalline solid that produces a large amount of black soot when burned. The item to be "dusted" is maintained above the burning camphor to allow the black smoke to cover the entire surface. The soot adheres selectively to the fingerprint residues and the excess is removed with a brush or under tap water; the fingerprint appears black. Camphor may be helpful for some substrates such as firearms cartridges (Sturelle et al. 2006), but is not used for conventional surfaces.

Iodine fuming

Iodine (I_2) fuming was described very early (Coulier 1863) for detecting forgeries and fingerprints. A substrate bearing latent fingerprints is exposed to the purple iodine vapors provided by crystals of iodine contained in a glass vessel through which an air stream is forced to pass. The lipids present in the secretions physically absorb the vapors, which results in a temporary yellowish or brownish staining of the marks. Visualization is temporary due to the re-sublimation of iodine vapors from the marks to the ambient atmosphere after a few minutes. It is possible, however, to fix the stained fingerprints by a chemical reaction between iodine and 7,8-benzoflavone (α -naphthoflavone) (Mashito and Makoto 1977, Haque et al. 1983), which results in a permanent blue image. The mark also can be lifted by using the corrosive nature of the iodine. A highly polished silver or tin plate is pressed firmly on the mark for 5 sec, then exposed to a strong white light source or UV lamp. This leads to the formation of a dark image on the plate (Adcock 1977, Arndt 1985).

This technique can be applied to various porous and nonporous surfaces, especially on difficult ones like leather or human skin (Adcock 1977, Gray 1978). It should be emphasized

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3 that only fresh fingerprints can be observed by using this technique. It should also be
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5 mentioned that iodine vapors are toxic and corrosive.
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10 *Lipid dyes: oil red O and solvent black 3*

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12 The lipid fraction of secretion residues constitutes a good target for fingerprint detection,
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14 because it is possible to dye them specifically using lysochrome molecules currently used in
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16 biology. Moreover, in the case of fingerprints that have been wet, it is generally accepted that
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18 the water soluble components, e.g., ions, amino acids or proteins, are no longer present on the
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20 substrate due to their solubilization, while the water insoluble components, e.g., lipids, remain
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22 and can still be retrieved.
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27 Oil red O (ORO; solvent red 27) is a relatively new technique for detecting
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29 fingerprints on porous surfaces (Beaudoin 2004, Rawji and Beaudoin 2006, Guigui and
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31 Beaudoin 2007). ORO is a lysochrome used in biology to stain lipids and lipoproteins. When
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33 a porous substrate bearing latent fingerprints is immersed in the ORO working solution, the
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35 fingerprints appear as visible red marks by accumulation of the dye molecules, whereas the
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37 substrate generally undergoes a slight coloration (light red) leading to sufficient contrast
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39 between the mark and the substrate to permit good visualization (Fig. 1). ORO also is used in
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41 forensic science to stain lip marks, especially when lipstick is used (Navarro et al. 2006).
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46 Solvent black 3 (or Sudan black) is another lysochrome used to stain lipids and
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48 phospholipids in histochemistry. Very few applications of this dye for fingerprint detection
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50 have been reported (Hart 2003, Home Office Scientific Development Branch 2005). Solvent
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52 black 3 is applied mostly on greasy, contaminated surfaces for which other methods give poor
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54 results or fail to detect fingerprints. The marks that are obtained after staining are dark
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56 blue.
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Enhancement of cyanoacrylate-developed fingermarks

Cyanoacrylate fuming cannot really be considered a physical staining method as defined in the introduction, because chemical processes take place with the secretion residue. This detection technique, however, generally is combined with a subsequent dyeing step, which does fall into this category.

Cyanoacrylate fuming, a method used worldwide, was developed toward the end of the 1970's (Lee and Gaensslen 1984) to detect fingermarks on nonporous surfaces. It involves the use of a colorless monomeric liquid, i.e., the cyanoacrylate ester (also known as Superglue, a rapid bonding, high strength glue) heated to its boiling point. Monomeric vapors are produced that react with some components of fingermarks by polymerization (Fig. 2), but not with the underlying substrate. The exact initiators of the polymerization are not yet clearly determined, because cyanoacrylate shows particular affinity for moisture, lipids and some water soluble components (Lewis et al. 2001, Wargacki et al. 2007). As a result of the polymerization process, which continues until the source of monomers is removed or the heating process is stopped, a white solid covers the developed fingermarks (Mankidy et al. 2006). At the microscopic level, polycyanoacrylate is not a massive and continuous film that covers the secretion residue, but rather can be described as tangled noodles. Owing to the toxicity of the vapors (Hughes 1993), treatment is performed in a cabinet where humidity and heating of the cyanoacrylate can be controlled precisely.

Cyanoacrylate fuming is a sensitive technique for detecting fingermarks on nonporous surfaces. The contrast obtained, however, sometimes is poor due to the white color of the polymer (Fig. 3), which results in barely visible or undetected fingermarks. Fortunately, cyanoacrylate-developed fingermarks can be stained using organic dyes that are trapped selectively in the polymer network and this significantly enhances the visual contrast (Fig. 3)

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3 (Champod et al. 2004). Dyes, typically luminescent ones, that are used routinely in forensic
4 laboratories around the world (Fig. 4) include Ardrex, basic red 28, basic yellow 40, Nile red
5 and rhodamine 6G (McCarthy 1990, Olenik 1992, Day and Bowker 1996). The choice of dye
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(Champod et al. 2004). Dyes, typically luminescent ones, that are used routinely in forensic laboratories around the world (Fig. 4) include Ardrex, basic red 28, basic yellow 40, Nile red and rhodamine 6G (McCarthy 1990, Olenik 1992, Day and Bowker 1996). The choice of dye relies mostly on the properties of the substrate, e.g., color, background luminescence, or solvent interactions that govern the specific excitation and emission wavelength domains (Mazzella and Lennard 1995). Solutions of mixed dyes (Olenik 1997), or even rare earth compounds, such as Europium complex, also can be used to widen the wavelength domains (Lock et al. 1995).

Chemical detection techniques

By chemical detection, we mean generally uncolored staining reagents that undergo chemical reaction, i.e., modifications of their molecular structure, with some components of the fingerprints to produce colored or luminescent compounds.

Silver nitrate

A reagent that must be cited for historical reasons is silver nitrate (AgNO_3). This was first used in 1878 to detect chloride ions contained in papillary secretions (Aubert 1877-1878, Olsen 1978). This method can be applied on porous surfaces including paper and untreated wood. The principle is simple: silver nitrate reacts with the chloride ions contained in the secretion residue to form silver chloride (AgCl). Upon exposure to light, AgCl decomposes to form metallic silver (Ag), resulting in a black fingerprint. Owing to the diffusion of chloride ions through the substrate, however, blurred fingerprints are obtained if they are aged more than one week. The kinetics of the diffusion has been studied to try to determine the age of the fingerprint (Angst 1962).

Amino acid reagents

Amino acids constitute a major organic component of sweat; 0.3–2.59 mg of amino acids are secreted per liter of sweat (Ramotowski 2001), which corresponds to 2.4–20.7 μM for a typical amino acid. Amino acids, therefore, represent a specific target for a fingerprint detection strategy based on chemical reaction. Amino acid detection (or quantification) agents used in biology, such as ninhydrin, have been applied successfully in forensic science as described below.

Amino acids generally, unlike chloride ions, show strong affinity for paper fibers, and their migration through such substrates is prevented. Fingermarks thus are well preserved in the paper matrix and can be detected satisfactorily tens of years after deposition. Note also that amino acids typically are water soluble constituents of sweat. This means that if the item has been wet, such compounds will not be recovered and the following techniques could not be successfully applied.

Ninhydrin (2,2-dihydroxy-1,3-indanedione) was the first amino acid reagent used for detecting latent fingerprints on porous surfaces such as paper and cardboard (Odén and Von Hofsten 1954) and it still is widely used. Ninhydrin can detect primary and secondary amines contained in amino acids or in polypeptides present in latent fingerprints. When reacting in situ with such free amines, ninhydrin produces a deep blue or purple product, Ruhemann's purple (Fig. 5). As a result, fingerprints appear as purple ridges on unmodified substrate (Fig. 6). Several studies have been performed to improve the sensitivity and to optimize the development conditions over wider temperature and relative humidity ranges (Lamothe and McCormick 1972, Grigg et al. 1986, Lennard et al. 1986, Almog 2001, Petraco et al. 2006).

Despite the great sensitivity of ninhydrin, the contrast between the fingerprint and the substrate can be weak, e.g., on dark backgrounds. In this case, observation of the fingerprints is impossible, because Ruhemann's purple is not luminescent. Efforts have been made to

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3 overcome this problem, especially by using post-ninhydrin treatments containing metal salts.
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5 When Ruhemann's purple is reacted with zinc or cadmium ions, luminescent coordination
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7 complexes are formed. As a result, fingerprints undergo a color change and become orange
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9 instead of purple with zinc and red instead of purple with cadmium. The derivatives also
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11 exhibit luminescence when cooled to 77 K with liquid nitrogen (Kobus et al. 1983, Stoilovic
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13 et al. 1986, Lennard et al. 1987). Cadmium toxicity and the inconvenience of storing liquid
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15 nitrogen, however, are drawbacks limiting application of such post-treatments. Extensive
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17 research has been carried out to discover ninhydrin-like reagents with increased sensitivity
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19 and room temperature luminescence (Almog et al. 1982, 1992, 2000, 2001, Lennard et al.
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21 1986, 1988, Joullié et al. 1991, Hansen and Joullié 2005). Many ninhydrin analogues have
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23 been synthesized and studied, but despite this, few have been used routinely for reasons of
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25 expense and time-consuming synthesis protocols.
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32 Another amino acid reagent that has been used widely for nearly 20 years is 1,8-
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34 diazafluoren-9-one (DFO) (Wilkinson 2000b). Unlike ninhydrin, DFO produces very faint
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36 pink fingerprints, barely visible to the naked eye, but which are extremely luminescent at
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38 room temperature (Grigg et al. 1990, McComiskey 1990, Pounds et al. 1990).
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42 More recently, a third amino acid reagent, 1,2-indanedione, has been shown to be
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44 effective for detecting fingerprints on porous surfaces. When using 1,2-indanedione, pink
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46 colored fingerprints characterized by strong luminescence at room temperature are obtained
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48 (Fig. 7). Extensive comparative studies showed that 1,2-indanedione is an alternative to DFO
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50 (Joullié et al. 1998, Roux et al. 2000, Wilkinson 2000a, Wiesner et al. 2001, Kasper et al.
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52 2002, Gardner and Hewlett 2003). Recent research also has shown that when zinc chloride
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54 (ZnCl_2) is introduced to the 1,2-indanedione working solution before treating the specimen,
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56 the luminescence of the detected fingerprints is markedly improved (Stoilovic et al. 2007,
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58 Wallace-Kunkel et al. 2007, Bicknell and Ramotowski 2008, Russell et al. 2008). For these
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3 reasons and others, such as faster detection procedure, DFO is being replaced by 1,2-
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5 indanedione in the forensic laboratory for detecting fingermarks on porous surfaces.
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8 Concerning the mechanisms of the reactions between amino acids and the reagents
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10 above, a recent study showed that the reactions between DFO or ninhydrin and all the amino
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12 acids go to completion (Spindler et al. 2009). This is not the case with 1,2-indanedione,
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14 however, which varies according to the amino acid involved. The addition of a small amount
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16 of zinc to the working solution seems to play a role in the completeness of the reaction,
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18 because zinc acts as a catalyst.
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22 Two other less widely used reagents for amino acids also are used. Genipin is a natural
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24 product extracted from *Gardenia jasminoides*. Upon reacting with amino acids, a blue
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26 reaction product is obtained that gives a red luminescence at room temperature (Lee et al.
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28 2003, Almog et al. 2004, Levinton-Shamuilov et al. 2005). This reagent constitutes the only
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30 "dual fingermark reagent" for amino acids, i.e., a reagent giving visible/colored fingermarks
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32 that also can be observed with luminescence. Genipin has the further advantage of being a
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34 natural and nontoxic product. Lawsone (2-hydroxy-1,4-naphthoquinone) is the compound
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36 thought to be responsible for the staining properties of henna, commonly used as a skin and
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38 hair dye. In forensic science, naphthoquinones represent a new group of compounds that can
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40 be used to detect the amino acids (Jelly et al. 2008). After treatment with lawsone,
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42 fingermarks are stained purple-brown and exhibit a strong red luminescence. Currently, only
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44 experimental results are available and further research is needed before considering it an
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46 effective reagent.
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55 ***p*-4-Dimethylaminocinnamaldehyde (pDMAC)**

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57 Urea is another component of sweat that could be targeted to detect fingermarks on porous
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59 surfaces. The first reagent proposed for this purpose was *p*-4-dimethylaminocinnamaldehyde
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3 (pDMAC). The resulting fingermarks are red and luminescent (Sasson and Almog 1978,
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5 Brennan et al. 1995, Ramotowski 1996). The sensitivity of pDMAC, however, has been
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7 shown to be less than the conventional amino acid reagents described above. Moreover,
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9 blurred fingermarks generally are obtained if they are more than two days old. Finally, the
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11 actual molecular target of pDMAC has been questioned (Home Office Scientific
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13 Development Branch 2006) and it appears that pDMAC actually could be reacting with amino
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15 acids rather than with urea. pDMAC is not used routinely.
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22 *Osmium tetroxide and ruthenium tetroxide*

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24 Osmium tetroxide (OsO_4) and ruthenium tetroxide (RuO_4 , RTX) are compounds that can react
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26 with the double bonds of the unsaturated organic compounds found in a fingermark deposit
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28 through an oxidation mechanism (Grzegorzewska and Filbrandt 2004). Direct exposure to
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30 OsO_4 vapors provides black fingermarks after 1 to 12 h (Olsen 1978), whereas RTX gives
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32 gray marks after only 10 to 20 min (Mashiko et al. 1991, Mashiko and Miyamoto 1998).
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34 Despite the excellent results that can be obtained on a variety of surfaces, especially the
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36 problematic ones like human skin, RTX and OsO_4 are not used routinely because of their high
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38 toxicity through inhalation or skin contact and hazards associated with their manipulation
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40 (Blackledge 1998). Only fully equipped laboratories can consider their use.
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48 *Fingermarks in blood*

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50 When the victim and/or the offender are injured, hands can become contaminated with blood.
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52 In such cases, bloody fingermarks are likely to be recovered. Depending on the amount of
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54 blood and on the nature of the substrate, some marks are obvious to the naked eye, but others
55
56 remain latent and require the application of specific blood reagents.
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1
2
3 Although two different categories of blood reagents exist, *viz.*, protein stains and heme
4 reagents, blood sometimes can be visualized directly due to a strong absorption at 415 nm
5
6 (Stoilovic 1991) that makes it appear black under illumination at this wavelength. Blood also
7
8 can be observed due to luminescence in the long-wave UV region when short-wave UV
9
10 exciting light is used (Springer et al. 1994). Before using any blood-specific reagent, the items
11
12 bearing latent fingerprints generally are dipped in methanol or in an aqueous 5-sulfosalicylic
13
14 acid solution (2% w/v) to avoid loss or diffusion of blood due to the water-based blood
15
16 reagents (Hussain and Pounds 1988, Sears and Prizeman 2000). Once fixed, effective blood
17
18 reagents can be applied.
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24 Protein stains are compounds adopted directly from biology. The application protocol
25
26 consists of immersing the item to be treated in a dye solution. The staining times vary from 10
27
28 sec to several minutes depending on the dye, the type of substrate and the amount of blood.
29
30 The dye binds to the proteins, which leads to a colored or luminescent complex. The item then
31
32 is washed with successive de-staining solutions, based on methanol or water, to remove the
33
34 excess dye and to clear the background so that contrast is improved (Fig. 8). Because
35
36 numerous protein stains can be applied, only the commonly used ones are noted here (Sears
37
38 and Prizeman 2000, Sears et al. 2001, 2005, Marchant and Tague 2007). A conventional non-
39
40 luminescent stain is amido black (naphthol blue black B), which gives dark blue marks on
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42 light blue or nearly colorless substrates. Luminescent stains also are available, e.g., acid
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44 yellow 7, acid fuchsine (Hungarian red) and acid violet 17 (Coomassie brilliant violet). Which
45
46 dye is used depends on the color and the nature of the substrate.
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52 Heme reagents are molecules that can undergo oxidation, initiated by hydrogen
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54 peroxide in presence of a catalyst, in this case the heme group of hemoglobin. As a result,
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56 colored, luminescent, or chemiluminescent (a product generating its own light with no need to
57
58 use excitation light source) products are formed. The reaction is selective for blood (possible
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1
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3 false positive results are not discussed here) and gives little background staining. Routinely
4
5 used stains include 3,3-diaminobenzidine (DAB) (Allman and Pounds 1991, 1992, Sahs
6
7 1992), which gives a dark brown color; ABTS (2,2'-azino-di-(3-ethyl-
8
9 benzthiazolinesulfonate) diammonium salt) (Caldwell et al. 2000), which gives a green
10
11 reaction product; and leucomalachite green, phenolphthalein, leucocrystal violet, o-tolidine
12
13 (Lee 1984), fluorescein (Cheeseman and DiMeo 1995), leuco rhodamine 6G (Yapping and
14
15 Yue 2004) or eosin Y (Wang et al. 2007). Another heme reagent that must be cited, even if it
16
17 is not used as a fingerprint reagent, is luminol. This gives rise to intense blue
18
19 chemiluminescence and is used to detect traces of blood as might be found on a crime scene
20
21 or on items that have been washed (Webb et al. 2006). Any fingerprints that are obtained,
22
23 however, are not sufficiently sharp to detect fingerprint ridges that are necessary for
24
25 identification of the person, and the procedure is used only to detect the presence of blood,
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27 which facilitates subsequent blood sampling for DNA analysis.
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34 Finally, note that the conventional fingerprint reagents, e.g., amino acid reagents or
35
36 cyanoacrylate, also can detect blood fingerprints even if they do not possess the sensitivity of
37
38 the specific blood reagents. Conventional stains, therefore, generally are applied before the
39
40 blood reagents to detect most of the existing bloody or non-bloody fingerprints.
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46 **Nanoparticles – a new generation of stains**

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48 During the last decade, considerable interest has emerged within the forensic science
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50 community in using nanoparticles or nanocomposites for detecting fingerprints.
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53 Nanoparticles already are applied successfully in cell and whole animal imaging (Bagwe et al.
54
55 2003, Medintz et al. 2005, Michalet et al. 2005, Murcia and Naumann 2005, Thurn et al.
56
57 2007), but constitute an emerging field in forensic science.
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1
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3 Interest in nanoparticles to detect fingerprints is due to their properties: 1) their size is
4
5 1,000 to 10,000 times smaller than a fingerprint ridge width, which ensures excellent
6
7 resolution upon detection, 2) their structure can be modified easily by addition of molecular
8
9 chains or chemical functionalities on their outer surface, and 3) their optical properties make
10
11 them excellent candidates for obtaining luminescent fingerprints. Nanoparticles such as
12
13 quantum dots are intrinsically luminescent (Bawendi et al. 1990, Wang and Herron 1991,
14
15 Murray et al. 1993), and others, e.g., silica nanoparticles, are made luminescent by
16
17 incorporating organic dyes in their structure during their synthesis.
18
19
20
21

22 The challenge for using nanoparticles to detect fingerprints lies in the fact that
23
24 nanoparticles *per se* generally have no specific affinity for the marks. It is necessary,
25
26 therefore, to find a way to make nanoparticles specifically target the secretion residues
27
28 without acquiring an affinity for the substrate; otherwise the resulting contrast would be poor
29
30 (Fig. 9). A successful application of nanoparticles generally is obtained by adding functional
31
32 groups onto the nanoparticles (Fig. 10) or by modifying the experimental conditions, such as
33
34 the pH, to influence parameters such as the charge on the nanoparticle.
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39 The following sections describe four approaches that were selected for their specificity
40
41 in terms of fingerprint detection processes: physico-chemical detection, lipid-lipid interaction,
42
43 and electrostatic interaction.
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48 ***Metal nanoparticles***

49
50 Use of metal nanoparticles to detect fingerprints on various kinds of substrates has interested
51
52 forensic scientists for several years (Choi et al. 2008b). Both in terms of research and practical
53
54 applications, most interest has focused on two kinds of metal nanoparticles, gold and silver.
55
56 The multimetal deposition technique (based on gold nanoparticles) and physical development
57
58 (based on silver nanoparticles) are not new; both were developed in the 1980s. Research
59
60

1
2
3 continues to clarify understanding of the underlying physico-chemical processes that permit
4
5 the detection of fingerprints, however, with the aim of enhancing and simplifying existing
6
7 protocols. Both approaches are described below, followed by a section dealing with additional
8
9 possibilities offered by metal nanoparticles.
10
11

12 13 14 15 *Multimetal deposition (MMD)*

16
17 Gold nanoparticles conventionally are synthesized in water. A gold (III) derivative (generally,
18
19 tetrachloroauric acid; HAuCl_4) is reduced using an agent such as sodium citrate or sodium
20
21 borohydride. The reduction of HAuCl_4 by sodium citrate in boiling water is one of the most
22
23 commonly used procedures for obtaining monodisperse spherical gold nanoparticles,
24
25 generally about 10–20 nm diameter, with time stabilities of several months to years
26
27 (Turkevich et al. 1951, Frens 1973, Slot and Geuze 1981, Turkevich 1985a,b). The origin of
28
29 such excellent stability is to be found in the capping of the gold nanoparticles in solution by
30
31 tri-negatively charged citrate ions. The electrostatic repulsion of the resulting negatively
32
33 charged nanoparticles prevents them from aggregating. The same negative charge also is the
34
35 basis of their ability to detect fingerprints by exploiting charges on organic components such
36
37 as proteins, amino acids and lipids as latent prints deposited on a substrate.
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44 If a sample bearing a fingerprint is immersed in an aqueous solution of freshly
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46 prepared colloidal gold (pH ~ 6.2), nothing happens. If the pH of the colloidal solution is
47
48 lowered beforehand, however, it is possible to find a small range of pH for which both of the
49
50 following conditions are met. 1) The tri-negatively charged citrate ions are not yet fully
51
52 protonated ($\text{pK}_{a1} = 3.13$, $\text{pK}_{a2} = 4.76$, $\text{pK}_{a3} = 6.40$) and still give the gold nanoparticles a
53
54 negative charge. It should be noted that the interaction of carboxylic groups with a metal
55
56 surface, as is the case with sodium citrate and gold, can lead to a decrease of the pKa values
57
58 (increasing acid strength of the groups) measured for the same molecule in solution
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1
2
3 (Królikowska and Bukowska 2007). 2) The proteins and amino acids that are entrapped in the
4
5 residue become progressively more positively charged due to the protonated amino groups (-
6
7 NH_3^+). These positive charges are progressively less counterbalanced by the carboxylate
8
9 groups ($-\text{COO}^-$) as they become protonated to neutral carboxylic moieties ($-\text{COOH}$) at low
10
11 pH.
12
13

14
15 In the case of fingerprints, the range of pH is extremely narrow and has been
16
17 determined experimentally as 2.5-2.8 (Schnetz and Margot 2001). When the pH of colloidal
18
19 gold is within this range and a sample bearing a fingerprint is dipped in it, negatively charged
20
21 gold nanoparticles in solution become attracted electrostatically by the positively charged
22
23 residue. Above this range, proteins are not sufficiently positively charged; below this range,
24
25 gold nanoparticles become uncharged by complete protonation of the sodium citrate ions (the
26
27 solution turns dark purple as the gold nanoparticles aggregate owing to lack of electrostatic
28
29 repulsion). This mechanism of staining selectivity is intriguingly like that of acid dyeing of
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31 proteins in biological staining (Horobin 1988) and textile dyeing (Zollinger 2003).
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37 These physico-chemical considerations are the basis of the MMD technique, (Fig. 11).
38
39 MMD was developed in 1989 (Saunders 1989) and improved in 2001 (Schnetz and Margot
40
41 2001). This technique is based on the electrostatic attraction between the gold nanoparticles
42
43 and secretions when the pH of the colloidal gold is adjusted to 2.65. Fingerprints detected by
44
45 gold nanoparticles only, however, are not readily visible to the naked eye, because they
46
47 produce very faint pink marks due to the small size of the nanoparticles. To increase the
48
49 contrast, a second metal deposition is performed by reducing silver ions to metallic silver on
50
51 the surface of the gold nanoparticles, which play the role of catalytic sites. As a result,
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53 fingerprints appear dark brown as illustrated in Fig. 12. MMD is an efficient technique for a
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55 wide range of substrates, e.g., porous, nonporous and wet, but it is highly labor intensive and
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57 produces dark brown marks only.
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3 An alternative to MMD was proposed in 2007, the single metal deposition (SMD)
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5 (Stauffer et al. 2007, Durussel et al. 2009). The procedure is similar to MMD, except that the
6
7 enhancement step, i.e., second metal deposition, is performed by reducing gold ions instead of
8
9 silver onto the gold nanoparticles. The results obtained with SMD are similar to MMD (Fig.
10
11 12), but the overall procedure is cheaper, less labor intensive, and easier to set up.
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15 Both MMD and SMD produce dark brown fingerprints. This means that the contrast
16
17 can be very good on transparent, white or light colored substrates, but these techniques fail to
18
19 produce sufficient contrast on dark colored or patterned substrates for which purpose
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21 luminescence is required. A modification of the enhancement step led to the formation of a
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23 zinc oxide shell instead of silver or gold around the gold nanoparticles after they were
24
25 deposited on the secretion residue (Becue et al. 2008). In this way, luminescent fingerprints
26
27 were obtained on dark, nonporous substrates (Fig. 12). This technique currently is limited to
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29 nonporous surfaces, however, unlike MMD and SMD, which can be applied on porous and
30
31 nonporous substrates.
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36 37 38 *Physical developer*

39
40 Use of a physical developer (PD) is another detection technique based on metal nanoparticles
41
42 and driven by physico-chemical mechanisms (Goode and Morris 1983). Unlike MMD, for
43
44 which metal colloids were synthesized prior to the development procedure, PD consists of
45
46 immersing samples bearing fingerprints in an acidic aqueous solution containing silver ions
47
48 e.g., silver nitrate; reducing agents, e.g., ferrous and ferric ions; positively charged ionic
49
50 surfactant, e.g., n-dodecylamine acetate; and other chemical components such as citric acid to
51
52 lower the pH of the solution. The accepted mechanism is that during the reduction process,
53
54 negatively charged silver colloids are formed by reduction of the silver ions (Cantu 2001,
55
56 Cantu and Johnson 2001). If such silver nanoparticles are formed in solution, they are
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3 surrounded rapidly by positively charged surfactant molecules, which prevent silver colloids
4 from aggregating and stops particle growth (Fig. 13). Such nonreactive nanoparticles do not
5 play a role in fingerprint detection and remain in solution. If the negatively charged silver
6 colloids are formed near a fingerprint, however, they immediately become attracted by the
7 positively charged residue because the pH of the solution is lowered to the acidic range. Once
8 entrapped in the secretion, these silver colloids act as reduction sites for the additional silver
9 ions. As a result, fingerprints become progressively visible as the reduction process
10 continues. At some point, the forensic scientist stops the process by removing the sample
11 from the reducing bath and by dipping it in a neutralizing/rinsing water bath. The fingerprints
12 obtained are a gray color (Fig. 13). Like MMD, PD is applied to white colored substrates
13 only.
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32 *Other applications of metal nanoparticles*

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34 In the cases of MMD, SMD and PD, metal nanoparticles are used without further surface
35 modifications; the detection is driven by physico-chemical mechanisms and especially
36 electrostatic interactions. Another strategy involves functionalizing the metal nanoparticles
37 with nonpolar hydrophobic aliphatic chains (Fig. 14) so that they can be applied in suspension
38 in organic solvents (Sametband et al. 2007) or as enhanced nanopowders once dried and
39 dusted using a brush (Choi et al. 2006, 2008a). The addition of aliphatic chains to the
40 nanoparticles enhances the interactions between the metal nanoparticles and the lipids
41 contained in the fingerprint residue (Choi et al. 2008b). Compared to classical (micro-sized)
42 powders, nanopowders have been shown to give better results in terms of ridge details and
43 less unwanted deposition on the underlying substrate. Despite this, the use of nanopowders
44 did not necessarily lead to a better contrast, because the detected fingerprints were a lighter
45 color than the dark colored classical powders.
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Another research strategy consisted of binding antibodies onto gold nanoparticles to take advantage of the highly selective antibody-antigen recognition process to detect specific drug metabolites in a fingerprint (Leggett et al. 2007). In an attempt to provide evidence of the consumption of a drug in addition to touching contaminated objects, anti-cotinine antibodies bound to gold nanoparticles have been used to target cotinine, a metabolite of nicotine, present in the sweat of tobacco smokers. Combined with a fluorescent marker, highly detailed fingerprints can be observed on glass slides (Fig. 15). Another research strategy is immunolabeling to identify antigenic sites with the secretion residue components or keratinized skin. Proteins present in sweat or skin remnants that have been identified in this way are keratins 1 and 10, cathepsin D and dermcidin using the corresponding antibodies (Drapel et al. 2009).

Metal Oxide nanoparticles

Nanopowder and small particle reagents

Metal oxides have been widely used for detecting fingerprints due to their optical properties, i.e., color and fluorescence, because it is possible to obtain both white particles, such as titanium dioxide (TiO_2) or zinc oxide (ZnO), and also black particles, such as iron oxide (Fe_2O_3). Such reagents can be used as conventional fingerprint powders or they can be suspended in aqueous media using a detergent to enhance their solubility. As a result, one can detect fingerprints on nonporous substrates or on the adhesive side of tapes using a process that has been described as a "wet version of the classical powdering process" or "wet powdering" (Cucè et al. 2004, Williams and Elliot 2005). Instead of using a brush, samples are treated by immersing them in particle suspensions before rinsing with water. Fingerprints are detected by entrapment of the metal oxide particles in the lipid matrix of the secretion residue. This technique uses "small particle reagents" (SPR) and gives good results with

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3 nonporous substrates on which latent fingerprints are present, especially if the substrate has
4 been wet (Goode and Morris 1983, Frank and Almog 1993, Champod et al. 2004). In addition
5
6 to the metal oxide particles noted above, black molybdenum sulfide (MoS_2) and white zinc
7
8 carbonate (ZnCO_3) particles also are used as SPR agents. According to the color of the
9
10 underlying substrate, white powder, e.g., TiO_2 , or black powder, e.g., Fe_2O_3 , is used to
11
12 maximize the contrast. A luminescent version of SPR has been proposed that involves
13
14 addition of luminescent organic dyes, e.g., basic yellow 40 in ethanol, to a conventional
15
16 aqueous SPR solution (Springer and Bergman 1995, Lee and Gaensslen 2001, Jasuja et al.
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18 2008).

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The size of the particles influences the result of SPR; smaller particles give better results than larger ones (Frank and Almog 1993). As for dried nanopowders, the use of nanoparticles in the context of SPR can greatly improve the resolution.

More recently, nanostructured ZnO was used to detect fingerprints on nonporous surfaces by taking advantage of its visible fluorescence when dry dusted or applied as an SPR (Choi et al. 2008a). These investigators combined oleylamine with the fluorescent dye, perylene dianhydride, to form an entity that then was adsorbed onto TiO_2 nanoparticles to form a powder exhibiting strong fluorescence at 650-700 nm when excited at 505 nm (Choi et al. 2007). When dusted onto nonporous substrates bearing latent fingerprints, this fluorescent nanopowder was slightly weaker in fluorescence intensity compared to conventional luminescent powders, but produced significantly less background. As a result, good contrast was obtained between the fingerprints and the substrate.

Titanium dioxide as a blood reagent

1
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3 Sub-micron titanium dioxide particles suspended in anhydrous methanol can be used to detect
4
5 blood fingermarks on nonporous and "semiporous" surfaces (Bergeron 2003). The results
6
7 were excellent on nonporous surfaces. No mechanistic explanation was offered for the affinity
8
9 of TiO₂ for blood. Uptake, however, may be related to the fact that TiO₂ has some affinity for
10
11 blood (Nygren et al. 1997) and for proteins by electrostatic interactions (Topoglidis et al.
12
13 2001).
14
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16 17 18 19 20 **Quantum dots (QDs)**

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22 QDs are crystalline semiconductor nanoparticles containing a few hundred to several
23
24 thousand atoms varying in size from 1 to 10 nm. Owing to quantum confinement, QDs exhibit
25
26 strong luminescence at room temperature. It is of interest that the emission wavelength is
27
28 dependent on the size of the nanoparticle; larger nanoparticles emit in the infrared/red range
29
30 and smaller ones emit in the ultraviolet/blue range of the spectrum. Another significant
31
32 feature of such nanoparticles is the fact that all the QDs have a broad excitation spectrum in
33
34 the ultraviolet range. This means that a variety of QDs, each emitting its own wavelength, can
35
36 be excited by a single excitation source. These nanoparticles are used in a wide variety of
37
38 applications such as components for lasers (Bukowski and Simmons 2002) or as fluorescent
39
40 probes for cellular imaging (Medintz et al. 2005). QDs high quantum yield and ability to resist
41
42 photobleaching make them superior markers compared to organic fluorescent dyes (Resch-
43
44 Genger et al. 2008). QDs can be synthesized in either water or organic solvents depending on
45
46 their external surface coating. Moreover, their outer surface can be functionalized easily with
47
48 chemical groups to provide new functionalities. Consequently, QDs are widely used in cell
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50 biology to stain different components of cells or tissues (Bruchez et al. 1998).
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58 In forensic sciences, particularly for fingerprint detection, the properties of QDs are
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60 stimulating great interest. Their surfaces can be coated specifically to target the secretion

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3 residue and the luminescence emission color can be tuned in case of a luminescent
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5 background. Since 2000, several studies have focused on the use of QDs to detect
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fingermarks. Cadmium sulfide (CdS) nanocrystals, used alone or combined with dendrimers, i.e., hyperbranched polymers, have been proposed as stains for cyanoacrylate-developed fingermarks (Bouldin et al. 2000, Menzel et al. 2000a,b). The results, however, were disappointing, because the staining times required were too long (24 h) and the working solutions were too unstable. More recent studies have provided only limited improvements (Yu-Juan et al. 2008). The combination of QDs and dendrimers does not seem to constitute an alternative to the existing cyanoacrylate stains.

In another recent study, researchers stabilized QDs in petroleum ether by grafting aliphatic chains on their outer surface and attempted to detect sebaceous fingermarks with these nanomaterials. Promising results were obtained on silicon wafers, but on porous surfaces, the results were limited due to high background luminescence (Sametband et al. 2007).

Cadmium sulfide (CdS) encapsulated in a biopolymeric chitosan matrix has been used as dusting powder for detecting fresh marks on aluminium foil (Dilag et al. 2009). Luminescent fingermarks were obtained, but the process involved in the detection is only physical. Moreover, the use of dried cadmium-based nanoparticles as dusting powder has serious health and safety implications. Water soluble cadmium selenide (CdSe) QDs have been synthesized and used to detect fresh marks on the adhesive surface of tape with promising results (Wang et al. 2009). Cadmium telluride (CdTe) QDs, synthesized in aqueous solution, were used to detect blood fingermarks on various nonporous surfaces (Becue et al. 2009) such as aluminium foil, black polyethylene, glass or transparent polypropylene (Fig. 16). A comparison with acid yellow 7, one of the best blood reagents for nonporous substrates

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2
3 (Sears et al. 2001), showed that the QDs were superior to acid yellow 7 on aluminium and
4
5
6 equally effective for the other substrates.

7
8 Currently, the use of QDs to detect fingerprints remains a research proposition and no
9
10 casework uses of QDs have been reported. QDs show great promise, however, for efficient
11
12 fingerprint detection. Issues remaining to be addressed before routine application is likely
13
14 include the toxicity of cadmium and improved targeting of secretion residue by
15
16 functionalization of the QD outer surface.
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21 22 *Silica nanoparticles*

23
24 Synthesis of silica nanoparticles is simple and consists of a succession of
25
26 hydrolysis/condensation processes on silicon alkoxide precursors ($\text{Si}(\text{O}-\text{R})_4$; R = an alkyl
27
28 chain). This leads to the formation of silicon oxide nanoparticles in suspension in water
29
30 (Stöber and Fink 1968) or in a water-in-oil emulsion (Arriagada and Osseo-Asare 1999,
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32 Bagwe et al. 2004). The introduction of organic dyes during the synthesis permits their
33
34 attachment to the nanoparticle surface or to their embedment within the particles (van
35
36 Blaaderen and Vrij 1992, Santra et al. 2001, Tapeç et al. 2002, Zhao et al. 2004, Johnston et
37
38 al. 2005). Such dye-doped nanoparticles currently are widely used as labeling agents for bio-
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40 imaging purposes. They are highly efficient alternatives to classical organic fluorophores,
41
42 because the silica coating (or encapsulation) of some dyes isolates them from the oxygen and
43
44 water of the outside environment. This both increases photostability and emission quantum
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46 yield of the dye and decreases photobleaching by preventing penetration of oxygen, which
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48 promotes photodecomposition (Tan et al. 2004).
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55 In the context of fingerprint detection, the use of luminescent silica nanoparticles for
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57 forensic purposes remains at the pilot study stage. Theaker et al. (2008) recently reported
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59 entrapment of a variety of colored and fluorescent dyes including basic red 28, basic yellow
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3 40, fluorescein, methylene blue, oxazine perchlorate, rhodamine B, rhodamine 6G and
4
5 thiazole orange within silica particles. The resulting doped nanoparticles were used in
6
7 aqueous suspensions to detect fingerprints. The process is very similar to that described for
8
9 small particle reagents. Micron-size particles also were used as dusting agents. Both fresh (20
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11 min) and aged fingermarks (40 days old) showed good definition after development.
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15 Current research projects in our laboratory aim to use dye-doped silica nanoparticles
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17 that are functionalized chemically to recognize the fingerprint residue. Such dye-doped
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19 nanoparticles can be considered inert labeling agents that require appropriate functionalization
20
21 to promote (bio)molecular interactions or reactions with components of interest. One
22
23 advantage of the silica nanoparticles is the versatility provided by their surfaces for chemical
24
25 modification. This is because the surfaces of silica nanoparticles carry silanol groups (Si-OH).
26
27 These moieties readily undergo chemical modifications, such as the addition of an additional
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29 siloxane layer using an organosilane or functionalized alkoxy silane, e.g., X-(CH₂)_m-Si-(OR)₃;
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31 where X is the functionality to be added. Other chemical modifications include addition of
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33 alkyl chains bearing amine, carboxyl, or thiol functionalities (Fig. 17). The incorporation of
34
35 chemical groups onto the silica surface allows tailoring of the chemical behavior of the
36
37 surface, and thus the nanoparticle as a whole, compared to inert "naked" particles. The
38
39 challenge in forensic science is to find the best functionalities to add to dye-doped silica
40
41 nanoparticles to enhance targeting of latent fingermarks on a wide range of substrates.
42
43 Preliminary results are encouraging and suggest the considerable potential of such a strategy
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45 for detecting fingermarks by combining the intense luminescence of the dye-doped silica
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47 nanoparticles and the selectivity of the grafted ligands.
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55 In forensic science, and more particularly in the context of fingerprint detection, stains
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57 play a key role. To cover the subject as completely as possible, we chose to define a "stain" as
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59 every chemical entity (organic molecule, metal nanoparticle, dye-doped nano- and micro-
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3 structured element, or other compound) that can create a contrast between a latent fingerprint
4 and its underlying substrate. This wide definition goes beyond the conventional organic dyes
5 or organic dye-doped nanoparticles that currently are used for bio-imaging applications,
6 because it also encompasses entities such as iodine, metal salts, or metal nano- or micro-
7 particles, which are excellent contrast elements.
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15 With this review, we aimed at presenting various aspects of fingerprint detection
16 using dyes. The possibilities offered to forensic scientists are numerous and include
17 conventional staining of the lipid fraction of fingerprints, chemical targeting of amino acids
18 or proteins, selective polymerization on the secretion residue, or the use of more complex
19 physico-chemical processes. The choice for one technique over another, or for a combination
20 of different ones, is determined mainly by the nature of the item or by the surface at the crime
21 scene, e.g., walls or furniture, and by the context of the criminal affair, e.g., timeline, bloody
22 case, evidence exposed to environmental elements, prolonged exposure to water. In addition,
23 the choice for a luminescent dye or a conventional stain is strongly dependant on the nature
24 and color of the substrate, even if it is generally recognized that luminescent techniques are
25 more sensitive compared to nonluminescent ones.
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41 During the last decade, several investigations have focused on the development of new
42 detection techniques based on the use of functionalized and/or luminescent nanoparticles for
43 detecting fingerprints. Nanoparticles have great promise for fingerprint detection due to their
44 high surface-to-volume ratio, their size-dependent qualities, their optical properties and the
45 fact that they can easily be tuned chemically. If surface modification and composition control
46 are combined, these properties impart high selectivity and sensitivity. The results currently are
47 promising and could provide forensic scientists with new and efficient alternatives to the
48 currently existing techniques.
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3 **Fig. 1.** Left: molecular structure of oil red O. Right: representative fingerprint detected using
4 this technique with paper as substrate.
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10 **Fig. 2.** Polymerization mechanism on the secretion residue during the cyanoacrylate fuming
11 process.
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17 **Fig. 3.** Left: cyanoacrylate-fumed fingerprint located on a beer glass. Note that the quality of
18 development is not optimal. Right: same fingerprint after enhancement using rhodamine 6G
19 as a cyanoacrylate dye and observed using luminescence.
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27 **Fig. 4.** Chart illustrating the excitation and emission wavelength ranges for the most
28 commonly used cyanoacrylate dyes, the colors of which are indicated on the diagram.
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30 Abbreviations: BY40, basic yellow 40; BR28 basic red 28; S7 Styryl 7.
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36 **Fig. 5.** Reaction mechanism between ninhydrin and amino acid leading to the formation of a
37 dark purple product, Ruhemann's purple.
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43 **Fig. 6.** Left: molecular structure of ninhydrin (2,2-dihydroxy-1,3-indanedione). Right:
44 representative fingerprint detected using this technique with paper as substrate.
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50 **Fig. 7.** Left: molecular structure of 1,2-indanedione. Right: representative fingerprint detected
51 using this technique with paper as substrate and observed using luminescence.
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3 **Fig. 8.** Left: molecular representation of acid yellow 7. Right: representative blood fingerprint
4 detected using this technique with aluminium foil as substrate.
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10 **Fig. 9.** Schematic illustration of the main characteristics of nanoparticles influencing their use
11 as fingerprint detection reagents. Good contrast is obtained when the ridges of the mark are
12 targeted specifically, but not the underlying substrate.
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18 **Fig. 10.** Schematic illustration of the functionalization strategy involving addition of ligands
19 on the outer surface of nanoparticles to increase affinity for the secretion residue.
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27 **Fig. 11.** Schematic illustration of the two-step metal deposition application protocols. This
28 chart shows MMD, SMD, and MMD_{lumin}, which differ in terms of which metal is used for
29 enhancement, silver, gold, and zinc, respectively.
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36 **Fig. 12.** Schematic representation of the differences among the three metal deposition
37 protocols in terms of final nanoparticle composition and detected fingerprints. MMD and
38 SMD produce dark colored fingerprints, whereas luminescent MMD (MMD_{lumin}) generates
39 luminescent fingerprints.
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48 **Fig. 13.** Schematic illustration of the two processes involved in the physical developer
49 detection technique in terms of the distance between the secretion residue and the site of
50 colloid formation. Top: in solution. Bottom: close to the residue.
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57 **Fig. 14.** Left: schematic representation of the oleylamine-functionalized gold nanoparticle
58 used as nanopowder dusting on fingerprints (Choi et al. 2006). Right: schematic
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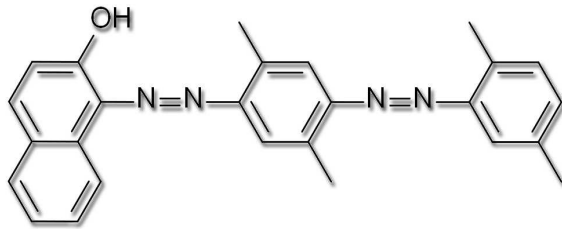
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3 representation of the alkanethiol-capped gold nanoparticle (Au-NP-C18) used to detect
4 sebaceous fingermarks on porous and nonporous surfaces in an enhanced physical developer
5 process (Sametband et al. 2007).
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12 **Fig. 15.** Schematic representation of the "protein A/antibody"-functionalized gold
13 nanoparticle used to detect fingermarks by antibody/antigen recognition. The illustrated
14 fingermark was obtained from a male smoker after 40 min sweating and detected using the
15 anti-cotinine-functionalized nanoparticles with Alexa Fluor 546 as luminescent marker.
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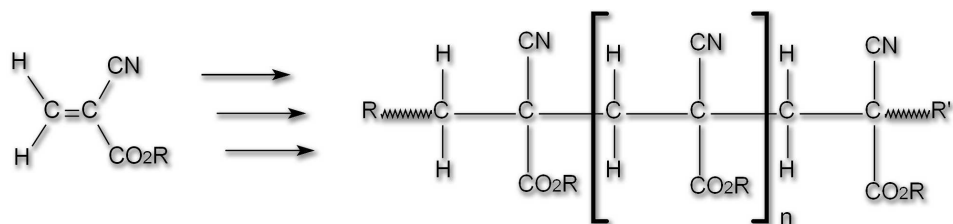
Fingermark reproduced with permission from (Leggett et al. 2007), Copyright Wiley-VCH Verlag GmbH & Co. KGaA.

Fig. 16. Left: schematic representation of the CdTe/CdS quantum dot used to detect
fingermarks on nonporous substrates (Becue et al. 2009). Right: representative fingermark
detected using this technique with aluminium foil as substrate.

Fig. 17. Potential outer shell functionalization of a silica nanoparticle with the objective of
promoting the interactions with organic compounds such as amino acids and proteins in the
secretion residue.



(Left) Molecular representation of Oil Red O; (Right) Representative fingerprint detected using this technique (paper as substrate).
109x61mm (400 x 400 DPI)

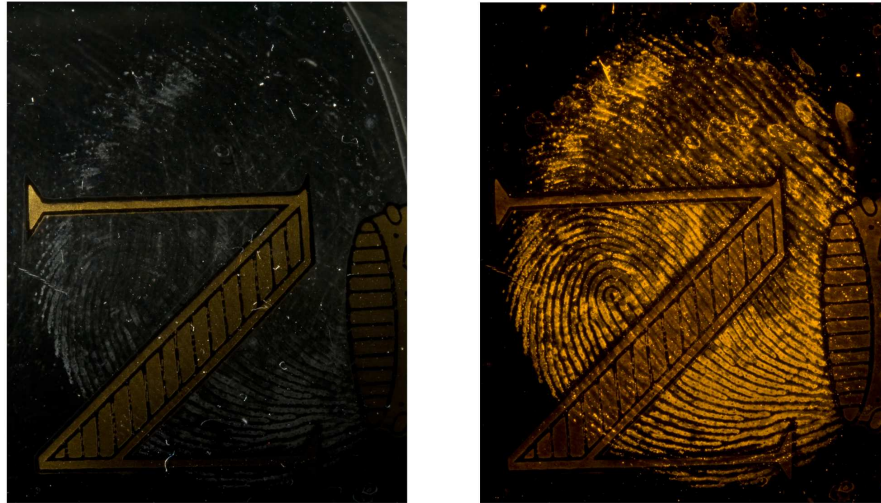


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Polymerization mechanism taking place on the secretion residue during the cyanoacrylate fuming process.

133x33mm (400 x 400 DPI)

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(Left) Cyanoacryle-fumed fingerprint located on a beer glass (the quality of development is not optimal), (Right) Same fingerprint after enhancement using Rhodamine 6G as cyanoacrylate dye (observation in luminescence).
140x76mm (400 x 400 DPI)

Review Only

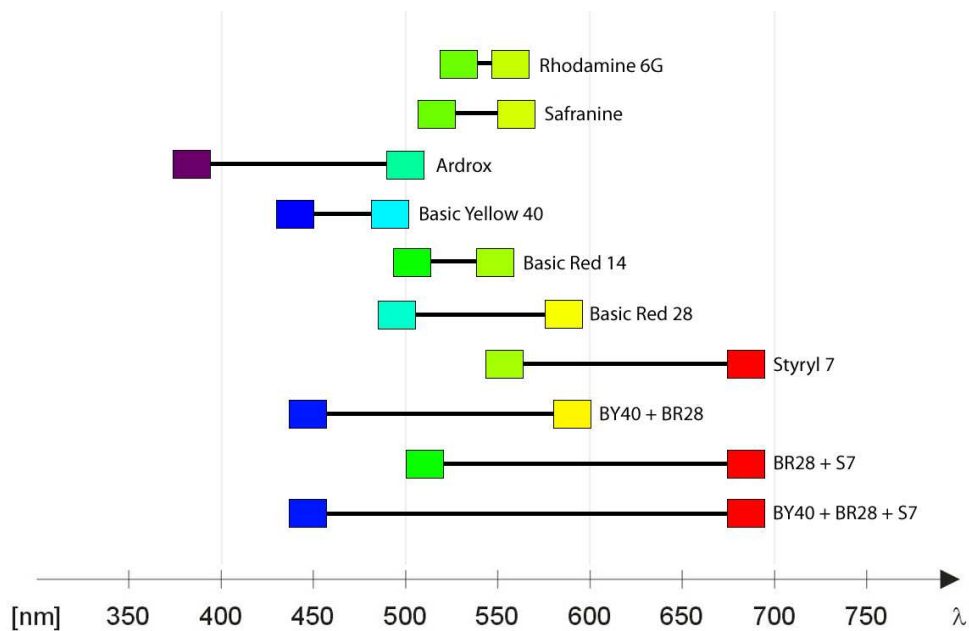
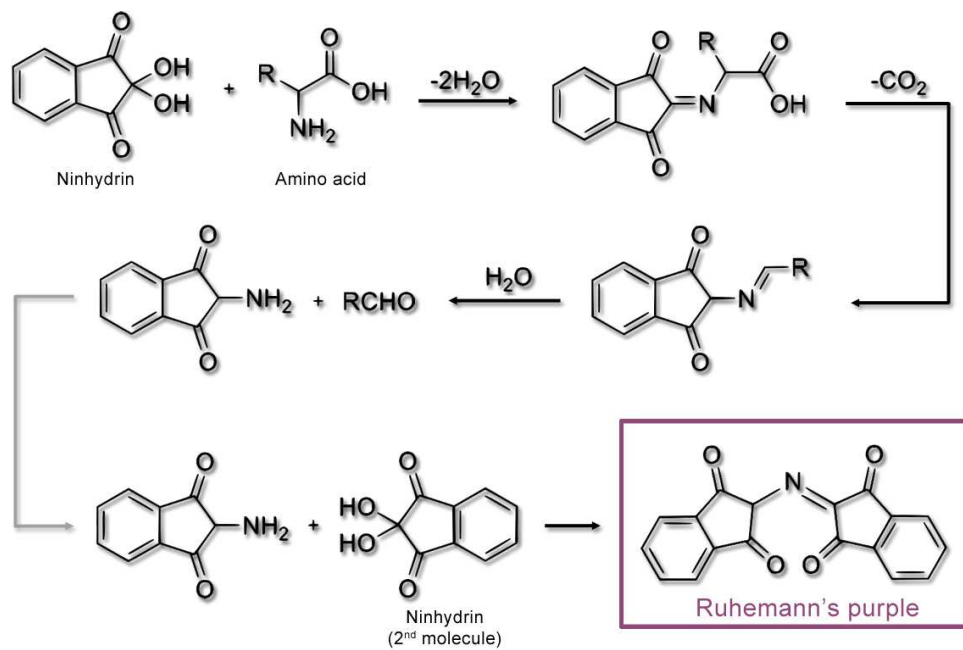
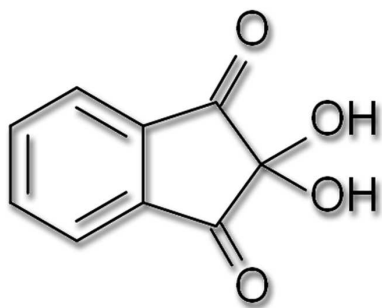


Chart illustrating the excitation and emission wavelength ranges for the most commonly used cyanoacrylate dyes.
99x63mm (300 x 300 DPI)

view Only



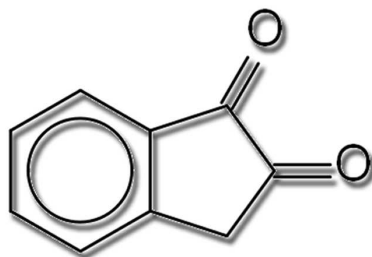
Reaction mechanism between Ninhydrin and amino acid, leading to the obtaining of a dark-purple product: the Ruhemann's purple.
99x66mm (300 x 300 DPI)



(Left) Molecular representation of Ninhydrin (2,2-dihydroxy-1,3-indanedione); (Right) Representative fingermark detected using this technique (paper as substrate).

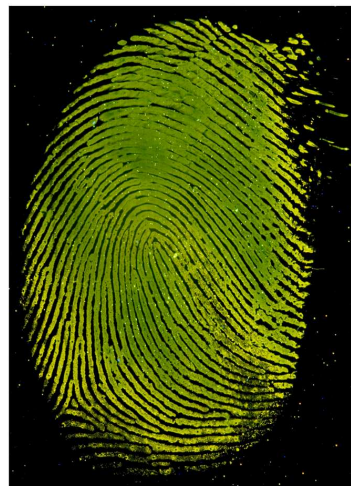
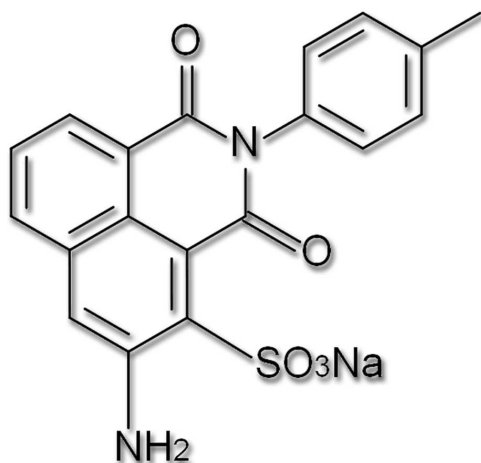
109x64mm (400 x 400 DPI)

Review Only



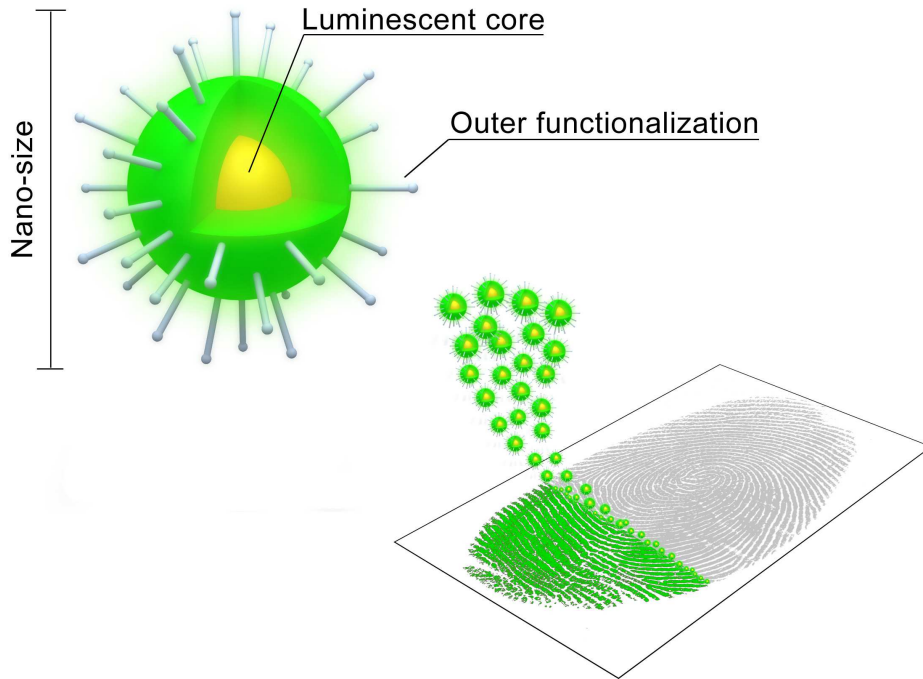
(Left) Molecular representation of 1,2-Indanedione; (Right) Representative fingerprint detected using this technique (paper as substrate; observation in luminescence).
108x64mm (400 x 400 DPI)

Review Only



(Left) Molecular representation of Acid Yellow 7; (Right) Representative blood fingerprint detected using this technique (aluminium foil as substrate).
120x64mm (400 x 400 DPI)

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Schematic illustration of the main characteristics of nanoparticles from which benefit should be taken to be used as fingerprint detection reagents. A good contrast is obtained if the ridges are specifically targeted, and not the underlying substrate.

150x118mm (400 x 400 DPI)

View Only

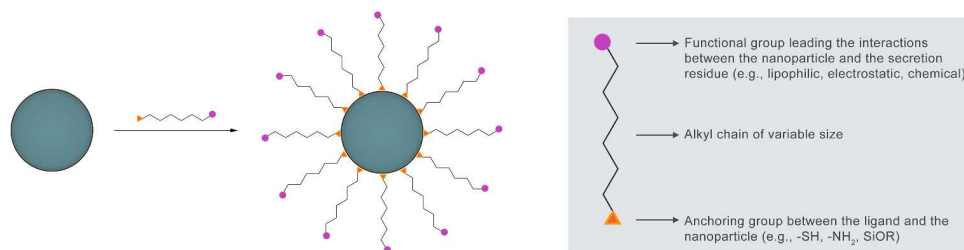
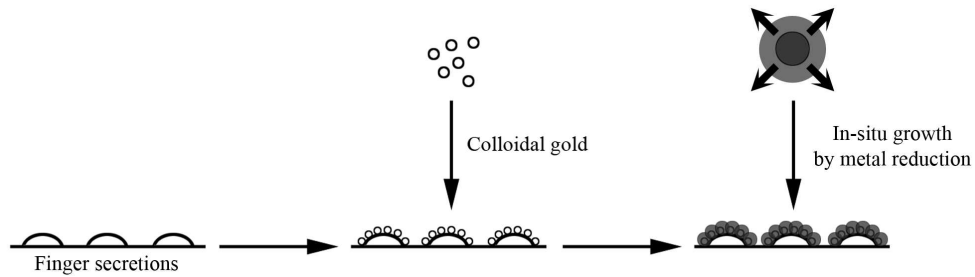


Illustration of the functionalization strategy consisting in adding ligands on the outer surface of nanoparticles to increase their chemical potential in terms of affinity for the secretion residue.
200x53mm (400 x 400 DPI)

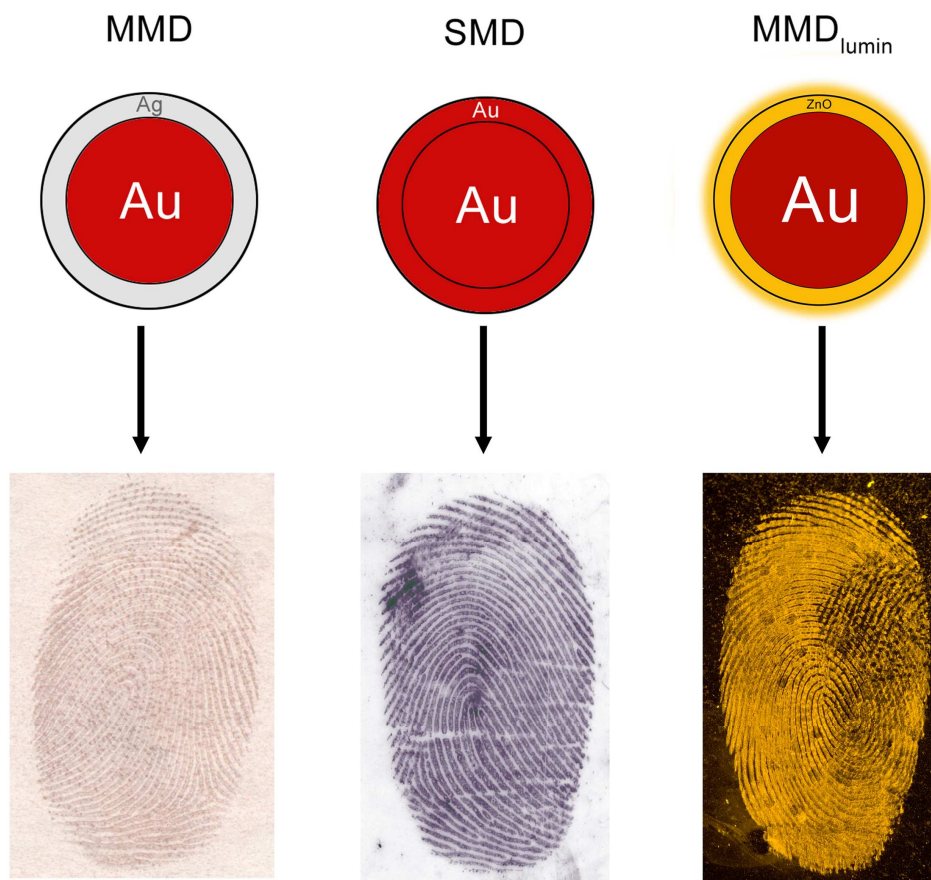
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Simplified illustration of the "two-steps" metal deposition application protocol. This chart is valid for multimetal deposition (MMD), single-metal deposition (SMD), and luminescent MMD (MMDlumin), which differ by the choice of the metal in the enhancement step: silver, gold, and zinc oxide, respectively.

127x39mm (400 x 400 DPI)

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Schematic representation of the differences between the three metal deposition protocols in terms of final nanoparticle composition and detected fingerprint aspect. Multimetal deposition (MMD) and single metal deposition (SMD) permit to obtain dark-coloured fingerprints, whereas luminescent MMD (MMD_{lumin}) permits the observation of luminescent fingerprints.

149x138mm (400 x 400 DPI)

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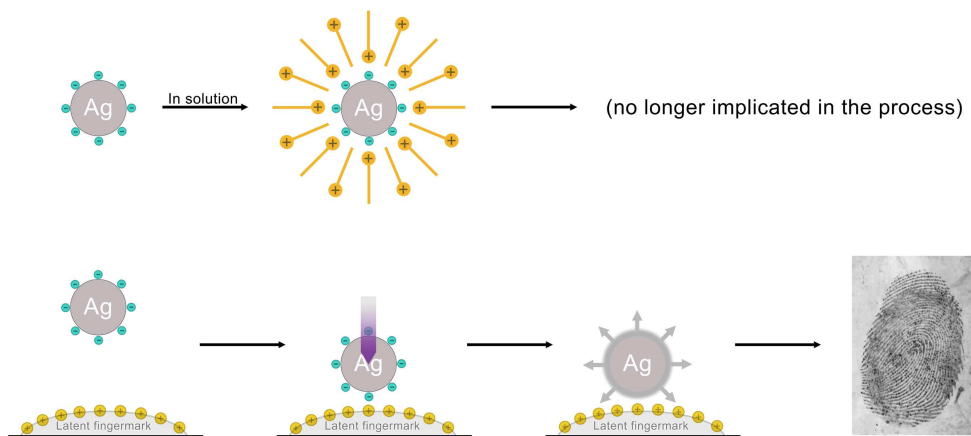
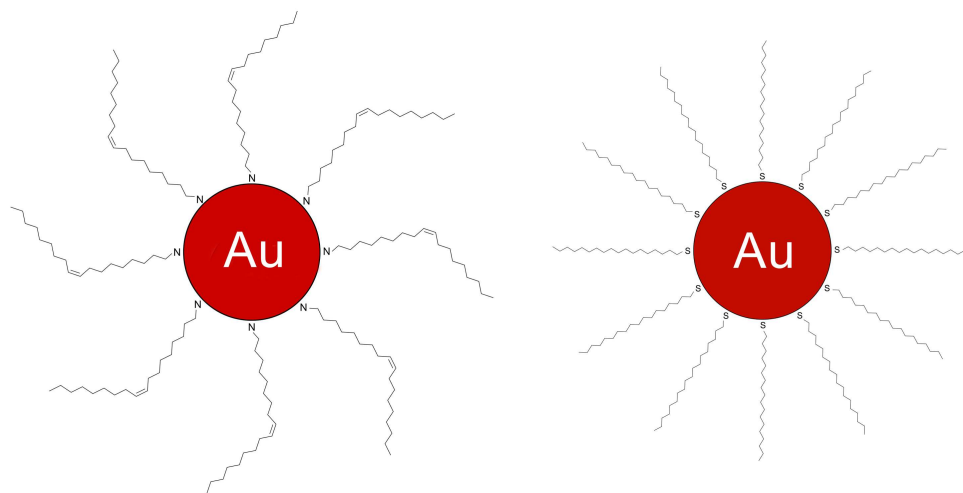


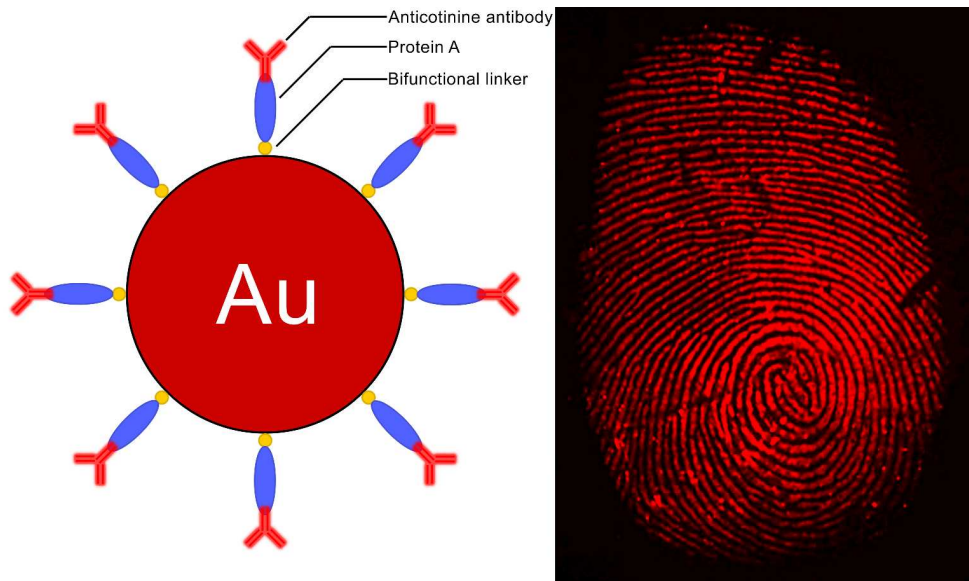
Illustration of the two processes involved in the physical developer detection technique, according to the distance between the secretion residue and the colloid formation site ("in solution" or "close to the residue").
175x79mm (400 x 400 DPI)

Review Only



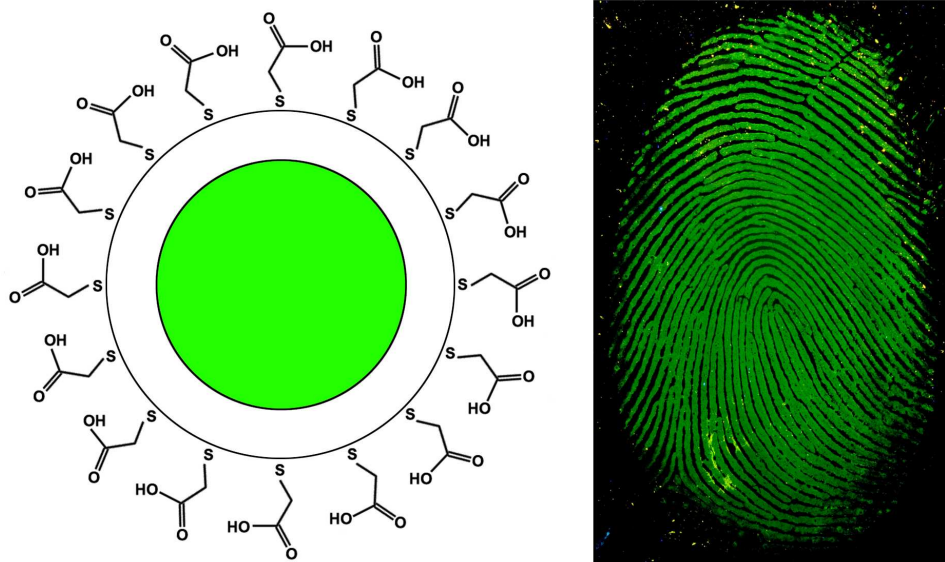
(Left) Schematic representation of the oleylamine-functionalized gold nanoparticle used as nanopowder to be dusted on fingerprints (Choi et al., 2006). (Right) Schematic representation of the alkanethiol-capped gold nanoparticle (Au-NP-C18) used to detect sebaceous fingerprints on porous and non-porous surfaces in an enhanced physical developer process (Sametband et al., 2007).

190x101mm (400 x 400 DPI)

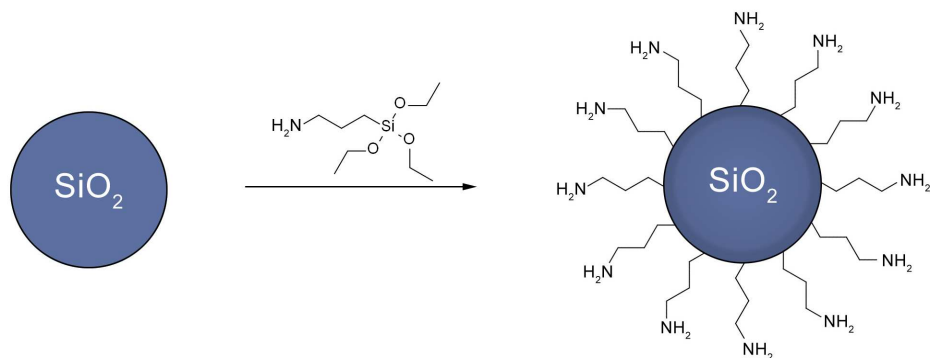


Schematic representation of the "protein A / antibody"-functionalized gold nanoparticle used to detect fingerprints by taking benefit of the highly-specific antibody/antigen recognition process. The illustrated fingerprint has been obtained from a male smoker after 40 min sweating and detected using the anti-cotinine-functionalized nanoparticles and Alexa Fluor 546 as luminescent marker (source of the fingerprint: (Leggett et al., 2007)).

200x114mm (400 x 400 DPI)



(Left) Schematic representation of the CdTe/CdS quantum dot used to detect fingermarks on non-porous substrates (Becue et al., 2009); (Right) Representative fingerprint detected using this technique (aluminium foil as substrate).
123x76mm (400 x 400 DPI)



Potential outer shell functionalization of a silica nanoparticle having for aim to promote the interactions with organic compounds such as amino acids and proteins contained in the secretion residue.

132x52mm (400 x 400 DPI)