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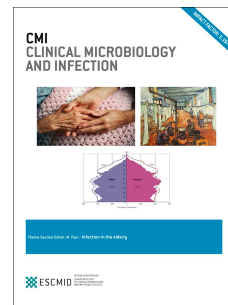
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SARS-CoV-2 ACE-Receptor detection in the placenta throughout pregnancy

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1 SARS-CoV-2 ACE-Receptor detection in the placenta throughout pregnancy

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21 To the Editor,

22 Fetal risks for women exposed to SARS-CoV-2 and subsequent pregnancy outcomes remain
23 uncertain (1). Recent reports have raised controversial concerns regarding the potential for
24 vertical transmission by transplacental infection. It has been proposed that the occurrence of
25 intrauterine transplacental SARS-CoV-2 among infected mother-infant dyads be based upon
26 identification of SARS-CoV-2 in chorionic villous cells using immunohistochemistry or
27 nucleic acid methods such as *in situ* hybridization (2). Hypothetically, two conditions are
28 necessary for transplacental transmission to be possible: 1) the virus must reach the placenta;
29 and 2) the receptor for the virus, angiotensin-converting enzyme 2 (ACE2), must be present in
30 the placenta.

31 Regarding the first condition, we and others recently reported RT-PCR or RNAscope data
32 supporting the presence of SARS-CoV-2 in placental tissue (3-6). To date, conflicting data
33 exists regarding the second condition (4, 7, 8).

34 In order to investigate ACE2 expression in the placenta throughout pregnancy, we selected
35 formalin-fixed placental tissues between 14 and 40 weeks gestation from 28 patients who
36 delivered before the current COVID-19 outbreak (2 different cases for each 2 weeks gestation
37 – i.e. 14, 16, 18, ... 38, 40 weeks, annex), as well as placental tissues at 19 weeks gestation
38 from a COVID-19 positive patient (figure 1). Kidney was used as positive (brush border of
39 proximal renal tubules and podocytes of glomeruli) and negative (distal renal tubules and
40 renal medulla) controls. Using a monoclonal anti-ACE2 antibody (Atlas antibodies, clone
41 CL4035, dilution 1/1000), we demonstrated *in situ* expression of ACE2 at the maternal-fetal
42 interface (✓ 1 and annex), a prerequisite for transplacental transmission. We observed a strong
43 and diffuse membranous staining of cytotrophoblast and syncytiotrophoblast cells of placental
44 villi, as well as a membranous expression in extra-villous trophoblast. By testing placental
45 tissues at various gestational ages in both COVID-19 positive and negative mothers, we

46 confirmed that ACE expression is present consistently throughout pregnancy regardless of
47 COVID-19 status.

48 Our *in situ* analysis by specific immunohistochemistry and SARS-CoV-2 detection by RT-
49 PCR indicate a possible placental infection by SARS-CoV2. Trophoblastic cells, which are in
50 direct contact with the maternal blood in the intervillous space, show strong expression of
51 ACE2 throughout pregnancy, supporting that SARS-CoV2 is able to infect the placenta via a
52 receptor-mediated mechanism. SARS-CoV-2 can cross the placental barrier, as it has been
53 demonstrated in a neonate born to a mother infected in the last trimester and presenting with
54 neurological compromise (5). However, the rate of vertical transmission to the fetus warrants
55 further investigation. By analogy to other pathogens (i.e. cytomegalovirus or toxoplasmosis),
56 this may occur once sufficient time has elapsed for the virus to breach the placental barrier
57 (6–8 weeks after infection).

58 SARS-CoV-2 infection of the maternal placental surface may induce acute or chronic
59 placental insufficiency (3-5), or be responsible for subsequent miscarriage or fetal growth
60 restriction as reported in 40% of MERS or SARS-1 maternal infections. In consequences, this
61 findings could lead to increase protective measures for pregnant women, as well as a
62 heightened level of concern/monitoring of pregnancies exposed to SARS-CoV-2.

63 **Author Contributions:**

64 Prof Baud and Dr Gengler had full access to all of the data in the study and takes
65 responsibility for the integrity of the data and the accuracy of the data analysis.

66 *Conceptualization:* Gengler, Dubruc, Baud

67 *Acquisition, analysis, or interpretation of data:* All authors

68 *Writing – Original Draft:* Gengler, Dubruc, Baud

69 *Writing – Review & Editing:* Gengler, Greub, Favre, de Laval, Baud

70 *Administrative, technical, or material support:* All authors

71 *Supervision and Validation:* Gengler, Baud

72

73 **Conflict of Interest Disclosures:**

74 Dr Greub reported having ongoing research agreements with Resistell and Becton-Dickinson
75 and being medical advisor for Resistell; he also reported being developer of a card game on
76 microbes and funding JeuPro, a start-up company that distributes the game Krobs. No other
77 disclosures were reported.

78

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81

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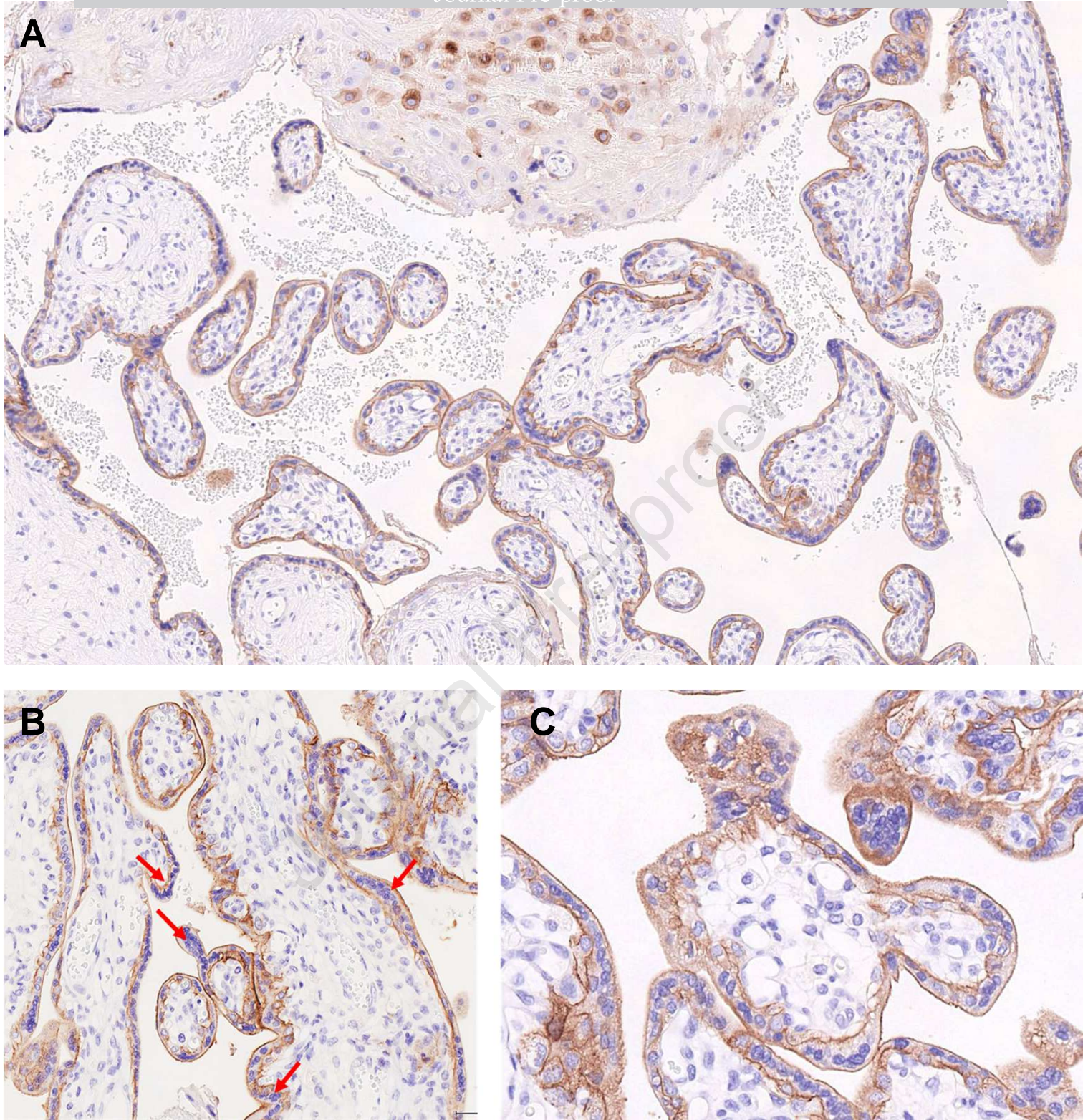
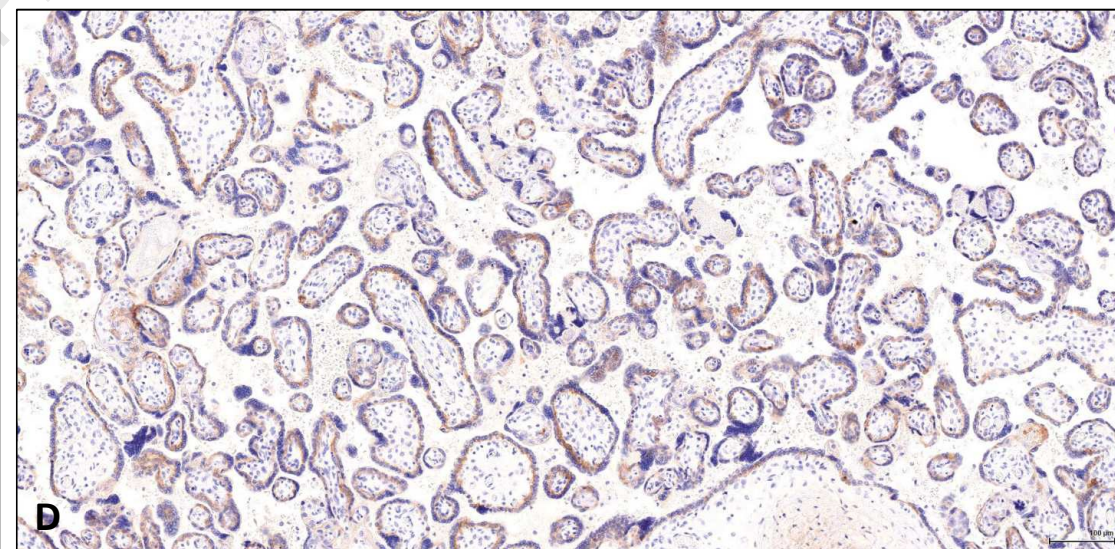
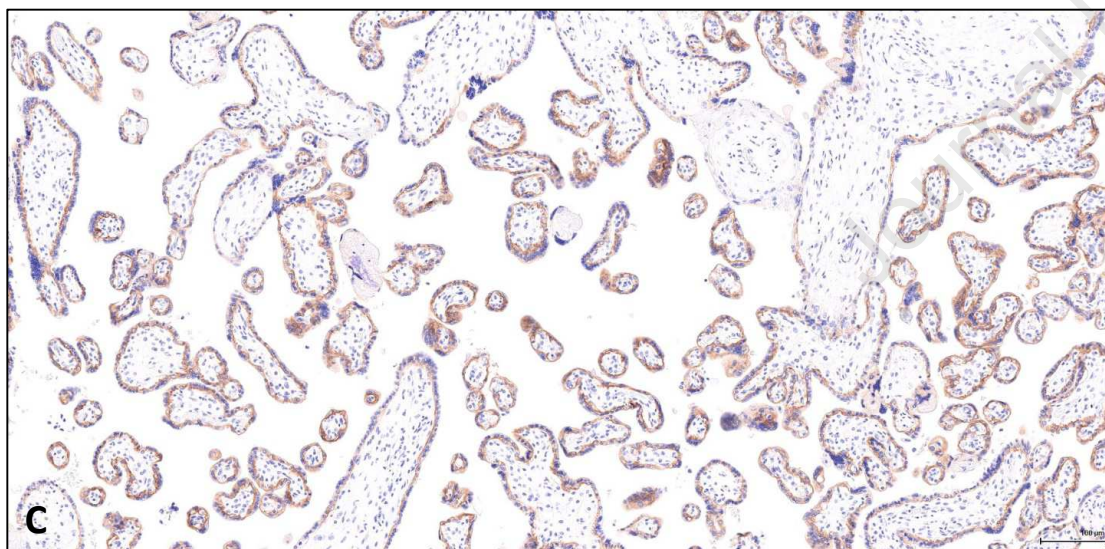
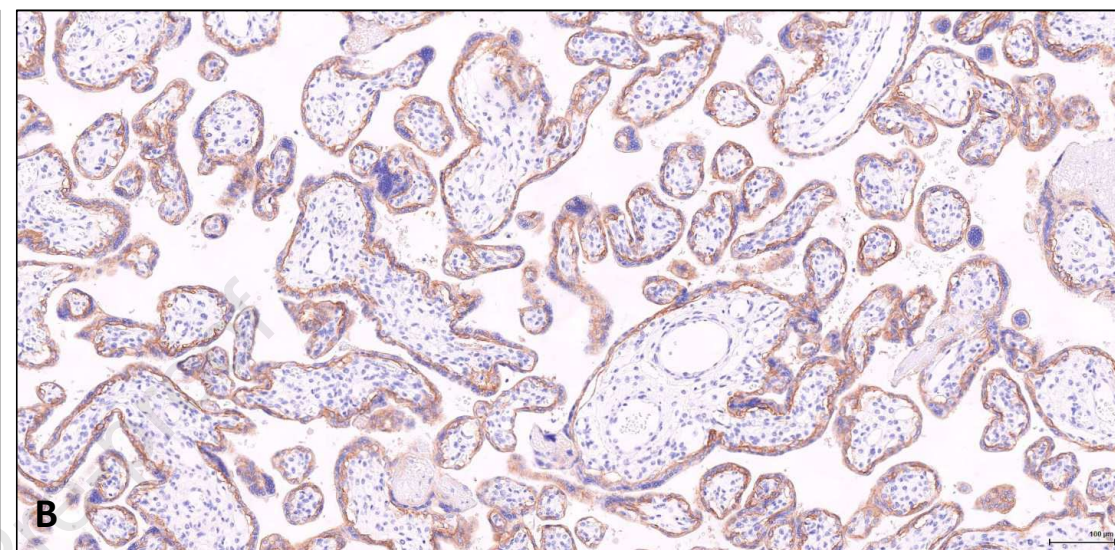
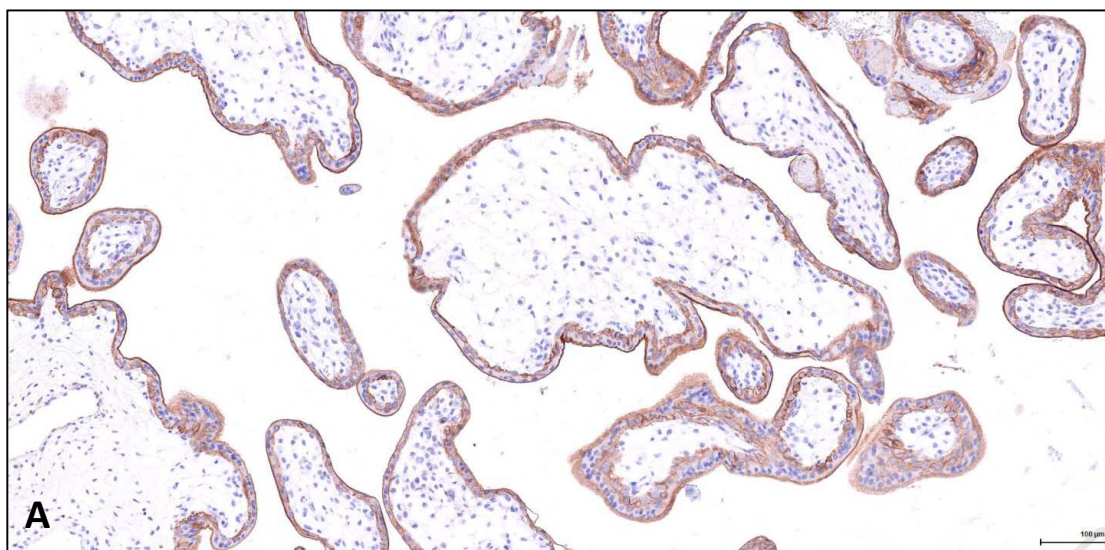


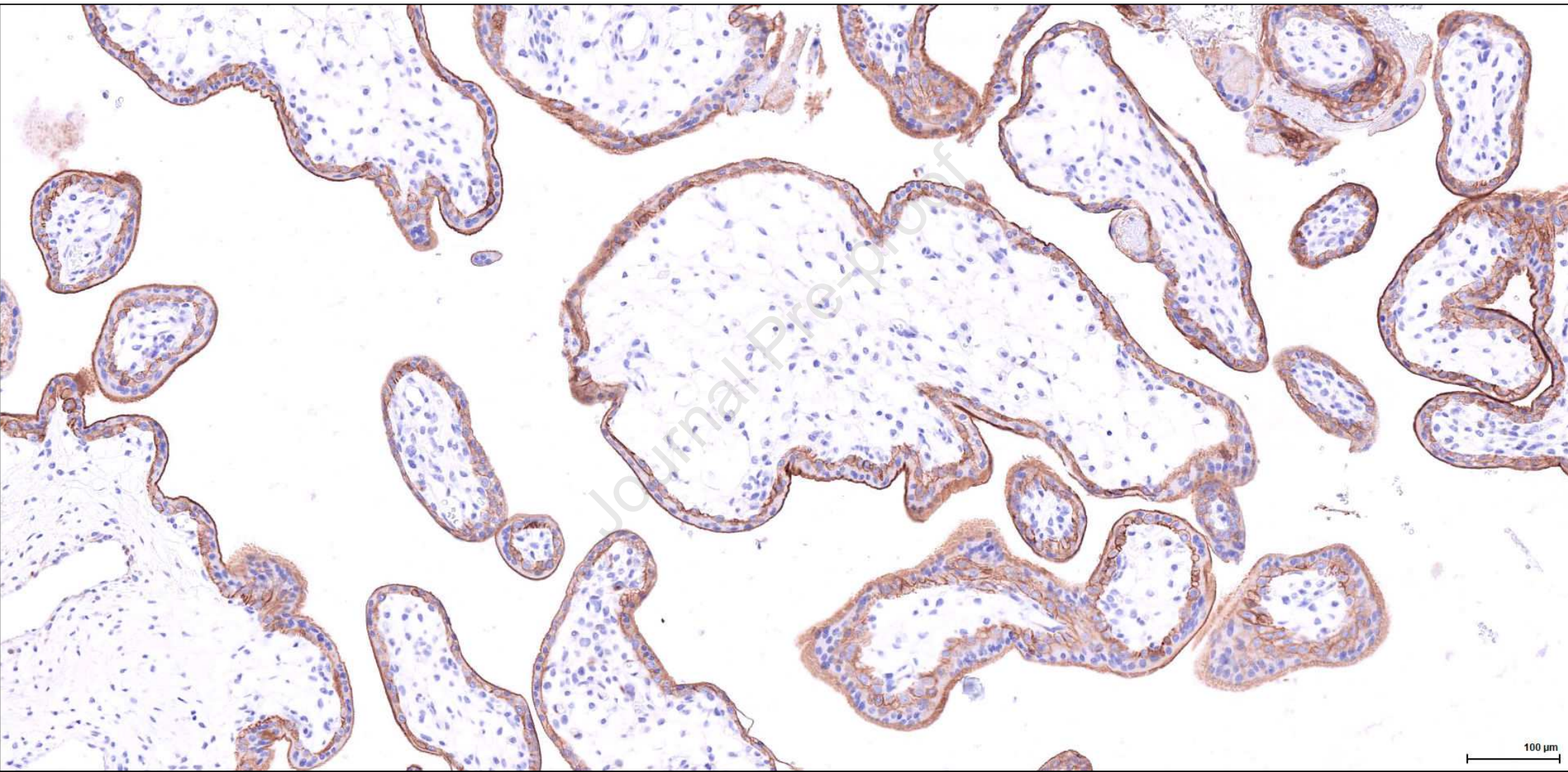
Figure 1:

Diffuse membranous staining of villous cytotrophoblast and syncytiotrophoblast cells (arrows) with monoclonal Anti-ACE2 antibody (clone CL4035), dilution 1/1000 in a COVID-19 positive mother, 19 weeks of amenorrhea.

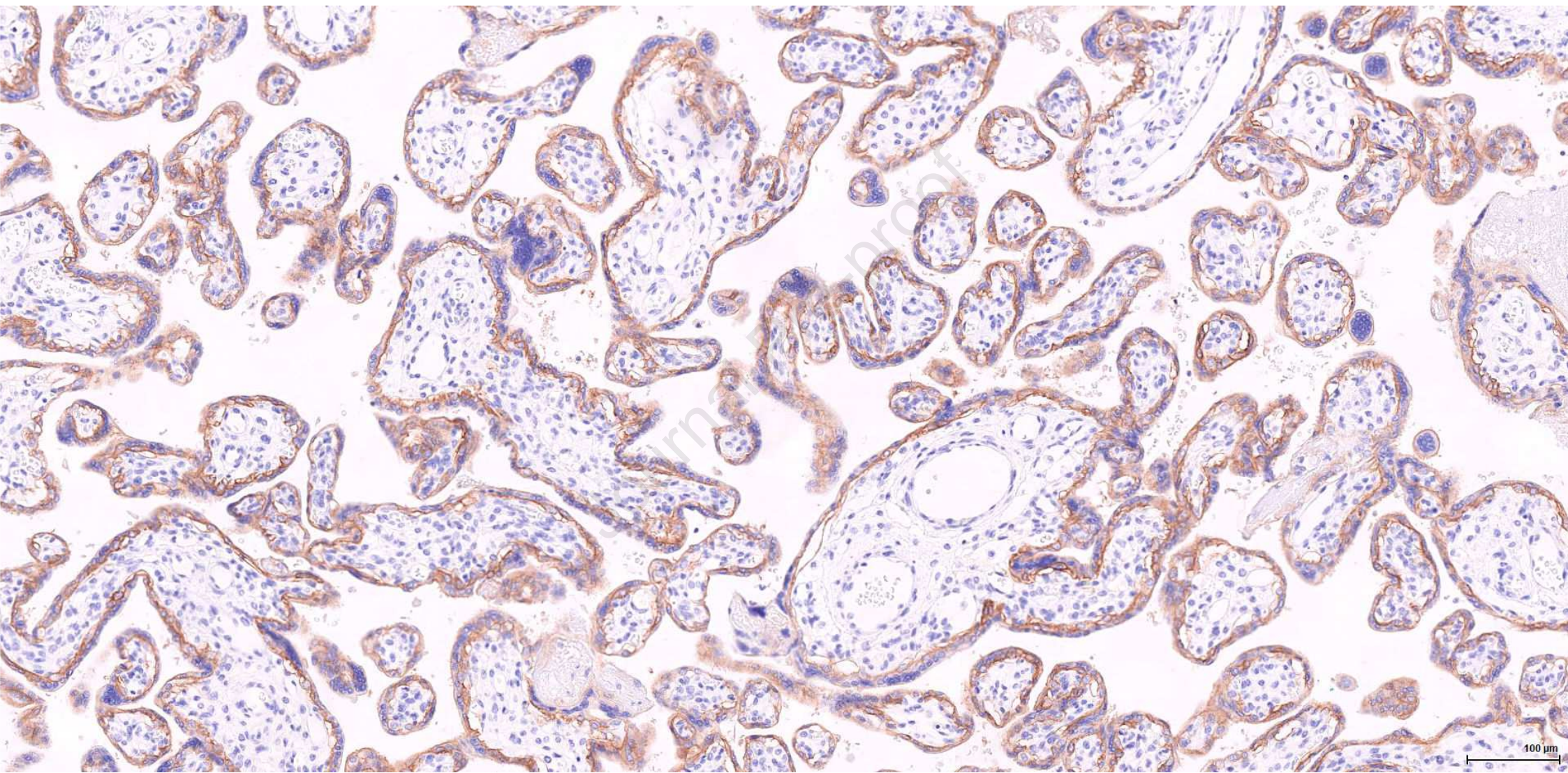


Annex: Diffuse membranous staining of villous cytotrophoblast cells with monoclonal Anti-ACE2 antibody (clone CL4035), dilution 1/1000, 10X at various gestational ages (16 (A), 22 (B), 31 (C) and 40 (D) weeks gestation) in control COVID-19 negative mothers.

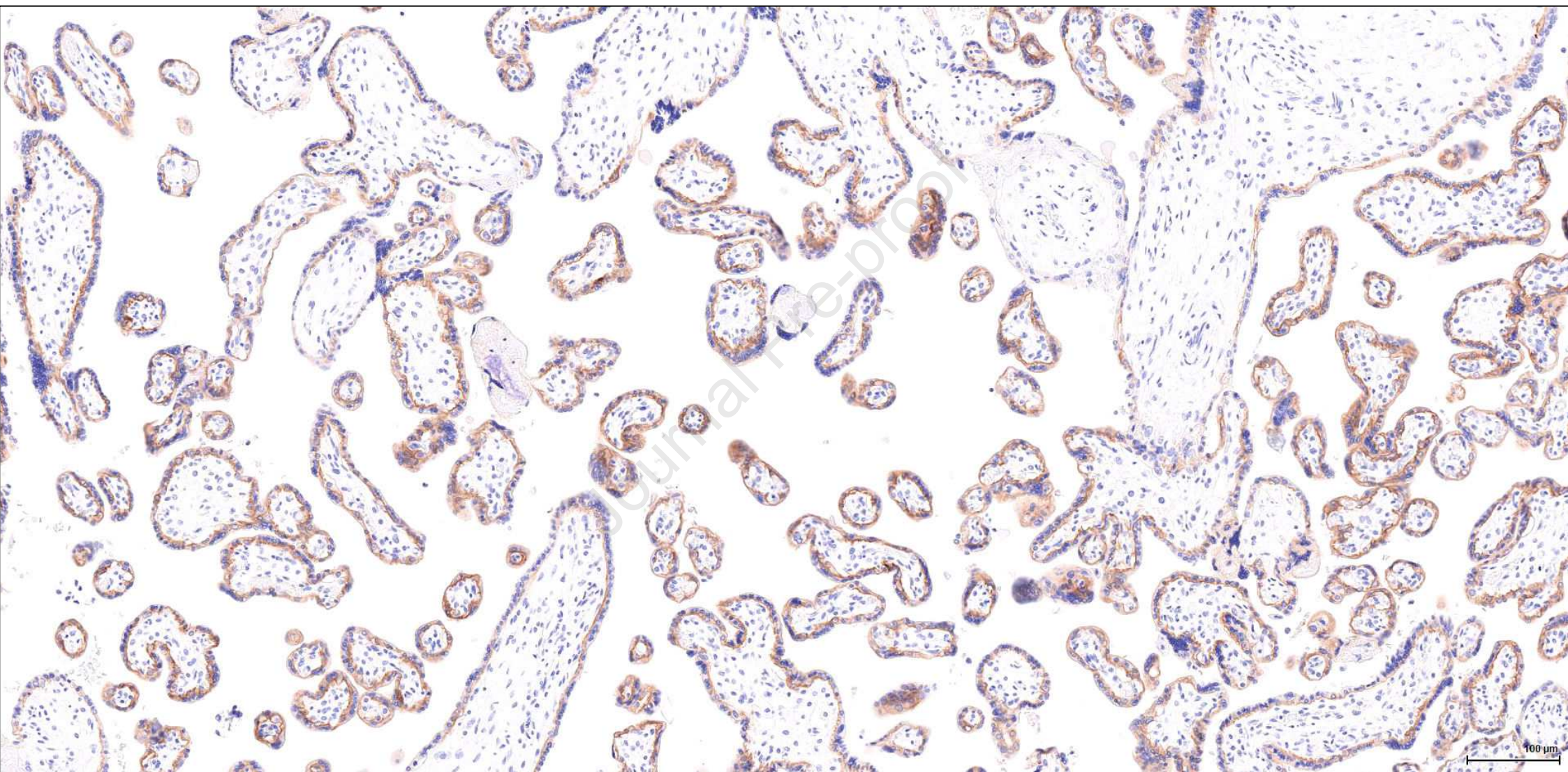
16 weeks



22 weeks



31 weeks



40 weeks

