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Sexual dimorphism in cancer

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Abstract

The incidence of many cancer types is significantly higher in the male than female populations, with associated differences in survival. Occupational and/or behavioral factors are well known underlying determinants. However, cellular/molecular differences between the two sexes are also likely to be important. We are focusing here on the complex interplay that sexual hormones and sex chromosomes can have in intrinsic control of cancer initiating cell populations, tumor microenvironment and systemic determinants of cancer development like the immune system and metabolism. A better appreciation of these differences between the two sexes could be of substantial value for cancer prevention as well as treatment.

Introduction (Epidemiology)

Epidemiological studies point to sexual dimorphism as a relevant factor for cancer incidence and survival. Overall, females have a lesser risk and better prognosis than males in a wide range of cancer types unrelated to reproductive function, such as those of colon, skin, head/neck, esophagus, lung and liver ^{1, 2} (Figure 1). Only very few exceptions exist in which incidence is higher in women, specifically thyroid cancer ³. Noteworthy, this trend is observed independently to the ethnicity of the studied population.

The generally higher cancer risk in the male population was previously attributed to environmental exposure to chemicals or carcinogens, diet and risk behaviors such as smoking and drinking. However, even after appropriate adjustment for risk factors and statistical evaluation of the data, females still exhibit a higher degree of overall cancer protection than males ^{4, 5}, with significant differences existing also in some childhood malignancies, specifically acute T cell lymphoblastic leukemia and hepatoblastoma ⁶. This review investigates the possible cellular/molecular basis of differences in cancer susceptibility between the two sexes, focusing on the complex impact that sexual hormones and sex chromosomes can have in this context. Importantly, a significant interplay is likely to exist between the two, which could be of value for novel forms of cancer prevention and intervention.

General impact of sex-related hormone signaling in cells and organ systems unrelated to reproductive function

Both steroid and protein hormones are involved in sexual development and reproductive functions. Starting around birth, gonadal steroids control production of protein hormones, so called "*somatotrophs*", by the hypothalamus-pituitary axis with a more pronounced impact in puberty and adulthood. Sex-related somatotrophs with a direct sex-related function, such as prolactin (PRL), luteinizing hormone (LH), follicle-stimulating hormone (FSH) and gonadotropin-releasing hormone (GnRH) are mainly implicated in cancer of reproductive tissues, like prostate, ovarian and breast ⁷⁻⁹. However, as detailed further below, prolactin impinges also on cancer development in non-reproductive organs, specifically the liver ^{10,11}. Among other somatotrophs with a broader function, growth hormone (GH) has been similarly implicated in liver cancer ¹². GH affects liver gene expression differentially in the two genders, possibly in connection with its pulsatile secretion into the plasma in men and constant in women ¹³. Beside its action in the liver, GH has been implicated in tumorigenesis of other several tissues, like breast, prostate, colon, skin, endometrium and brain, even if in these cases autocrine GH production by cancer cells may be more important than pituitary secretion ¹⁴.

Relative to protein hormones, much more abundant information exists on the impact of sex steroid hormones on cancer development in non reproductive organs. These hormones, being lipid soluble, can enter the plasma membrane of target cells and interact directly with intracellular receptors that can shuttle to the nucleus to

affect gene expression¹⁵. Action of these hormones extends at the epigenetic level to DNA methylation and chromatin conformation^{16,17}. Analysis of gene expression in different non reproductive tissues indicates the existence of a gender specific influence on transcription^{18, 19}, which is paralleled at the level of chromatin organization, by specific differences between the sexes²⁰. As discussed here below, sex hormone signaling pathways are likely to affect cancer susceptibility through multiple mechanisms, impacting on intrinsic self renewal mechanisms, tumor microenvironment, immune system and metabolism. The vast majority of accrued information relates to the involvement of three major sex steroid hormone receptors, estrogen receptor (ER) α , β and androgen receptor (AR), on which we will be focusing (Fig. 2). It is important to note, however, that the progesterone receptor, under intense investigation in breast cancer²¹, may also be involved in cancer of non-reproductive organs, like what recently reported for lung NSCLC²² and colon cancer²³.

a) Stem cell renewal : Specific ablation of estrogen receptor α , β and androgen receptor affects homeostasis and/or regeneration of organs unrelated to reproductive functions, such as lung²⁴, colon²⁵ and skin²⁶⁻²⁸. Sex steroids have an impact on stem cell populations of various types. Estrogens enhance self renewal of embryonic stem cells²⁹, promote endothelial progenitor cells³⁰ and, during pregnancy, contribute to the expansion of hematopoietic stem cell populations³¹. Neural stem cells have been reported to fluctuate during the estrous cycle and exogenous estrogen administration is sufficient to increase neural stem cell division

³². 17 β -estradiol, the most potent form of estrogen, can also counteract senescence through p53-antagonizing mechanisms and/or control of telomerase expression ³³, ³⁴.

Like estrogens, androgens are involved in control of hematopoietic cell renewal. Aging men, that experience a reduction in androgen production, are prone to develop normocytic anemia, a condition characterized by a reduced number of red blood cells, that can be corrected with androgen supplementation ³⁵. Androgen supplementation has also ameliorating effects on acquired aplastic anemia, characterized by a reduced production of all blood cells, and dyskeratosis congenita, a genetic syndrome interesting several parts of the body ^{36,37}. These two diseases manifest telomere dis-function, bone marrow failure due to hematopoietic stem cells impairment and are linked to cancer development. Molecularly, in primary hematopoietic cells, androgens induce telomerase expression through aromatase-dependent conversion of testosterone into estrogen and ER α activation ³⁸.

The role of ER α signaling in breast cancer initiating cells is an area of active investigation ^{39,40}, and so is that of AR in prostate cancer ^{41,42}. By contrast, the role that these pathways play in self-renewal of initiating cancer cells of other types is a mostly unexplored topic and important area of future investigation.

b) Cancer stroma : The tumor microenvironment plays a crucial role in both initiation and progression of the disease ^{43,44}. Various components of the stromal compartment are under sex hormone control. Still not well depicted is the impact of sex hormones on the behavior of stromal fibroblasts. Cancer Associated Fibroblasts

(CAFs) can be viewed like persistently activated fibroblasts sharing properties with fibroblasts at wound sites ⁴⁴. While well known differences exist in skin wound healing between the two sexes ^{26,27}, where fibroblasts exert an important role in the first phases of the process, studies on the impact of sex hormones on CAF function have mostly focused on prostate ⁴⁵⁻⁴⁷, breast ^{48, 49} and cervical cancer ⁵⁰ types. Estrogens were shown to promote mobilization of bone marrow derived precursors to breast cancer stroma ⁴⁸ and increase stromal density ⁴⁹. In cervical cancer, ER α expression in stromal fibroblasts is sustained during the course of the disease ⁵⁰, pointing to a possibly detrimental role. By contrast, in prostate tumors, expression of ER α is positively associated with survival and restrains cancer associated fibroblast behavior reducing angiogenesis and tumor growth ⁴⁵. Such beneficial role contrasts with additional observations that implicate ER α activity as a positive regulator of prostate epithelial cells ⁵¹, suggesting that this receptor functions differently in the two compartments. By contrast to ER α , ER β activation has been reported to restrain prostate hyperplasia and cancer by inducing cell death of both epithelial and stromal cells ⁵².

Conflicting results exist concerning the role of AR signaling in prostate CAFs. While animal model studies indicate that this receptor in stromal fibroblasts promotes initial stages of the disease ⁴⁶, clinical evidence points to the opposite possibility that sustained AR expression in this context is linked to less aggressive cancer and better prognosis ⁴⁷.

Angiogenesis is another key determinant of cancer development. Estrogens have a well established pro-angiogenesis function, best known in the endometrium

during the female reproductive cycle, which is mediated by enhancement of endothelial cell proliferation and migration ^{30, 53}. This is mediated by diffusible factors like PAF ⁵⁴ and VEGF ⁵⁵, which are frequently secreted by cancer cells in response to estrogens. In contrast to ER α , ER β activation in neoplastic breast cells negatively affects blood vessel formation ⁵⁶. Androgens also stimulate endothelial cell migration and new blood vessel formation through VEGF, especially in cells taken from male tissues, due to higher AR expression ⁵⁷.

As for cancer initiating cells, the role that ER and AR signaling play in the stromal compartment of tumors of non-reproductive systems is an area of great importance for future studies.

c) Inflammatory/immune system: The microenvironment encompasses different leukocyte populations, with macrophage M1 polarization playing a key role in chronic inflammation-associated cancer development ⁴³. In macrophages, ER α significantly reduces their pro-inflammatory activity ⁵⁸ and tumor promoting properties ⁵⁹. In contrast, AR favors the migration of macrophages and the release of proinflammatory cytokines delaying wound healing ²⁸. Mice lacking this receptor in inflammatory cells resolve the wound quicker than wild type mice. Macrophage release of cytokines is dependent on AR and contributes to prostate tumorigenesis ⁶⁰.

Besides macrophages, other cells of the myeloid and lymphoid lineages like neutrophils or lymphocytes express estrogen and androgen receptors and these sex hormones can modulate the immune system at multiple levels through control of

immune-modulatory genes like *IFN- γ* ⁶¹. Pathogenesis of autoimmune diseases, to which females are much more susceptible than males, involves an altered balance between effector T cells, with cytotoxic function, and regulatory (CD4+CD25+) T cells (Treg), a subpopulation of T cells that promotes self tolerance and restrains cancer immune surveillance⁶². The Foxp3 transcription factor is a master regulator of Treg cell number and function⁶³. It belongs to the fork-head family of transcription factors and is encoded by the X-linked *foxp3* gene. Polymorphisms and mutations of this gene are associated with a number of diseases with immunodysregulation and poly-endocrinopathies. *foxp3* expression and function are under combined control of T cell receptor signaling and multiple cytokines through both transcription and post-transcription regulatory mechanisms⁶³. In women, size of Treg cell populations positively correlates with estradiol levels during the luteal cycle and in pregnancy⁶¹, with estradiol treatment inducing both Treg expansion and Foxp3 expression⁶⁴. Testosterone stimulation causes also a strong increase of Treg cells both *in vivo* and *in vitro*, through a mechanism that involves AR recruitment to the *foxp3* locus and its enhanced expression⁶⁵.

The fact that both estrogens and androgens are involved in positive regulation of the *foxp3* gene suggests that additional more indirect mechanisms are involved in the differential activity of Treg cells in the female versus male populations. One such mechanism may involve differential production and function of IL6. This cytokine, together with IL-1, promotes degradation of the Foxp3 protein and ensuing Treg suppression⁶⁶. Importantly, IL-6 expression is positively

regulated by estrogens ⁶⁷ and genetic polymorphisms in the IL6 locus were identified that may be involved in its differential control between the two sexes ⁶⁸.

Besides a differential impact on various cell populations, sex hormones can also affect the immune system indirectly, through the gut microbiome. In Non Obese Diabetic (NOD) mice, susceptible to autoimmune diabetes, the rise of testosterone after puberty promotes a divergent bacterial commensal population from adult females and prepuberal animals ⁶⁹. Such effect, which is reversed upon castration, is associated with reduced susceptibility to autoimmune reaction linked with changes in cytokine expression in lymphoid and myeloid cells.

The X chromosome is also highly enriched in immune-related genes ⁶¹ and there are several X-linked microRNAs that may contribute to sex-specific regulation of the immune response by targeting immune-related genes ^{70,71}. In one study of an autoimmune disease, Systemic lupus erythematosus (SLE), several X-linked miRNAs were found overexpressed in T cells of female versus male patients ⁷². Several of the differentially expressed miRNAs are located within 5000bp of an estrogen response element and, among the predicted targets, was CBL, a negative regulator of T cell receptor activity, which is down-regulated in T cells of the SLE patients ⁷².

Overall, a complex picture emerges whereby sexual dimorphism in the immune system is the combined result of multiple determinants ⁶¹. Irrespectively of the detailed mechanisms, the greater susceptibility of women to develop autoimmune diseases is likely accompanied by enhanced immune surveillance

against various tumor types. As discussed at the end, further insights into this topic will likely lead to improved forms of cancer immune therapy and prevention.

d) Metabolism : Steroid hormones are also central regulators of systemic metabolism, acting at several levels ⁷³. Both estrogen and androgen receptors increase glucose tolerance and restrain visceral fat accumulation ⁷⁴. Reduced ER α signaling in post-menopausal women and aromatase deficiency, with compromised conversion of androgens to estrogens, are associated with increased adiposity ⁷³, a known risk factor for cancer development ⁷⁵. Cancer is often associated with a severe body wasting syndrome called cachexia ⁷⁶. Cachexia is a sexually dimorphic trait ⁷⁷ that is linked to higher levels of autophagy in male hearts and is associated with reduced survival ⁷⁸. Androgens have also an important muscle trophic function and the majority of male cancer patients in the seventh decade of life experiences sarcopenia, a condition characterized by the loss of muscle mass and strength, linked to a reduction in testosterone ⁷⁹. A decrease in androgen levels has been noticed in multiple studies in male cancer patients ⁸⁰ that may be a result of chemotherapy ⁸¹. Selective androgen modulators, able to activate AR mainly in the muscle, have been tested for their ability to revert or compensate for body mass wasting with improvement of lean body mass and physical condition ⁸².

Sex hormone signaling in specific cancer types

A clear example of greater cancer susceptibility of the male versus female populations due to hormonal sex influences is represented by *hepatocellular carcinoma* (HCC), which occurs more frequently in men ^{1, 2}. A detailed analysis indicates that AR and ER α contribute to hepatocarcinogenesis in an antagonistic way regulating gene expression (Fig. 3): the first stimulates while the second restrains proliferation and nucleotide/ amino acid metabolism ⁸³. Such estrogen induced resistance and androgen mediated enhancement of tumorigenesis is dependent on FOXA1 and FOXA2 pioneer factors, required for chromatin recruitment of nuclear receptors. Ablation of FOXA proteins eliminates gender differences converting the tumor suppressive role of estrogen into tumor promoting ⁸³. Estrogens exert a protective function reducing the production of inflammatory mediators like IL6 ⁸⁴. However, important estrogen effects on liver physiology can also be mediated by its effects on the gonadal-hypophyseal axis ^{85,86}. In rodents, hypophysectomy abolishes sex-dependent differences in HCC ⁸⁷. Estrogen can modulate growth hormone signaling acting on pituitary GH secretion ⁸⁸, as well as on control of GH receptor (GHR) expression ⁸⁹. GH in turn targets the liver in a sex-specific manner, which is particularly evident in rodents at puberty, but is also found in humans ⁹⁰. GH function is mediated by the transcriptional activator STAT5, which can protect hepatocytes from chronic injuries and malignant transformation ¹², providing another possible mechanism for sexual dimorphism in HCC development.

The lesser susceptibility of women to HCC can also result from estrogen stimulated secretion of prolactin, another pituitary-secreted hormone ^{10,11}. In this

context, prolactin may protect females from liver tumorigenesis by restricting innate immunity signaling in hepatocytes, which may be of special relevance for the virally-associated forms of the disease ¹⁰. More specifically, prolactin stimulation of hepatic cell lines was shown to suppress response to IL1R, TLF4 and TNFR1 activation by a post-transcriptional mechanism, thereby suppressing c-Myc activation ¹⁰.

Several other cancer types exhibit higher incidence and aggressive behavior in the male and post- versus pre-menopausal female populations, with similar gender differences occurring in corresponding animal models. These include *colon cancer, gliomas, melanoma and non melanoma skin cancer, head/neck squamous cell carcinoma (SCC), esophageal and non small cell lung cancer (NSCLC)*. Protection in women extends to hematological malignancies, especially lymphomas, even after menopause ^{1,2}.

In many cases, higher estrogen signaling, leading to ER β activation, has been implicated as the likely explanation. ER β expression is often diminished in these various cancer types and sustained ER β proliferation is a favorable prognostic marker. In colon cancer, a polymorphism in the ER β promoter, possibly influencing its activity levels, has been associated with increased survival ⁹¹. Polymorphisms in the EGFR gene have also been identified that are positively associated with colon cancer survival in women and negatively in males ⁹², pointing to a possible interplay between EGFR and ER β signaling underlying the differences between the two sexes.

In skin and head/neck SCC cells in which ER β expression is down-modulated, increased ER β expression and/or agonist-specific stimulation promotes

differentiation through induction of NOTCH1 gene transcription and function ¹⁹. Also in melanoma, ER β expression levels are inversely related to primary cancer progression in both men and women ⁹³ and, in a mouse model of the disease, tamoxifen treatment significantly inhibited metastatic spread in female animals ⁹⁴. ER β function is required for normal lung and colon morphogenesis ^{24, 25} and its activation exerts beneficial consequences in experimental models of colon ⁹⁵, glioma ⁹⁶, mesothelioma ⁹⁷, renal cell carcinoma (RCC) ⁹⁸ and T cell lymphoma ⁹⁹.

In contrast to the above tumor types, the incidence of thyroid cancer is significantly higher in the female than male populations. Even though several studies tried to find associations between this neoplasm and hormonal or dietary factors, the reasons for this discrepancy are still unclear ³. However, experimentally, it should be noted that AR expression in thyroid follicular cells was reported to reduce proliferation ¹⁰⁰, while estrogen treatment induced proliferation and suppressed apoptosis ¹⁰¹, suggesting a possibly opposite role of sex hormones in this context.

Influence of sex chromosomes on oncogenesis

The most significant genetic differences between the two sexes reside in the X and Y chromosomes. As discussed below, mutations and/or epigenetic deregulation of genes on sex chromosomes could contribute to the oncogenetic process. Besides protein-encoding genes, deregulated expression of non-coding

RNAs is likely to be involved, with a likely interplay with sex hormone action, including the fact that the androgen receptor gene is X-linked (Fig. 4).

a) X chromosome

To counteract differences in X chromosome gene dosage, mammals utilize the X inactivation (Xi) process, involving transcriptional silencing and chromatin compaction promoted by the long non coding RNA (lncRNA) Xist^{102, 103}. Female tissues are mosaic for most X-linked genes, resulting from Xi in every cell. Although inactivation of one X chromosome over the other is generally random, preferential inactivation of one chromosome (skewed Xi) can protect women against the consequences of X-linked gene mutations¹⁰⁴. On the contrary the absence of Xi can have detrimental effects: for instance loss of Xist has been reported in breast, ovarian and cervical cancer cell lines^{105, 106} and, only in female mutant mice, it caused deregulation of hematopoietic lineages and emergence of myeloid neoplasms associated with genome wide changes in gene expression¹⁰⁷.

In other settings, cancer development has been linked to increased rather than decreased Xist expression. Xist expression can be up-regulated in glioblastoma cells of both sexes and its knockdown results in tumour-suppression¹⁰⁸. In testicular tumors, increased Xist expression is also frequently observed, which is not coupled with methylation of X linked genes, indicating that its role may be distinct from this process¹⁰⁹.

Despite the X chromosome-wide silencing mechanism, there are regions escaping Xist-dependent inactivation. Besides the pseudoautosomal regions (PAR),

which have corresponding regions of homology on the Y chromosome, other single genes escaping inactivation are spread throughout the chromosome. In particular, UTX (KDM6A) is a H3K27 demethylase expressed from both X chromosomes ¹¹⁰. In men, UTX has a homolog on the Y chromosome, called UTY (KDM6C), whose catalytic activity is very low or absent ¹¹¹. UTX has a tumor suppressing function and inactivating mutations in this gene are not compensated by UTY (Fig. 4). This appears to be of importance in human T-acute lymphocyte leukemia (ALL), a childhood tumor more common in males than in females and a corresponding experimental model of the disease ¹¹²⁻¹¹⁴. Additional X-linked players altered in T-ALL are the PHF6 and RPL10 genes, which are mutated almost exclusively in male patients ^{115, 116}. Loss-of-function UTX mutations have also been found in oesophageal SCCs and renal cell carcinomas, which are more frequent in the male population ^{117,118}. Group 4 medulloblastomas are also more prevalent in males and it is noteworthy that, besides UTX, several other mutated oncogenes and tumor suppressor genes in these tumors are located on the X chromosome ¹¹⁹ (suppl. Table 1). These include the genes for a component of histone-deacetylase complexes, ZMYM3, and the RNA helicase DDX3X, which has a homologue on the Y chromosome that is not translated. While ZMYM3 is mutated exclusively in male patients' tumors, DDX3X is also found to be mutated in females' tumors, in the allele that escapes X inactivation ¹²⁰. Loss of function mutations in DDX3X are also observed in Natural killer/T-cell lymphoma (NKTCL) ¹²¹ and NSCLC, where the absence of the helicase promotes malignancy ¹²².

Other tumor suppressor genes mapping in the X chromosome include *WTX*, which is involved in pediatric kidney cancer Wilms tumors ¹²³, and *KDM5C*, coding for a histone demethylase, whose truncating mutations are found in renal carcinoma cells and trigger genomic instability ¹²⁴ (suppl. Table 1). Of note, *KDM5C* is also overexpressed in HCCs in which it seems to play a pro-oncogenic function ¹²⁵.

As mentioned, *foxp3* is a X-linked gene, which plays an important role in the immune system, by controlling transcription of regulatory T cells. Besides Tregs, expression of this gene has been observed in different tumor cells, like lung, melanoma, colon cancer cells ¹²⁶, as well as pancreatic carcinoma cells ¹²⁷, which share some Foxp3-dependent immune suppressing functions with Tregs. Furthermore, in breast and prostate cancer Foxp3 inhibits NFκB activation, through a Foxp3-miR-146-NFκB axis, which promotes tumor suppression and apoptosis ¹²⁸. We further note that a specific member of the SOX family of transcription factors, *SOX3*, is encoded in the X chromosome (Fig. 4). This will be discussed below in the context of the Y-encoded *SRY* gene.

Besides protein-coding genes and lncRNAs like *Xist*, the X chromosome is also highly enriched in miRNAs, whose density is almost two-fold higher than in the autosomes (suppl. Table 1). Among these are miR-221 and miR-222, which function as oncomiRs through repression of the cell cycle inhibitor p27 ¹²⁹. Interestingly, these two miRs map relatively close to *KDM6A* gene, raising the possibility that they may also escape Xi. Together with miR-222, miR-223 (also X-linked) is up-regulated in gastric cancer, promoting cancer cell migration and metastasis ¹³⁰. Expression of

two other X-linked miRs, miR-20b and miR-361, has potential prognostic significance for oropharyngeal carcinoma ¹³¹. There is also a cluster of several miRs (miR-506, miR-507, miR-508, miR-513a-1, miR-513a-2, miR-513b, and miR-513c) that might fine-tune the response of cells to PI3K/Akt activation. Expression of these miRs is under direct control of the Akt-dependent FOXO1 and FOXO3 transcription factors. In turn, the miRs target these transcription factors as well as components of the PI3K and of MAPK cascade ¹³².

Although all these miRs map to the X chromosome, whether or not their differential expression contributes to gender differences in cancer risk remains to be established.

b) Y chromosome

Broad as well as restricted alterations of the Y chromosome are frequent in cancer development. Loss of the entire Y chromosome (LOY) has been reported with various frequencies in prostate ¹³³⁻¹³⁵, pancreatic ¹³⁶, colorectal ¹³⁷ and bladder ¹³⁸, which has also a strong male prevalence. Short arm deletions have also been found in numerous cancer types ¹³⁹ (<http://cgap.nci.nih.gov/Chromosomes/Mitelman>). Importantly, Y chromosome loss occurs also in normal tissues; it is frequently observed in normal hematopoietic cells of elderly men and is significantly associated with a greater risk of cancer ^{140, 141}. Y chromosome loss in blood cells could be used as predictive biomarker and is also induced by smoking ¹⁴¹. However, it remains to be determined whether or not widespread loss of Y chromosomal regions

promotes cancer development or simply results from elimination of unnecessary genetic material, as proposed during evolution¹⁴².

More limited alterations of the Y chromosome can also occur. Of likely importance are those affecting the pseudoautosomal region and the closely located SRY gene, responsible for male sex development¹⁴³. Chromosomal amplifications of the pseudoautosomal region Y11p2 adjacent to SRY (located at Y11p3) have been found in human HCCs¹⁴⁴ possibly contributing to gender disparity in HCC. Indeed SRY ablation can impair development of this cancer type¹⁴⁵, decreasing also chemoresistance¹⁴⁴ (Figure 3). SRY codes for a HMG box transcription factor of the SOX family. Among various members, SRY shares a significant amount of homology with Sox1, 2 and 3, and may have evolved from these¹⁴⁶. Like Sox2¹⁴⁷, SRY is likely to play a role in cancer stem cell renewal, as suggested by its control of OCT4 expression (Fig. 4), another key cell reprogramming and stem cell factor¹⁴⁴ and of a c-Myc coactivator, sgf29¹⁴⁵. Interestingly, together with these growth promoting functions, SRY has also been reported to negatively regulate AR gene transcription through some as yet unclarified mechanism¹⁴⁸.

TSPY is another Y-linked gene, coding for a SET/NAP (SET proto-oncogene /nucleosome assembly protein), with a known function in male germ cell differentiation, mitosis, and meiosis¹⁴⁹. Increased TSPY expression occurs in testicular germ cell tumors¹⁴⁹, as well as various types of somatic cancer, including melanoma¹⁵⁰, prostate¹⁵¹ and liver cancer¹⁵². Interestingly, while TSPY appears to exert a pro-oncogenic function, it has a homologue on the X chromosome, TSPX, which can be tumour suppressive¹⁵³ and protects from hepatitis B viral infection, a

major risk for HCC ¹⁵⁴. In fact, both ectopic TSPY activation and TSPX silencing can contribute to the initiation and progression of hepatocellular carcinoma, which, as discussed in a previous section, is a male-predominant cancer ¹⁵² (Fig. 3). In these tumors, there is a positive correlation between TSPY and AR expression, with TSPY promoting AR expression ¹⁵⁵. Conversely, at least in prostate cancer cells, AR stimulation by androgen leads to TSPY up-regulation ¹⁵⁶.

Finally, RBMY (RNA binding motif gene on the Y chromosome) codes for a male germ cell-specific regulator of RNA splicing involved in spermatogenesis. RBMY is aberrantly activated in male HCC ¹⁵⁷, where, like TSPY, it can enhance AR expression and activity as well as cancer development ¹⁵⁸ (Fig. 3).

Conclusions and future perspectives

Sex-specific differences in cancer incidence and mortality across ethnic backgrounds point to significant influences of gender on both cancer initiation and progression (Fig. 1). An important notion is that cancer as a disease is not limited to a group of genetically deranged cells and their immediate environment, but is linked to general alterations in organ homeostasis as a result of perturbed developmental/morphogenetic signals and systemic factors ^{43, 159}. Selection of cells with cumulative somatic mutations ¹⁶⁰ or a “Big Bang” expansion of single cell populations with pre-existing dominant alterations ¹⁶¹ have been implicated in cancer development. However, there can be clonal expansion of cells harboring many cancer driver mutations without any clinical signs of the disease ¹⁶². Indeed, in

most cases, premalignant lesions do not progress into malignancy for reasons that are poorly understood ^{163, 164}. This led to the view of an “ecological cellular environment” playing a key role in restraining or unleashing tumor growth ^{164, 165}. As discussed throughout this review, sex hormones and sex chromosome-encode information can impact on all aspects of the cancer process (Fig. 5).

As for the tumor cells themselves, important molecular insights can be provided by recent developments in large-scale genome sequencing of tumors coupled with epigenetic, transcriptomic and proteomic analysis and associated clinical information. For instance, analysis of the TCGA database showed differences in the mutational landscape of Head/Neck SCCs in the male versus female populations, with distinct gene expression profiles ¹⁹. It will be interesting to assess whether similar differences apply to other cancer types, possibly in a cell type or cancer subtype-specific manner, and to determine whether subsets of cancer patients with mutational and/or gene expression profiles closer to those of the other gender can be identified, with distinct clinical behavior.

At the level of the cancer stroma and organ homeostasis, sex hormone regulation of angiogenesis stands out as a key element of divergent cancer progression between the two sexes, as does inflammation. Systemically, the most promising area where understanding sex bias can be of value is at the level of the immune system. Immunotherapy based on immune checkpoint inhibitors is rapidly emerging as a key therapeutic mainstay of cancer types with high gene mutation frequencies ¹⁶⁶, most of which exhibit much greater incidence in the male than female populations. As we discussed, sex hormone modulation of the immune

system can be involved as well as X-linked expression of cytokine genes and immune-modulatory miRNAs. It will be important to assess to what extent these elements have an impact on key immune checkpoints, like the PD-1/PD-L1 axis, acting on various T cell populations, neighboring immunomodulatory cells and cancer cells themselves.

Sexual dimorphisms that favour females are also observed in lifespan and aging ¹⁶⁷. Such differences are once again defined by multiple elements. Sex hormones, especially estrogens, can play an important role in control of cellular senescence and aging-associated tissue deteriorations. Concomitantly, at the chromosomal level, it has been suggested that X-linked mutations are more likely to exert detrimental effects in male than female cells, which have the advantageous possibility of selective inactivation of the mutations-carrying X chromosome ¹⁰⁴.

Besides its basic value, detailed knowledge of the impact of sex hormones on cancer development in organs of non reproductive function has translational importance, given the available pharmacological tools to either enhance or suppress their function. This is especially true for steroid hormones ^{168,169}. Along these lines, a topic of great interest and translational potential is that of the interplay between their influence and epigenetic reprogramming of cells, with a well known interplay between ER and AR function and key histone and DNA modifying enzymes ¹³, which can be targeted by a large battery of natural and synthetic compounds ¹⁷⁰. Interventions at this level are going to be of great promise for both cancer prevention and treatment.

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Figure legends

Figure 1. Gender differences in cancer incidence and mortality in organs unrelated to reproductive function.

Incidence (A) and mortality (B) rates of various cancer types for the year 2012 in males (M) and females (F) of European, African, Americas and South East Asian regions. Rates (number of cases per 1000) are for all ages and were retrieved from GLOBOCAN 2012 (IARC) (<http://globocan.iarc.fr>).

Figure 2. Role of sex steroid hormone receptors in non reproductive tissues.

Summary scheme of the complexity of biological functions of the three sex steroid hormone receptors that have been so far implicated. As discussed in the text, activation of ER α elicits hematopoietic stem cell expansion and mobilization, favors skin wound healing enhancing keratinocyte proliferation, promotes blood vessel formation and endothelial cell precursor mobilization, reduces liver cell proliferation and restrains the inflammatory role of macrophages. ER β also blocks macrophage activation and, contrary to ER α , negatively regulates angiogenesis. ER β promotes differentiation and suppresses tumor formation in several districts such as the skin (acting on both keratinocytes and melanocytes) and is required for the proper morphogenesis of lung and colon. AR suppresses wound healing, promotes angiogenesis, liver cell proliferation as well as macrophage activation. Androgens activity is not limited to AR activation. In fact, through the action of aromatase, they

are converted into estrogens, thus controlling indirectly hematopoietic stem cell expansion through ER α activity.

Figure 3. Possible mechanisms contributing to gender dimorphism in hepatocellular carcinoma (HCC).

Diagrammatic summary of factors implicated in the greater incidence of HCC in the male versus female populations. At hormonal level, AR and ER α activation have opposing effect on hepatocytes proliferation and nucleic acid metabolism, acting on gene transcription. Estrogen exerts also indirect effects through stimulation of prolactin and growth hormone (GH) secretion by the pituitary gland, with these hormones in turn suppressing inflammatory and proliferative signals in the liver. At the chromosomal level, the X-linked gene TSPX protects from hepatitis B viral infection, a major risk for HCC development and loss of TSPX has been observed in male patients with HCC. On the other hand, TSPY, the TSPX homolog gene on Y chromosome, induces AR expression and is found ectopically expressed in HCC male patients. Amplification of two other Y-linked genes, RBMY and SRY, occurs in HCC, with RBMY enhancing AR expression and SRY promoting tumorigenesis as well as chemo-resistance.

Figure 4. Interplay between sex chromosomes in human cancer.

Representative illustration of X- and Y-linked genes of functional significance discussed in the text. In women, some parts of the X chromosome escape Xist-mediated inactivation, as is the case for the tumor suppressor gene UTX, which is

not compensated by its Y homologue UTY, resulting in differential gene dosage effects with likely cancer-protective consequences. Three other genes on the Y chromosome, SRY, RBMY and TSPY, have the potential of promoting cancer development, which may be mediated, in part, by their modulation of X-linked AR expression. Increased SRY activity may also drive cancer stem cell expansion through increased expression levels of autosomal genes like OCT4.

Figure 5. Hormonal and chromosomal contributions to sexual dimorphism in cancer.

As discussed in the text, a complex interplay exists between sex hormones and X and Y chromosomes, with epigenetic control of gene expression as a link. These combined factors can differentially affect cancer development at multiple levels: i) cancer target cells of origin, through modulation of ubiquitous or cell type specific intracellular processes; ii) cancer stroma and surrounding “field cancerization” tissues, through angiogenesis and inflammation; iii) systemically, through alterations of the immune system and metabolism. It can be further proposed that drug targeting of sex hormone action and associated epigenetic control mechanisms can have beneficial effects for multiple cancer types, well beyond those of organs with reproductive functions.

References

1. Siegel, R.L., Miller, K.D. & Jemal, A. Cancer statistics, 2015. *CA Cancer J Clin* **65**, 5-29 (2015).
2. Torre, L.A. et al. Global cancer statistics, 2012. *CA Cancer J Clin* **65**, 87-108 (2015).
3. Rahbari, R., Zhang, L. & Kebebew, E. Thyroid cancer gender disparity. *Future Oncol* **6**, 1771-9 (2010).
4. OuYang, P.Y. et al. The significant survival advantage of female sex in nasopharyngeal carcinoma: a propensity-matched analysis. *Br J Cancer* **112**, 1554-61 (2015).
5. Wisnivesky, J.P. & Halm, E.A. Sex differences in lung cancer survival: do tumors behave differently in elderly women? *J Clin Oncol* **25**, 1705-12 (2007).
6. Dorak, M.T. & Karpuzoglu, E. Gender differences in cancer susceptibility: an inadequately addressed issue. *Front Genet* **3**, 268 (2012).
7. Wang, Y. et al. Effect of luteinizing hormone-induced prohibitin and matrix metalloproteinases on ovarian epithelial tumor cell proliferation. *Am J Cancer Res* **5**, 114-24 (2015).
8. Mertens-Walker, I., Baxter, R.C. & Marsh, D.J. Gonadotropin signalling in epithelial ovarian cancer. *Cancer Lett* **324**, 152-9 (2012).
9. Jacobson, E.M., Hugo, E.R., Borcherdig, D.C. & Ben-Jonathan, N. Prolactin in breast and prostate cancer: molecular and genetic perspectives. *Discov Med* **11**, 315-24 (2011).
10. Hartwell, H.J., Petrosky, K.Y., Fox, J.G., Horseman, N.D. & Rogers, A.B. Prolactin prevents hepatocellular carcinoma by restricting innate immune activation of c-Myc in mice. *Proc Natl Acad Sci U S A* **111**, 11455-60 (2014).
11. Yamamoto, R. et al. Correlation between serum prolactin levels and hepatocellular tumorigenesis induced by 3'-methyl-4-dimethylaminoazobenzene in mice. *Br J Cancer* **72**, 17-21 (1995).
12. Mueller, K.M. et al. Impairment of hepatic growth hormone and glucocorticoid receptor signaling causes steatosis and hepatocellular carcinoma in mice. *Hepatology* **54**, 1398-409 (2011).
13. Gabory, A., Attig, L. & Junien, C. Sexual dimorphism in environmental epigenetic programming. *Mol Cell Endocrinol* **304**, 8-18 (2009).
14. Lea, R.W., Dawson, T., Martinez-Moreno, C.G., El-Abry, N. & Harvey, S. Growth hormone and cancer: GH production and action in glioma? *Gen Comp Endocrinol* **220**, 119-23 (2015).
15. Matsumoto, T. et al. The androgen receptor in health and disease. *Annu Rev Physiol* **75**, 201-24 (2013).
16. Nugent, B.M. et al. Brain feminization requires active repression of masculinization via DNA methylation. *Nat Neurosci* **18**, 690-7 (2015).
17. Fullwood, M.J. et al. An oestrogen-receptor-alpha-bound human chromatin interactome. *Nature* **462**, 58-64 (2009).

18. Mele, M. et al. Human genomics. The human transcriptome across tissues and individuals. *Science* **348**, 660-5 (2015).
19. Brooks, Y.S. et al. Multifactorial ERbeta and NOTCH1 control of squamous differentiation and cancer. *J Clin Invest* **124**, 2260-76 (2014).
20. Sugathan, A. & Waxman, D.J. Genome-wide analysis of chromatin states reveals distinct mechanisms of sex-dependent gene regulation in male and female mouse liver. *Mol Cell Biol* **33**, 3594-610 (2013).
21. Brisken, C. Progesterone signalling in breast cancer: a neglected hormone coming into the limelight. *Nat Rev Cancer* **13**, 385-96 (2013).
22. Skjefstad, K. et al. The prognostic role of progesterone receptor expression in non-small cell lung cancer patients: Gender-related impacts and correlation with disease-specific survival. *Steroids* **98**, 29-36 (2015).
23. Simon, M.S. et al. Estrogen plus progestin and colorectal cancer incidence and mortality. *J Clin Oncol* **30**, 3983-90 (2012).
24. Patrone, C. et al. Regulation of postnatal lung development and homeostasis by estrogen receptor beta. *Mol Cell Biol* **23**, 8542-52 (2003).
25. Wada-Hiraike, O. et al. Role of estrogen receptor beta in colonic epithelium. *Proc Natl Acad Sci U S A* **103**, 2959-64 (2006).
26. Campbell, L. et al. Estrogen promotes cutaneous wound healing via estrogen receptor beta independent of its antiinflammatory activities. *J Exp Med* **207**, 1825-33 (2010).
27. Ashcroft, G.S. & Mills, S.J. Androgen receptor-mediated inhibition of cutaneous wound healing. *J Clin Invest* **110**, 615-24 (2002).
28. Lai, J.J. et al. Monocyte/macrophage androgen receptor suppresses cutaneous wound healing in mice by enhancing local TNF-alpha expression. *J Clin Invest* **119**, 3739-51 (2009).
29. Han, H.J., Heo, J.S. & Lee, Y.J. Estradiol-17beta stimulates proliferation of mouse embryonic stem cells: involvement of MAPKs and CDKs as well as protooncogenes. *Am J Physiol Cell Physiol* **290**, C1067-75 (2006).
30. Masuda, H. et al. Estrogen-mediated endothelial progenitor cell biology and kinetics for physiological postnatal vasculogenesis. *Circ Res* **101**, 598-606 (2007).
31. Nakada, D. et al. Oestrogen increases haematopoietic stem-cell self-renewal in females and during pregnancy. *Nature* **505**, 555-8 (2014).
32. Pawluski, J.L., Brummelte, S., Barha, C.K., Crozier, T.M. & Galea, L.A. Effects of steroid hormones on neurogenesis in the hippocampus of the adult female rodent during the estrous cycle, pregnancy, lactation and aging. *Front Neuroendocrinol* **30**, 343-57 (2009).
33. Bayne, S. et al. Estrogen deficiency leads to telomerase inhibition, telomere shortening and reduced cell proliferation in the adrenal gland of mice. *Cell Res* **18**, 1141-50 (2008).
34. Kato, K. et al. Contribution of estrogen receptor alpha to oncogenic K-Ras-mediated NIH3T3 cell transformation and its implication for escape from senescence by modulating the p53 pathway. *J Biol Chem* **277**, 11217-24 (2002).

35. Bain, J. Andropause. Testosterone replacement therapy for aging men. *Can Fam Physician* **47**, 91-7 (2001).
36. Dokal, I. & Vulliamy, T. Dyskeratosis congenita: its link to telomerase and aplastic anaemia. *Blood Rev* **17**, 217-25 (2003).
37. Calado, R.T. & Young, N.S. Telomere maintenance and human bone marrow failure. *Blood* **111**, 4446-55 (2008).
38. Calado, R.T. et al. Sex hormones, acting on the TERT gene, increase telomerase activity in human primary hematopoietic cells. *Blood* **114**, 2236-43 (2009).
39. Fillmore, C.M. et al. Estrogen expands breast cancer stem-like cells through paracrine FGF/Tbx3 signaling. *Proc Natl Acad Sci U S A* **107**, 21737-42 (2010).
40. Zhang, Y., Eades, G., Yao, Y., Li, Q. & Zhou, Q. Estrogen receptor alpha signaling regulates breast tumor-initiating cells by down-regulating miR-140 which targets the transcription factor SOX2. *J Biol Chem* **287**, 41514-22 (2012).
41. Schroeder, A. et al. Loss of androgen receptor expression promotes a stem-like cell phenotype in prostate cancer through STAT3 signaling. *Cancer Res* **74**, 1227-37 (2014).
42. Huang, C.K., Luo, J., Lee, S.O. & Chang, C. Concise review: androgen receptor differential roles in stem/progenitor cells including prostate, embryonic, stromal, and hematopoietic lineages. *Stem Cells* **32**, 2299-308 (2014).
43. Dotto, G.P. Multifocal epithelial tumors and field cancerization: stroma as a primary determinant. *J Clin Invest* **124**, 1446-53 (2014).
44. Ohlund, D., Elyada, E. & Tuveson, D. Fibroblast heterogeneity in the cancer wound. *J Exp Med* **211**, 1503-23 (2014).
45. Slavin, S. et al. Estrogen receptor alpha in cancer-associated fibroblasts suppresses prostate cancer invasion via modulation of thrombospondin 2 and matrix metalloproteinase 3. *Carcinogenesis* **35**, 1301-9 (2014).
46. Lai, K.P., Yamashita, S., Huang, C.K., Yeh, S. & Chang, C. Loss of stromal androgen receptor leads to suppressed prostate tumorigenesis via modulation of pro-inflammatory cytokines/chemokines. *EMBO Mol Med* **4**, 791-807 (2012).
47. Leach, D.A. et al. Stromal androgen receptor regulates the composition of the microenvironment to influence prostate cancer outcome. *Oncotarget* **6**, 16135-50 (2015).
48. Gupta, P.B. et al. Systemic stromal effects of estrogen promote the growth of estrogen receptor-negative cancers. *Cancer Res* **67**, 2062-71 (2007).
49. Pequeux, C. et al. Stromal estrogen receptor-alpha promotes tumor growth by normalizing an increased angiogenesis. *Cancer Res* **72**, 3010-9 (2012).
50. den Boon, J.A. et al. Molecular transitions from papillomavirus infection to cervical precancer and cancer: Role of stromal estrogen receptor signaling. *Proc Natl Acad Sci U S A* **112**, E3255-64 (2015).
51. Ellem, S.J. & Risbridger, G.P. Treating prostate cancer: a rationale for targeting local oestrogens. *Nat Rev Cancer* **7**, 621-7 (2007).

52. McPherson, S.J. et al. Estrogen receptor-beta activated apoptosis in benign hyperplasia and cancer of the prostate is androgen independent and TNFalpha mediated. *Proc Natl Acad Sci U S A* **107**, 3123-8 (2010).
53. Hamada, H. et al. Estrogen receptors alpha and beta mediate contribution of bone marrow-derived endothelial progenitor cells to functional recovery after myocardial infarction. *Circulation* **114**, 2261-70 (2006).
54. Seo, K.H. et al. Estrogen enhances angiogenesis through a pathway involving platelet-activating factor-mediated nuclear factor-kappaB activation. *Cancer Res* **64**, 6482-8 (2004).
55. Stoner, M. et al. Estrogen regulation of vascular endothelial growth factor gene expression in ZR-75 breast cancer cells through interaction of estrogen receptor alpha and SP proteins. *Oncogene* **23**, 1052-63 (2004).
56. Hartman, J. et al. Estrogen receptor beta inhibits angiogenesis and growth of T47D breast cancer xenografts. *Cancer Res* **66**, 11207-13 (2006).
57. Sieveking, D.P. et al. A sex-specific role for androgens in angiogenesis. *J Exp Med* **207**, 345-52 (2010).
58. Campbell, L. et al. Estrogen receptor-alpha promotes alternative macrophage activation during cutaneous repair. *J Invest Dermatol* **134**, 2447-57 (2014).
59. Yang, W. et al. Estrogen represses hepatocellular carcinoma (HCC) growth via inhibiting alternative activation of tumor-associated macrophages (TAMs). *J Biol Chem* **287**, 40140-9 (2012).
60. Fang, L.Y. et al. Infiltrating macrophages promote prostate tumorigenesis via modulating androgen receptor-mediated CCL4-STAT3 signaling. *Cancer Res* **73**, 5633-46 (2013).
61. Fish, E.N. The X-files in immunity: sex-based differences predispose immune responses. *Nat Rev Immunol* **8**, 737-44 (2008).
62. von Boehmer, H. & Daniel, C. Therapeutic opportunities for manipulating T(Reg) cells in autoimmunity and cancer. *Nat Rev Drug Discov* **12**, 51-63 (2013).
63. Nie, J., Li, Y.Y., Zheng, S.G., Tsun, A. & Li, B. FOXP3(+) Treg Cells and Gender Bias in Autoimmune Diseases. *Front Immunol* **6**, 493 (2015).
64. Polanczyk, M.J. et al. Cutting edge: estrogen drives expansion of the CD4+CD25+ regulatory T cell compartment. *J Immunol* **173**, 2227-30 (2004).
65. Walecki, M. et al. Androgen receptor modulates Foxp3 expression in CD4+CD25+Foxp3+ regulatory T-cells. *Mol Biol Cell* **26**, 2845-57 (2015).
66. Yang, X.O. et al. Molecular antagonism and plasticity of regulatory and inflammatory T cell programs. *Immunity* **29**, 44-56 (2008).
67. Isse, K. et al. Estrogen stimulates female biliary epithelial cell interleukin-6 expression in mice and humans. *Hepatology* **51**, 869-80 (2010).
68. Olivieri, F. et al. The -174 C/G locus affects in vitro/in vivo IL-6 production during aging. *Exp Gerontol* **37**, 309-14 (2002).
69. Yurkovetskiy, L. et al. Gender bias in autoimmunity is influenced by microbiota. *Immunity* **39**, 400-12 (2013).
70. Bianchi, I., Lleo, A., Gershwin, M.E. & Invernizzi, P. The X chromosome and immune associated genes. *J Autoimmun* **38**, J187-92 (2012).

71. Dai, R. & Ahmed, S.A. Sexual dimorphism of miRNA expression: a new perspective in understanding the sex bias of autoimmune diseases. *Ther Clin Risk Manag* **10**, 151-63 (2014).
72. Hewagama, A. et al. Overexpression of X-linked genes in T cells from women with lupus. *J Autoimmun* **41**, 60-71 (2013).
73. Barros, R.P. & Gustafsson, J.A. Estrogen receptors and the metabolic network. *Cell Metab* **14**, 289-99 (2011).
74. Mauvais-Jarvis, F. Estrogen and androgen receptors: regulators of fuel homeostasis and emerging targets for diabetes and obesity. *Trends Endocrinol Metab* **22**, 24-33 (2011).
75. Calle, E.E. & Kaaks, R. Overweight, obesity and cancer: epidemiological evidence and proposed mechanisms. *Nat Rev Cancer* **4**, 579-91 (2004).
76. Tisdale, M.J. Cachexia in cancer patients. *Nat Rev Cancer* **2**, 862-71 (2002).
77. Baracos, V.E., Reiman, T., Mourtzakis, M., Gioulbasanis, I. & Antoun, S. Body composition in patients with non-small cell lung cancer: a contemporary view of cancer cachexia with the use of computed tomography image analysis. *Am J Clin Nutr* **91**, 1133S-1137S (2010).
78. Cospers, P.F. & Leinwand, L.A. Cancer causes cardiac atrophy and autophagy in a sexually dimorphic manner. *Cancer Res* **71**, 1710-20 (2011).
79. von Haehling, S., Morley, J.E. & Anker, S.D. An overview of sarcopenia: facts and numbers on prevalence and clinical impact. *J Cachexia Sarcopenia Muscle* **1**, 129-133 (2010).
80. Vigano, A. et al. Male hypogonadism associated with advanced cancer: a systematic review. *Lancet Oncol* **11**, 679-84 (2010).
81. Basaria, S. Male hypogonadism. *Lancet* **383**, 1250-63 (2014).
82. Dobs, A.S. et al. Effects of enobosarm on muscle wasting and physical function in patients with cancer: a double-blind, randomised controlled phase 2 trial. *Lancet Oncol* **14**, 335-45 (2013).
83. Li, Z., Tuteja, G., Schug, J. & Kaestner, K.H. Foxa1 and Foxa2 are essential for sexual dimorphism in liver cancer. *Cell* **148**, 72-83 (2012).
84. Naugler, W.E. et al. Gender disparity in liver cancer due to sex differences in MyD88-dependent IL-6 production. *Science* **317**, 121-4 (2007).
85. Matic, M. et al. Estrogen signalling and the metabolic syndrome: targeting the hepatic estrogen receptor alpha action. *PLoS One* **8**, e57458 (2013).
86. Yamamoto, R., Tatsuta, M. & Terada, N. Suppression by oestrogen of hepatocellular tumourigenesis induced in mice by 3'-methyl-4-dimethylaminoazobenzene. *Br J Cancer* **68**, 303-7 (1993).
87. Porsch Hallstrom, I., Svensson, D. & Blanck, A. Sex-differentiated deoxycholic acid promotion of rat liver carcinogenesis is under pituitary control. *Carcinogenesis* **12**, 2035-40 (1991).
88. Kerrigan, J.R. & Rogol, A.D. The impact of gonadal steroid hormone action on growth hormone secretion during childhood and adolescence. *Endocr Rev* **13**, 281-98 (1992).
89. Leung, K.C. et al. Estrogen inhibits GH signaling by suppressing GH-induced JAK2 phosphorylation, an effect mediated by SOCS-2. *Proc Natl Acad Sci U S A* **100**, 1016-21 (2003).

90. Mode, A. & Gustafsson, J.A. Sex and the liver - a journey through five decades. *Drug Metab Rev* **38**, 197-207 (2006).
91. Passarelli, M.N. et al. Common single-nucleotide polymorphisms in the estrogen receptor beta promoter are associated with colorectal cancer survival in postmenopausal women. *Cancer Res* **73**, 767-75 (2013).
92. Press, O.A. et al. Gender-related survival differences associated with EGFR polymorphisms in metastatic colon cancer. *Cancer Res* **68**, 3037-42 (2008).
93. Schmidt, A.N., Nanney, L.B., Boyd, A.S., King, L.E., Jr. & Ellis, D.L. Oestrogen receptor-beta expression in melanocytic lesions. *Exp Dermatol* **15**, 971-80 (2006).
94. Matsuoka, H. et al. Tamoxifen inhibits tumor cell invasion and metastasis in mouse melanoma through suppression of PKC/MEK/ERK and PKC/PI3K/Akt pathways. *Exp Cell Res* **315**, 2022-32 (2009).
95. Hartman, J. et al. Tumor repressive functions of estrogen receptor beta in SW480 colon cancer cells. *Cancer Res* **69**, 6100-6 (2009).
96. Sareddy, G.R. et al. Therapeutic significance of estrogen receptor beta agonists in gliomas. *Mol Cancer Ther* **11**, 1174-82 (2012).
97. Pinton, G. et al. Estrogen receptor-beta affects the prognosis of human malignant mesothelioma. *Cancer Res* **69**, 4598-604 (2009).
98. Yu, C.P. et al. Estrogen inhibits renal cell carcinoma cell progression through estrogen receptor-beta activation. *PLoS One* **8**, e56667 (2013).
99. Yakimchuk, K. et al. Effect of ligand-activated estrogen receptor beta on lymphoma growth in vitro and in vivo. *Leukemia* **25**, 1103-10 (2011).
100. Stanley, J.A. et al. Androgen receptor expression in human thyroid cancer tissues: a potential mechanism underlying the gender bias in the incidence of thyroid cancers. *J Steroid Biochem Mol Biol* **130**, 105-24 (2012).
101. Lee, M.L. et al. Induction of thyroid papillary carcinoma cell proliferation by estrogen is associated with an altered expression of Bcl-xL. *Cancer J* **11**, 113-21 (2005).
102. Sarma, K. et al. ATRX directs binding of PRC2 to Xist RNA and Polycomb targets. *Cell* **159**, 869-83 (2014).
103. Minajigi, A. et al. Chromosomes. A comprehensive Xist interactome reveals cohesin repulsion and an RNA-directed chromosome conformation. *Science* **349** (2015).
104. Migeon, B.R. The role of X inactivation and cellular mosaicism in women's health and sex-specific diseases. *JAMA* **295**, 1428-33 (2006).
105. Kawakami, T. et al. Characterization of loss-of-inactive X in Klinefelter syndrome and female-derived cancer cells. *Oncogene* **23**, 6163-9 (2004).
106. Benoit, M.H. et al. Global analysis of chromosome X gene expression in primary cultures of normal ovarian surface epithelial cells and epithelial ovarian cancer cell lines. *Int J Oncol* **30**, 5-17 (2007).
107. Yildirim, E. et al. Xist RNA is a potent suppressor of hematologic cancer in mice. *Cell* **152**, 727-42 (2013).
108. Yao, Y. et al. Knockdown of long non-coding RNA XIST exerts tumor-suppressive functions in human glioblastoma stem cells by up-regulating miR-152. *Cancer Lett* **359**, 75-86 (2015).

109. Kawakami, T. et al. The roles of supernumerical X chromosomes and XIST expression in testicular germ cell tumors. *J Urol* **169**, 1546-52 (2003).
110. Bellott, D.W. et al. Mammalian Y chromosomes retain widely expressed dosage-sensitive regulators. *Nature* **508**, 494-9 (2014).
111. Walport, L.J. et al. Human UTY(KDM6C) is a male-specific N-methyl lysyl demethylase. *J Biol Chem* **289**, 18302-13 (2014).
112. Mar, B.G. et al. Sequencing histone-modifying enzymes identifies UTX mutations in acute lymphoblastic leukemia. *Leukemia* **26**, 1881-3 (2012).
113. Ntziachristos, P. et al. Contrasting roles of histone 3 lysine 27 demethylases in acute lymphoblastic leukaemia. *Nature* **514**, 513-7 (2014).
114. Van der Meulen, J. et al. The H3K27me3 demethylase UTX is a gender-specific tumor suppressor in T-cell acute lymphoblastic leukemia. *Blood* **125**, 13-21 (2015).
115. Van Vlierberghe, P. et al. PHF6 mutations in T-cell acute lymphoblastic leukemia. *Nat Genet* **42**, 338-42 (2010).
116. De Keersmaecker, K. et al. Exome sequencing identifies mutation in CNOT3 and ribosomal genes RPL5 and RPL10 in T-cell acute lymphoblastic leukemia. *Nat Genet* **45**, 186-90 (2013).
117. van Haften, G. et al. Somatic mutations of the histone H3K27 demethylase gene UTX in human cancer. *Nat Genet* **41**, 521-3 (2009).
118. Dalgliesh, G.L. et al. Systematic sequencing of renal carcinoma reveals inactivation of histone modifying genes. *Nature* **463**, 360-3 (2010).
119. Northcott, P.A. et al. Medulloblastomics: the end of the beginning. *Nat Rev Cancer* **12**, 818-34 (2012).
120. Robinson, G. et al. Novel mutations target distinct subgroups of medulloblastoma. *Nature* **488**, 43-8 (2012).
121. Jiang, L. et al. Exome sequencing identifies somatic mutations of DDX3X in natural killer/T-cell lymphoma. *Nat Genet* **47**, 1061-6 (2015).
122. Wu, D.W. et al. DDX3 loss by p53 inactivation promotes tumor malignancy via the MDM2/Slug/E-cadherin pathway and poor patient outcome in non-small-cell lung cancer. *Oncogene* **33**, 1515-26 (2014).
123. Huff, V. Wilms' tumours: about tumour suppressor genes, an oncogene and a chameleon gene. *Nat Rev Cancer* **11**, 111-21 (2011).
124. Rondinelli, B. et al. Histone demethylase JARID1C inactivation triggers genomic instability in sporadic renal cancer. *J Clin Invest* (2015).
125. Ji, X. et al. Lysine-specific demethylase 5C promotes hepatocellular carcinoma cell invasion through inhibition BMP7 expression. *BMC Cancer* **15**, 801 (2015).
126. Karanikas, V. et al. Foxp3 expression in human cancer cells. *J Transl Med* **6**, 19 (2008).
127. Hinz, S. et al. Foxp3 expression in pancreatic carcinoma cells as a novel mechanism of immune evasion in cancer. *Cancer Res* **67**, 8344-50 (2007).
128. Liu, R. et al. FOXP3 Controls an miR-146/NF-kappaB Negative Feedback Loop That Inhibits Apoptosis in Breast Cancer Cells. *Cancer Res* **75**, 1703-13 (2015).

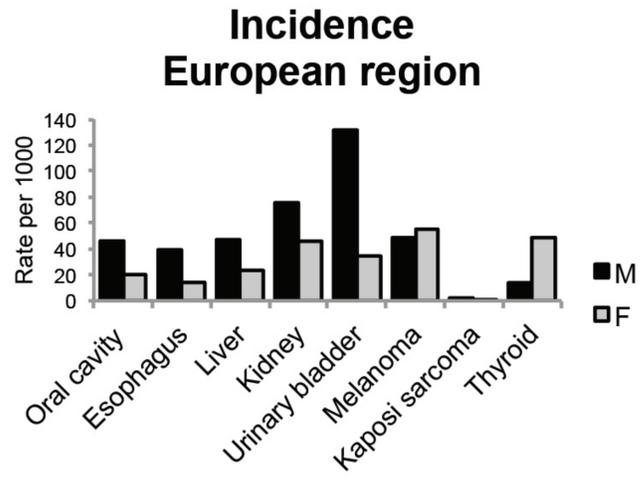
129. le Sage, C. et al. Regulation of the p27(Kip1) tumor suppressor by miR-221 and miR-222 promotes cancer cell proliferation. *EMBO J* **26**, 3699-708 (2007).
130. Li, X. et al. miRNA-223 promotes gastric cancer invasion and metastasis by targeting tumor suppressor EPB41L3. *Mol Cancer Res* **9**, 824-33 (2011).
131. Hui, A.B. et al. Potentially prognostic miRNAs in HPV-associated oropharyngeal carcinoma. *Clin Cancer Res* **19**, 2154-62 (2013).
132. Singhal, R., Bard, J.E., Nowak, N.J., Buck, M.J. & Kandel, E.S. FOXO1 regulates expression of a microRNA cluster on X chromosome. *Aging (Albany NY)* **5**, 347-56 (2013).
133. Nadal, M. et al. Aneuploidy of chromosome Y in prostate tumors and seminal vesicles: a possible sign of aging rather than an indicator of carcinogenesis? *Mol Carcinog* **46**, 543-52 (2007).
134. Konig, J.J., Teubel, W., Romijn, J.C., Schroder, F.H. & Hagemeyer, A. Gain and loss of chromosomes 1, 7, 8, 10, 18, and Y in 46 prostate cancers. *Hum Pathol* **27**, 720-7 (1996).
135. Stahl, P.R. et al. Y chromosome losses are exceedingly rare in prostate cancer and unrelated to patient age. *Prostate* **72**, 898-903 (2012).
136. Kowalski, J. et al. Chromosomal abnormalities of adenocarcinoma of the pancreas: identifying early and late changes. *Cancer Genet Cytogenet* **178**, 26-35 (2007).
137. Bottarelli, L. et al. Sex chromosome alterations associate with tumor progression in sporadic colorectal carcinomas. *Clin Cancer Res* **13**, 4365-70 (2007).
138. Fadl-Elmula, I. et al. Karyotypic characterization of urinary bladder transitional cell carcinomas. *Genes Chromosomes Cancer* **29**, 256-65 (2000).
139. Mitelman F, J.B.a.M.F. (2015).
140. Forsberg, L.A. et al. Mosaic loss of chromosome Y in peripheral blood is associated with shorter survival and higher risk of cancer. *Nat Genet* **46**, 624-8 (2014).
141. Dumanski, J.P. et al. Mutagenesis. Smoking is associated with mosaic loss of chromosome Y. *Science* **347**, 81-3 (2015).
142. Bianchi, N.O. Y chromosome structural and functional changes in human malignant diseases. *Mutat Res* **682**, 21-7 (2009).
143. Kido, T. & Lau, Y.F. Roles of the Y chromosome genes in human cancers. *Asian J Androl* **17**, 373-80 (2015).
144. Murakami, S. et al. SRY and OCT4 Are Required for the Acquisition of Cancer Stem Cell-Like Properties and Are Potential Differentiation Therapy Targets. *Stem Cells* **33**, 2652-63 (2015).
145. Murakami, S. et al. The male-specific factor Sry harbors an oncogenic function. *Oncogene* **33**, 2978-86 (2014).
146. Collignon, J. et al. A comparison of the properties of Sox-3 with Sry and two related genes, Sox-1 and Sox-2. *Development* **122**, 509-20 (1996).
147. Boumahdi, S. et al. SOX2 controls tumour initiation and cancer stem-cell functions in squamous-cell carcinoma. *Nature* **511**, 246-50 (2014).

148. Yuan, X., Lu, M.L., Li, T. & Balk, S.P. SRY interacts with and negatively regulates androgen receptor transcriptional activity. *J Biol Chem* **276**, 46647-54 (2001).
149. Lau, Y.F., Li, Y. & Kido, T. Gonadoblastoma locus and the TSPY gene on the human Y chromosome. *Birth Defects Res C Embryo Today* **87**, 114-22 (2009).
150. Gallagher, W.M. et al. Multiple markers for melanoma progression regulated by DNA methylation: insights from transcriptomic studies. *Carcinogenesis* **26**, 1856-67 (2005).
151. Kido, T., Hatakeyama, S., Ohyama, C. & Lau, Y.F. Expression of the Y-Encoded TSPY is Associated with Progression of Prostate Cancer. *Genes (Basel)* **1**, 283-93 (2010).
152. Kido, T. et al. The potential contributions of a Y-located protooncogene and its X homologue in sexual dimorphisms in hepatocellular carcinoma. *Hum Pathol* **45**, 1847-58 (2014).
153. Delbridge, M.L. et al. TSPY, the candidate gonadoblastoma gene on the human Y chromosome, has a widely expressed homologue on the X - implications for Y chromosome evolution. *Chromosome Res* **12**, 345-56 (2004).
154. Kido, T., Ou, J.H. & Lau, Y.F. The X-linked tumor suppressor TSPX interacts and promotes degradation of the hepatitis B viral protein HBx via the proteasome pathway. *PLoS One* **6**, e22979 (2011).
155. Li, S. et al. Over-expressed Testis-specific Protein Y-encoded 1 as a novel biomarker for male hepatocellular carcinoma. *PLoS One* **9**, e89219 (2014).
156. Lau, Y.F. & Zhang, J. Expression analysis of thirty one Y chromosome genes in human prostate cancer. *Mol Carcinog* **27**, 308-21 (2000).
157. Tsuei, D.J. et al. RBMY, a male germ cell-specific RNA-binding protein, activated in human liver cancers and transforms rodent fibroblasts. *Oncogene* **23**, 5815-22 (2004).
158. Tsuei, D.J. et al. Male germ cell-specific RNA binding protein RBMY: a new oncogene explaining male predominance in liver cancer. *PLoS One* **6**, e26948 (2011).
159. Potter, J.D. Morphogens, morphostats, microarchitecture and malignancy. *Nat Rev Cancer* **7**, 464-74 (2007).
160. Tomasetti, C. & Vogelstein, B. Cancer etiology. Variation in cancer risk among tissues can be explained by the number of stem cell divisions. *Science* **347**, 78-81 (2015).
161. Sottoriva, A. et al. A Big Bang model of human colorectal tumor growth. *Nat Genet* **47**, 209-16 (2015).
162. Martincorena, I. et al. Tumor evolution. High burden and pervasive positive selection of somatic mutations in normal human skin. *Science* **348**, 880-6 (2015).
163. Cardiff, R.D. & Borowsky, A.D. Precancer: sequentially acquired or predetermined? *Toxicol Pathol* **38**, 171-9 (2010).
164. Gatenby, R. Perspective: Finding cancer's first principles. *Nature* **491**, S55 (2012).
165. Merlo, L.M., Pepper, J.W., Reid, B.J. & Maley, C.C. Cancer as an evolutionary and ecological process. *Nat Rev Cancer* **6**, 924-35 (2006).

166. Sharma, P. & Allison, J.P. The future of immune checkpoint therapy. *Science* **348**, 56-61 (2015).
167. Austad, S.N. Why women live longer than men: sex differences in longevity. *Genet Med* **3**, 79-92 (2006).
168. Paterni, I., Granchi, C., Katzenellenbogen, J.A. & Minutolo, F. Estrogen receptors alpha (ERalpha) and beta (ERbeta): subtype-selective ligands and clinical potential. *Steroids* **90**, 13-29 (2014).
169. Taplin, M.E. Drug insight: role of the androgen receptor in the development and progression of prostate cancer. *Nat Clin Pract Oncol* **4**, 236-44 (2007).
170. Ahuja, N., Sharma, A.R. & Baylin, S.B. Epigenetic Therapeutics: A New Weapon in the War Against Cancer. *Annu Rev Med* **67**, 73-89 (2016).

Figure 1

A



B

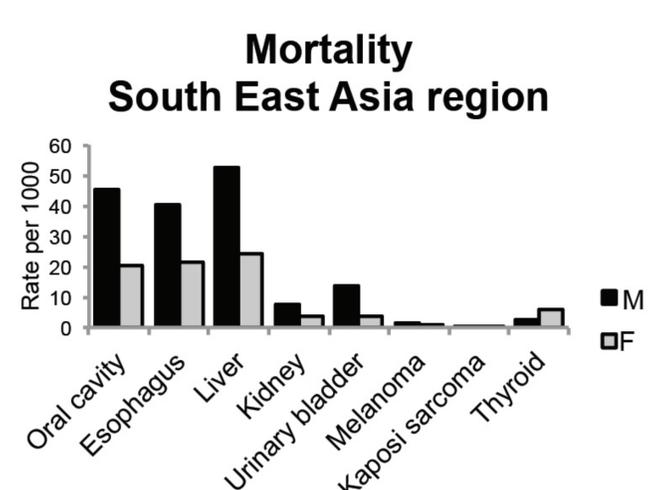
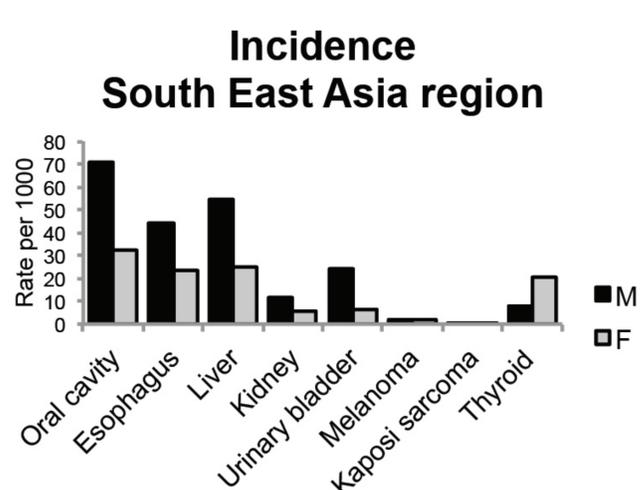
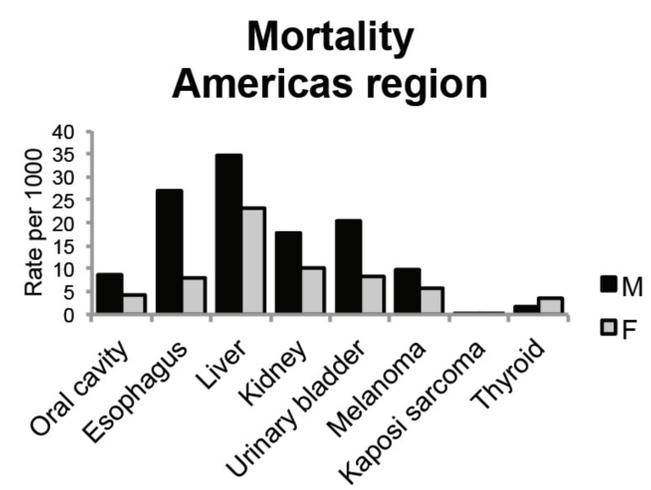
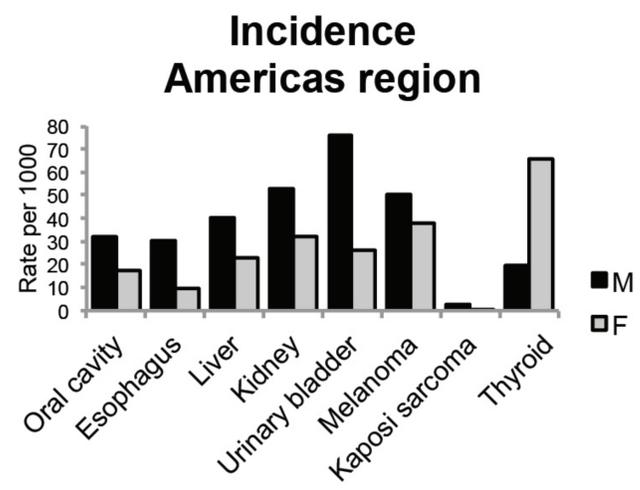
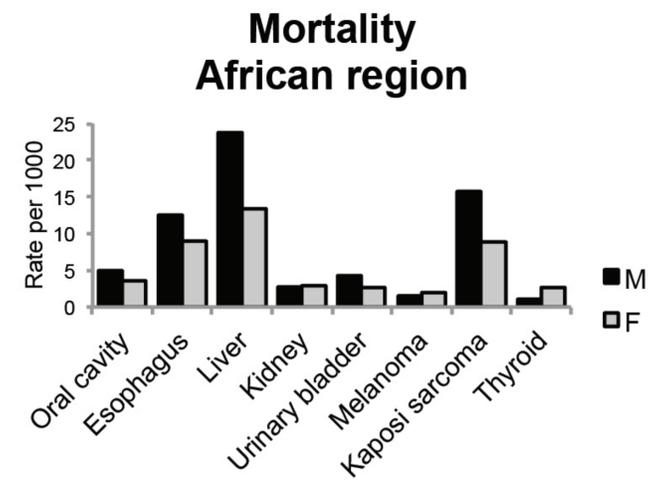
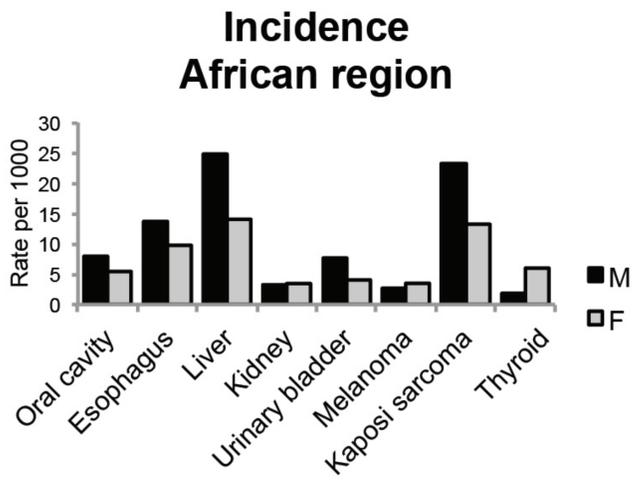
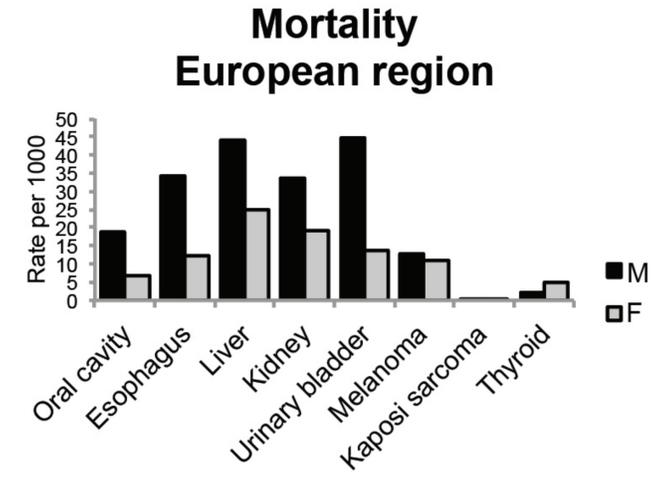


Figure 2

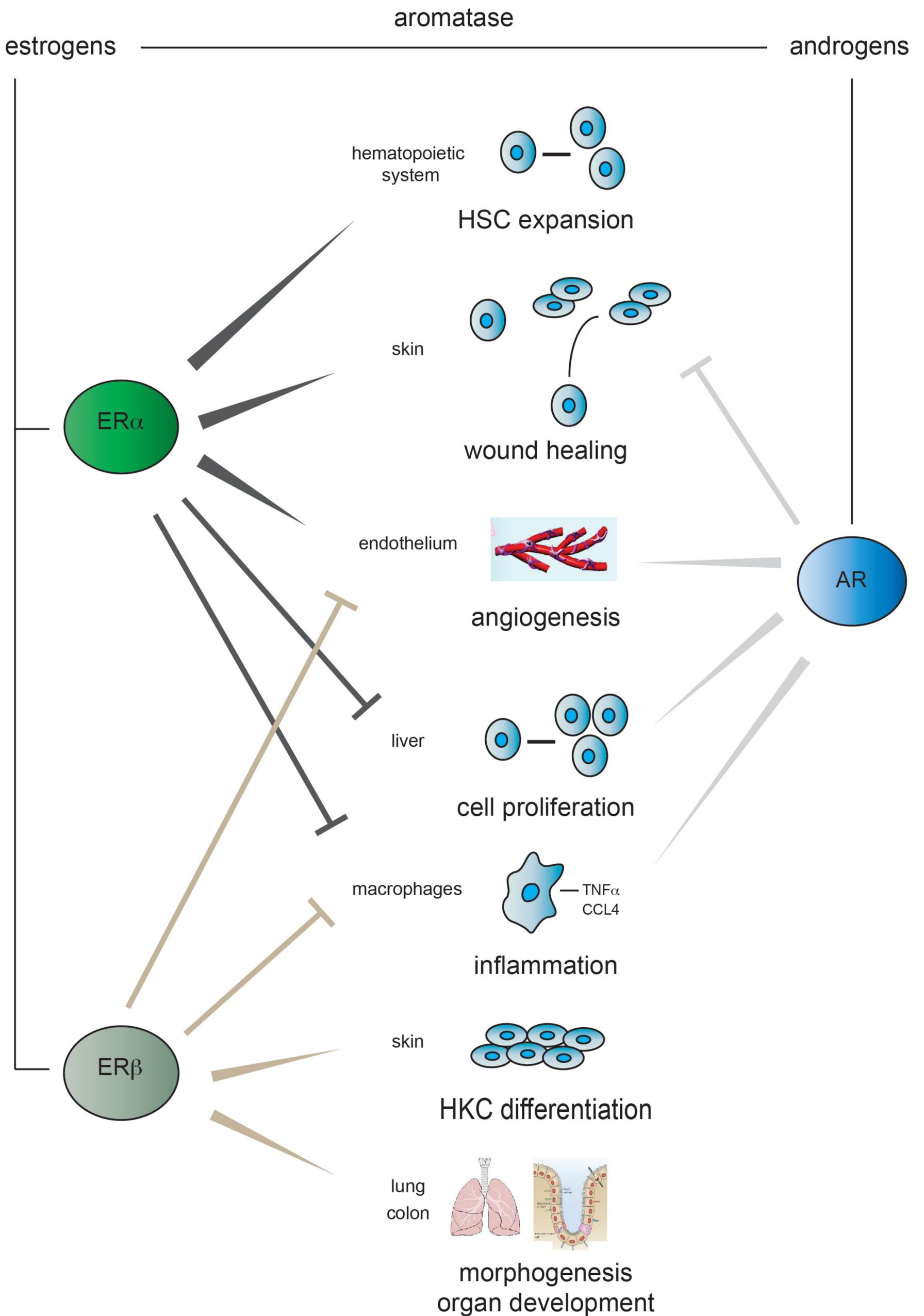


Figure 3

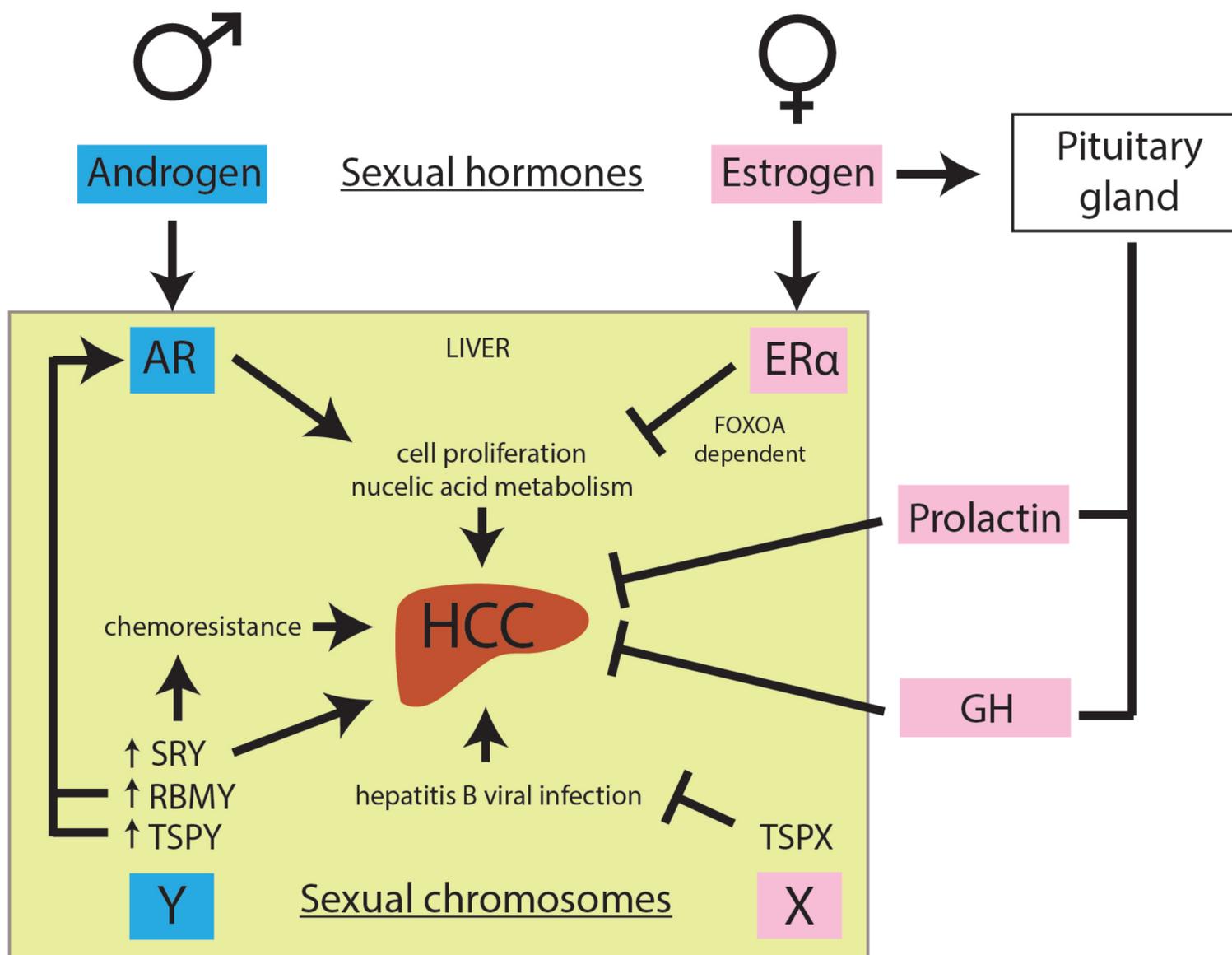


Figure 4

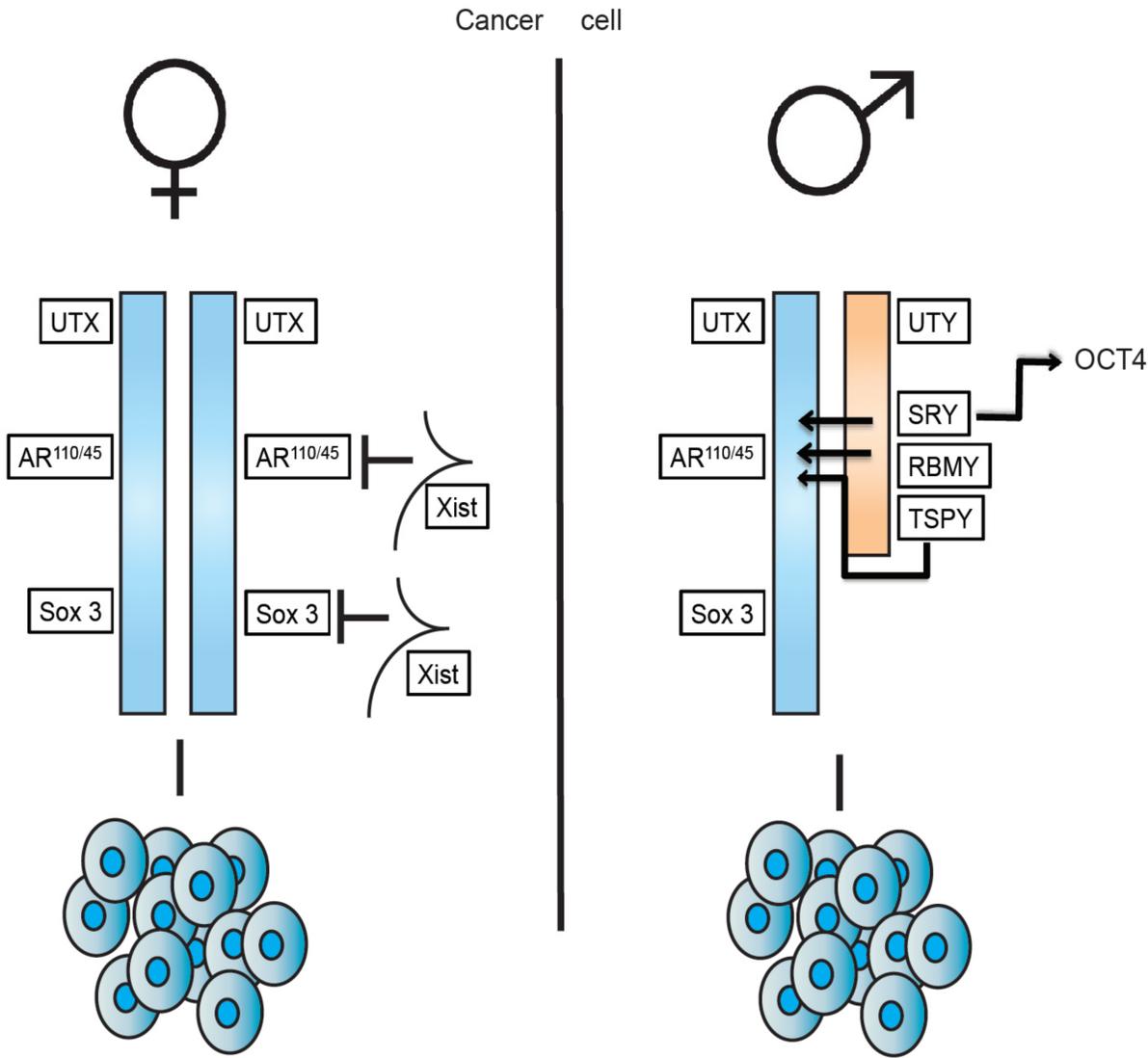


Figure 5

