Serveur Académique Lausannois SERVAL serval.unil.ch

Author Manuscript

Faculty of Biology and Medicine Publication

This paper has been peer-reviewed but dos not include the final publisher proof-corrections or journal pagination.

Published in final edited form as:

Title: CRTC2 polymorphism as a risk factor for the incidence of metabolic syndrome in patients with solid organ transplantation.

Authors: Quteineh L, Bochud PY, Golshayan D, Crettol S, Venetz JP, Manuel O, Kutalik Z, Treyer A, Lehmann R, Mueller NJ, Binet I, van Delden C, Steiger J, Mohacsi P, Dufour JF, Soccal PM, Pascual M, Eap

CB, and The Swiss Transplant Cohort Study

Journal: The pharmacogenomics journal

Year: 2015 Dec 8

DOI: 10.1038/tpj.2015.82





1	CRTC2 polymorphism as a risk factor for the incidence of metabolic
2	syndrome in patients with solid organ transplantation
3	
4	CRTC2 and Post-transplant metabolic syndrome.
5	
6	Lina Quteineh, MD, PhD (1), Pierre-Yves Bochud, MD (2), Dela Golshayan, MD, PhD (3),
7	Severine Crettol, PhD (1), Jean-Pierre Venetz, MD (3), Oriol Manuel, MD (2,3), Zoltán
8	Kutalik, PhD (4,5), Andrea Treyer, PharmD (1), Roger Lehmann, MD (6), Nicolas J Mueller,
9	MD (7), Isabelle Binet, MD (8), Christian van Delden, MD (9), Jürg Steiger, MD (10), Paul
10	Mohacsi, MD (11), Jean-francois Dufour, MD, PhD (12), Paola M. Soccal, MD (13), Manuel
11	Pascual, MD (3), Chin B Eap, PhD (1, 14) and the Swiss Transplant Cohort Study.
12	
13	¹ Unit of Pharmacogenetics and Clinical Psychopharmacology, Department of Psychiatry,
14	Lausanne University Hospital, Prilly, Switzerland.
15	² Service of Infectious Diseases, Lausanne University Hospital, Lausanne, Switzerland
16	³ Transplant Center, Lausanne University Hospital, Lausanne, Switzerland.
17	⁴ Institute of Social and Preventive Medicine (IUMSP), Lausanne University Hospital,
18	Lausanne, Switzerland.
19	⁵ Swiss Institute of Bioinformatics, Lausanne, Switzerland.
20	⁶ Service of Endocrinology and Diabetes, University Hospital, Zurich, Switzerland.
21	⁷ Division of Infectious Diseases and Hospital Epidemiology, University Hospital, Zurich,
22	Switzerland.
23	⁸ Service of Nephrology and Transplantation Medicine, Kantonsspital, St Gallen, Switzerland.
24	⁹ Service of Infectious Diseases, University Hospitals, Geneva, Switzerland.
25	¹⁰ Service of Nephrology, University Hospital, Basel, Switzerland.
26	¹¹ Swiss Cardiovascular Center Bern, University Hospital, Bern, Switzerland.

27	Department of Clinical P	harmacology, University Hosp	pital, Bern, Switzerland.
28	¹³ Service of Pulmonary Me	edicine, University Hospital, C	Geneva, Switzerland.
29	¹⁴ School of Pharmaceutical	Sciences, University of Gene	eva, University of Lausanne, Geneva,
30	Switzerland.		
31			
32			
33			
34			
35	For correspondence:		
36	Prof CB. Eap		
37	Hôpital de Cery,		
38	1008 Prilly – Lausanne, Switze	erland	
39	Tel: 0041 21 314 26 04	Fax: 0041 21 314 24 44	Email:chin.eap@chuv.ch
40			
41			
40			
42			
43			
44			
45			
46			
47			
48			
49			

ABSTRACT

Metabolic syndrome after transplantation is a major concern following solid organ transplantation
(SOT). The CREB-regulated transcription coactivator 2 (CRTC2) regulates glucose metabolism. The
effect of CRTC2 polymorphisms on new-onset diabetes after transplantation (NODAT) was
investigated in a discovery sample of SOT recipients (n ₁ =197). Positive results were tested for
replication in two samples from the Swiss Transplant Cohort Study (STCS, n_2 =1294 and n_3 =759).
Obesity and other metabolic traits were also tested. Associations with metabolic traits in population-
based samples (n_4 =46'186, n_5 =123'865, n_6 >100,000) were finally analyzed. In the discovery sample,
CRTC2 rs8450-AA genotype was associated with NODAT, fasting blood glucose and BMI
(p _{corrected} <0.05). CRTC2 rs8450-AA genotype was associated with NODAT in the second STCS
replication sample (OR=2.01, p=0.04). In the combined STCS replication samples, the effect of
rs8450-AA genotype on NODAT was observed in patients having received SOT from a deceased
donor and treated with tacrolimus (n=395, OR=2.08, p=0.02) and in non-kidney transplant recipients
(OR=2.09, p=0.02). Moreover, rs8450-AA genotype was associated with overweight or obesity
(n=1215, OR=1.56, p=0.02), new-onset hyperlipidemia (n=1007, OR=1.76, p=0.007), and lower
HDL-cholesterol (n=1214, β =-0.08, p=0.001). In the population-based samples, a proxy of
rs8450G>A was significantly associated with several metabolic abnormalities. CRTC2 rs8450G>A
appears to play an important role in the high prevalence of metabolic traits observed in patients with
SOT. A weak association with metabolic traits was also observed in the population-based samples.

Key terms: Transplantation, metabolic syndrome after transplantation, genetic polymorphisms.

Introduction

75

76

77

78

79

80

81

82

83

84

85

86

87

88

89

90

91

92

93

94

95

96

97

98

99

100

The introduction of calcineurin inhibitors (CNIs) - cyclosporine (CSA) and tacrolimus (TAC) - in solid organ transplantation (SOT) has reduced the incidence of acute rejection episodes and improved short-term graft survival^{1, 2}. However, both drugs are not devoid of metabolic complications, such as glucose intolerance, hypertension, and hyperlipidemia which can be very pronounced and have a detrimental impact on patients' life quality, and increase the mortality risk due to cardiovascular events³. Indeed, cardiovascular disease is responsible for approximately 20-40% of non-graft-related death after the first post-transplantation year⁴⁻⁶. New onset diabetes mellitus after transplantation (NODAT) is a serious complication partly related to the use of CNIs, mainly TAC^{7,8}. It is also associated with increased cardiovascular events, infectious complications, and graft loss^{9, 10}. There are several risk factors which increase the risk of NODAT in transplantation such as obesity, increased age, male sex, deceased donor, hepatitis C (HCV) status, acute rejection, and African-American or Hispanic descent^{9, 11}. The identification of genetic factors involved in the development of NODAT may greatly benefit from a more detailed understanding of the complex metabolic pathways involved in glucose metabolism. Genome-wide association studies (GWAS) conducted to date explain only 10% of type 2 diabetes heritability and more diabetic susceptibility genes remain to be discovered (reviewed in 12 and 13). A recent GWAS investigating NODAT identified eight polymorphisms¹⁴. However, only one of these was replicated in another study¹⁵. Whereas GWAS have been extremely valuable, other approaches are needed to further understand the pathophysiology of NODAT. The cAMP-regulated transcriptional coactivator 2 (CRTC2), a transcriptional coactivator that promotes the transcription of genes targeted by the cAMP response element-binding protein¹⁶, is an interesting target protein in glucose metabolism. CRTC2 belongs to the CRTC family which comprises 2 other members, CRTC1 and CRTC3¹⁷. CRTC1 is mainly expressed in the central nervous system and we recently showed an association between CRTC1 polymorphisms and obesity markers (body mass index (BMI) and fat mass) in psychiatric and population-based samples¹⁸, CRTC2 is highly expressed in the thymus, and present in both T and B-lymphocytes¹⁷. It is also expressed in the liver and it plays a direct role during the fasting state in the induction of gluconeogenic genes¹⁶, It further enhances hepatic insulin signalling by stimulating expression of the insulin receptor substrate 2 gene, thus triggering a feedback response that limits glucose output from the liver during fasting¹⁹, Calcineurin, the target of both TAC and CSA, plays an important role in the activation of CRTC2^{20, 21}. Overexpression of CRTC2 induced by a mutation at 2 regulatory sites rendered CRTC2 constitutively active in an animal model and permitted CRTC2-target gene activation even when calcineurin was inhibited by CNIs²². The authors of this study also showed that CRTC2 is required for β-cell function and proliferation and promoting this pathway could ameliorate symptoms of NODAT²². CRTC2 single nucleotide polymorphisms (SNPs) were previously investigated in two Asian populations and one coding SNP (R379C), with a very low minor allele frequency (MAF), was associated with type II diabetes²³ and with lung cancer²⁴. So far CRTC2 SNPs have not been investigated in other ethnic populations or with other phenotypes. In this work, we aimed to study the influence of CRTC2 SNPs on the incidence of NODAT in a sample of Caucasian SOT recipients, and positive results were then tested for replication in the Swiss transplant cohort Study (STCS). We also aimed to extend our analysis to obesity and other variables of the metabolic syndrome (MetS) following SOT. Finally, we aimed to test if the associations with MetS components could be found in general population-based samples (>100,000 subjects).

119

120

121

122

123

124

125

126

127

128

101

102

103

104

105

106

107

108

109

110

111

112

113

114

115

116

117

118

Materials and Methods

Discovery study sample:

This study aimed initially to investigate the effect of genetic polymorphisms of drug metabolizing enzymes and/or transporters on the incidence of different post-transplant complications and on immunosuppressive doses and blood levels^{25, 26}. For this study, a total of 197 patients were enrolled between 2003 and 2005 from the outpatient clinic of the Transplant Center of the University Hospital of Lausanne, Switzerland. Patients with functional graft for more than 12 months after transplantation were eligible to participate in the study. Data regarding patient's age, gender, BMI, ethnic origin, donor's age, HLA mismatch, duration of graft cold ischemia and delayed graft function were collected

retrospectively from patients' medical files. Immunosuppressive regimens, doses and blood levels were obtained retrospectively at different time-points during the first year post-transplantation. The study was approved by the ethics committee of the University of Lausanne. Patients gave their written informed consent to participate in this pharmacogenetic study. Once included, venous blood samples were collected for DNA extraction and further genotyping analysis. For the present study, data were collected between October 2011 and April 2012 as related to the development of NODAT during the first 5 years following transplantation. Data regarding fasting blood glucose (FBG), glycated haemoglobin (HbA1c), 2hrs oral glucose tolerance test (OGTT), insulin and oral anti-diabetic treatment were collected retrospectively from the medical files at the time of transplantation, at 1, 3, 6, 9 and 12 month post-transplantation and at the yearly follow-up until 5-year post-transplantation. NODAT was diagnosed if a patient needed anti-diabetic treatment (either insulin or oral anti-diabetic agents) for at least 6 months following transplantation or had several abnormal glucose profiles during the follow-up period that fulfil the criteria given by the WHO and ADA consensuses²⁷, including FBG \geq 7.0 mmol/l (in \geq two occasions) or 2 hours plasma glucose \geq 11.1 mmol/l during OGTT ²⁷ (even if not treated with anti-diabetic drugs). Patients who had diabetes or prediabetes before transplantation or were transplanted because of diabetic nephropathy were excluded from the present study.

The Swiss Transplant Cohort Study (STCS):

The STCS is an ongoing prospective multicenter cohort project (Basel, Bern, Genève, Lausanne, St. Gallen and Zurich), aiming at a nationwide comprehensive and structured data collection in all SOT recipients. All recipients of SOTs in Switzerland are prospectively registered since May 2008. Currently more than 3500 patients are included in the STCS. No particular eligibility or exclusion criteria exist for enrolment. After transplantation, all patients are mandatorily followed in their respective transplant centers. After baseline assessment, STCS follow-up assessments take place at 6-, 12 months and yearly thereafter. Biological samples are collected in relation to the case at baseline, and at 6-, and the 12-month visits. Data regarding patient's age, gender, BMI, blood pressure and lipid profiles (total cholesterol, HDL- and LDL-cholesterol) were available for all patients at the different

time-points of the STCS follow-up. Full description of this cohort is published elsewhere 28, 29. For the first STCS replication sample, patients transplanted from May 2008 to 8th of May 2011 with a functional graft for at least 12 months after transplantation were included in the analyses (n=1294). NODAT was diagnosed if a patient needed anti-diabetic treatments following transplantation or if such new metabolic event was reported in their case report forms. For the second STCS replication sample, patients transplanted from 9th of May 2011 to May 2013 with a functional graft for at least 12 months after transplantation were included in the analyses (n=759). For this second STCS replication sample, several random and FBG were available in the database, as well as HbA1c. Therefore, as for the discovery sample, NODAT was diagnosed if a patient needed anti-diabetic treatment (either insulin or oral anti-diabetic agents) or fulfilled the criteria given by the WHO and ADA consensuses mentioned earlier. For both STCS samples, patients who were diagnosed with diabetes or were prediabetics before transplantation or were transplanted because of diabetic nephropathy were excluded from the NODAT analyses. For the combined STCS sample, new-onset hypertension and new-onset hyperlipidemia were diagnosed if patients needed anti-hypertensive and hypolipidemic treatment post-SOT. Patients with previous hypertension or hyperlipidemia were excluded from the new-onset hypertension and newonset hyperlipidemia analyses, respectively. The study was approved by the ethics committee of their respective centers. Patients gave their written informed consent to participate in this pharmacogenetic study. Recipients younger than 18 years old and recipients with multiple organ transplantation were excluded from the whole analyses. If a patient received more than one transplantation during the inclusion period, only data from the first SOT was included in the analyses. For both the discovery and STCS samples, as abdominal obesity (waist circumference) was not

181

182

183

184

157

158

159

160

161

162

163

164

165

166

167

168

169

170

171

172

173

174

175

176

177

178

179

180

Population based samples:

available, BMI was used as a marker of obesity.

We aimed to replicate results with MetS traits in several population-based samples: the Meta-Analyses of Glucose and Insulin-related traits Consortium (MAGIC)(n=46'186)³⁰, Genetic Investigation of

ANtropometric Traits (GIANT) consortium (n=123'865)³¹⁻³³, and Genome Wide Associations Scans 185 for Total Cholesterol, HDL-C, LDL-C and triglycerides (n>100'000)^{34, 35}. More details are in the 186 187 Supplementary Material. 188 189 **Polymorphism selection and genotyping:** Two tagging SNPs within CRTC2 gene (rs8450G>A and rs12117078G>C) were selected using 190 191 HapMap Genome Browser by limiting the search to SNPs with a MAF>5% in the Caucasian population and r^2 cutoff of 0.8. Genotyping in the discovery study sample was done using Taqman 192 allelic discrimination assays. Genetic analyses for the STCS replication samples were done using the 193 fluorescence-based competitive allele-specific PCR technology (KASPar)³⁶. More details are in the 194 195 Supplementary Material. 196 197 **Statistical analysis:** 198 Detailed statistical methods are presented in the Supplementary Material. 199 Discovery and STCS samples: 200 We assessed the association of CRTC2 SNPs with MetS traits 1) by applying a logistic regression for 201 binary outcomes adjusted for recipient's age at transplantation and sex. Other variables identified 202 through the univariate analysis (P<0.10) were also added as covariates in these models, 2) by using 203 linear mixed models for continuous variables, and 3) by fitting a Generalized Additive Mixed Model (GAMM) to include a smooth trend for the response in time (allowing multiple observations for each 204 205 patient) for non-linear continuous variables (BMI and FBG). 206 For the discovery sample, the p-values of the models were adjusted for multiple comparisons using 207 Bonferroni correction and the SNP that survived this correction was investigated in the STCS. 208 Data for both the discovery and replications samples were analyzed using Stata 12 (StataCorp, College Station TX, USA) and R version 2.13.0 software (http://www.R-project.org). 209

210

Population-based samples:

We analyzed the associations of *CRTC2* SNP with different MetS traits using multivariate linear regression with allele dosage in which potential confounding factors such as age, sex, and smoking status were added as covariates.

Results:

Discovery and STCS replication samples:

- General characteristics of the discovery and STCS replication samples are presented in the Supplementary Material and Supplementary Tables 1 and 2. Supplementary Table 2 represents the global STCS sample without the exclusion of patients with previous history of diabetes or other metabolic abnormalities before transplantation.
- The distribution of *CRTC2* genotypes in all studies did not deviate from the Hardy-Weinberg equilibrium (P>0.05) and the MAFs were similar to those reported in HapMap (Supplementary Table 3).

CRTC2 SNPs and NODAT, FBG, and BMI in the discovery study sample:

Carriers of the *CRTC2 rs8450-AA* genotype showed increased risk of NODAT as compared to wild-type *GG* genotype (odd ratio (OR)=6.91, 95%CI:1.52-31.36, p_{corrected}=0.02). An increased risk was also observed by analyzing the *rs8450* SNP in a recessive model (Table 1). Neither cumulative prednisone dosages nor calcineurin inhibitors' doses, trough levels or concentration/dose ratios tested at 2 time periods (at 1 month or at 12 months post-SOT) were associated with NODAT in the univariate analysis or when used in the multivariate model including *CRTC2* SNPs and the other covariables (data not shown).

FBG were also available at several time-points after SOT. By applying the GAMM model, increased FBG levels were found in patients with the *CRTC2 rs8450-GA and AA* genotypes (0.24 mmol/L, p_{corrected}=0.06 and 0.47 mmol/L, p_{corrected}=0.02, respectively) as compared to wild-type *GG* genotype. We observed an equal increased of FBG levels by analyzing the *rs8450* SNP in dominant model (Table 1).

Patients with the *CRTC2 rs8450-GA* genotype showed a 1.36 kg/m2 increased BMI compared to the wild-type genotype (p_{corrected}=0.02). By analyzing the SNP in a dominant model, we observed a significant association between *CRTC2 rs8450* and BMI (β=1.17, 95%CI:0.17-2.35, p_{corrected}=0.04).

We observed no association between *CRTC2 rs12117078G>C* SNP and NODAT, FBG or BMI (Table 1).

CRTC2 SNP in the STCS replication samples:

The *CRTC2 rs8450G>A* SNP that survived corrections for multiple testing in the discovery study sample was investigated in the STCS. Recessive models for this SNP are presented in the text and in Tables 2 and 3 while additive and dominant models regarding NODAT are presented in Supplementary Table 4.

CRTC2 SNP and NODAT:

In the univariate analyses, increased recipient's age (p=0.0003), male sex (p=0.03), positive HCV status (p=0.003), deceased donor (p=0.04), treatment with TAC (p=0.001) and baseline BMI (p=0.0002) were among the non-genetic factors associated with an increased risk of NODAT and were used as covariates in the genetic models. In the first STCS replication samples, we observed a non-significant association between *rs8450G>A* and NODAT (Table 2). However, by analyzing the SNP in the subgroup of patients having received SOT from a deceased donor and treated with TAC, a significant effect of the SNP was observed (n=281, OR=2.36, 95%CI:1.16-4.78, p=0.01)(Table 2). In the second replication sample (n=438), in which the diagnostic criteria for NODAT was the same as for the discovery sample, recipients carrying the *rs8450-AA* genotype showed a significant increased risk of NODAT (n=438, OR=2.01, 95%CI:1.03-3.91, p=0.04). In the combined STCS sample, the effect of the *CRTC2* genotype was only observed in the subgroup of patients having received SOT from a deceased donor and treated with TAC (n=395, OR=2.08, 95%CI:1.15-3.74, p=0.02) (Table 2).

CRTC2 SNP and BMI, new-onset hypertension/hyperlipidemia: 265 We investigated the influence of CRTC2 rs8450G>A on BMI and on the incidence of new-onset 266 267 hypertension/hyperlipidemia in the combined STCS sample. Increased recipient's age (p<0.0001), male sex (p<0.0001) and type of transplanted organ (p<0.0001) 268 were among the non-genetic factors associated with increased BMI in the univariate analyses and were 269 used as covariates in the genetic models. BMI values were also dichotomized into normal versus 270 overweight or obese (BMI\ge 25 kg/m2). At 12-months post-SOT, overweight or obese patients were 271 more prevalent in the CRTC2 rs8450-AA genotype compared to the other genotypes (n=1215, 272 273 OR=1.56, 95%CI:1.08-2.25, p=0.02). We observed the same association but with lower significance 274 for the combined STCS sample at 6-months post-SOT (n=1389, OR=1.41, 95%CI:1.01-1.97, p=0.04), 275 and a non-significant association with baseline BMI (n=1515, OR=1.20, 95%CI:0.87-1.65, p=0.27), 276 suggesting that time post-SOT modulates the effect of the CRTC2 genotypes. 277 By applying the GAMM model to test the association between BMI over time and CRTC2 rs8450G>A SNP (n=1625), patients with the *CRTC2 rs8450-AA* genotype showed a 0.47 kg/m² (95%CI:0.11-1.02, 278 279 p=0.01) increase in BMI compared to the other genotypes. 280 We found no association between the CRTC2 rs8450G>A and the incidence of new-onset 281 hypertension (Supplementary Material). On the other hand, carriers of the CRTC2 rs8450-AA 282 genotype showed increased risk of new-onset hyperlipidemia as compared to the other genotypes (n=1007, OR=1.76, 95% CI:1.16-2.66, p=0.007). Additionally, patients with the CRTC2 rs8450-AA 283 284 genotype also showed a significant decrease of HDL-Cholesterol levels in the combined STCS sample

More details can be found in the Supplementary Material.

288

289

290

291

285

286

CRTC2 SNP and MetS traits in kidney and non-kidney transplant recipients:

Patients were dichotomized into kidney versus non-kidney transplant recipients (Table 3). The *CRTC2 rs8450-AA* genotype was associated with significantly higher OR for NODAT in non-kidney

(n=1214, β = -0.08 mmol/L, p=0.001). CRTC2 rs8540G>A SNP was not associated with total

cholesterol or with LDL-cholesterol blood levels in the combined STCS sample (data not shown).

transplant recipients (n=409, OR=2.09, 95%CI:1.13-3.86, p=0.02), while the *rs8450 AA*-genotype was only associated with higher risk of new-onset hyperlipidemia in kidney transplant recipients (n=573, OR=1.95, 95%CI:1.14-3.15, p=0.01).

Population-based samples:

- Several glycemic traits were available in the MAGIC study (Supplementary Table 5); each *rs1572788 C-allele* (in complete linkage disequilibrium (LD) with *rs8450 A-allele*) increased FBG levels by 0.01 mmol/l (n=46'186, p=0.004). Additionally, each *rs1572788 C-allele* decreased HOMA-B by 0.008% (n=46'186, p=0.03). No association was observed between *rs1572788T>C* and HOMA-IR or the 2 hour OGTT.
- Decreased HDL-cholesterol and increased triglycerides were observed for each rs1572788 *C-allele* in the Genome Wide Associations Scans for Total Cholesterol, HDL-C, LDL-C and triglycerides (n=96'908, β =-0.008, p=0.013 and n=93'562, β =0.009, p=0.004, respectively). We observed non-significant associations for rs1572788T>C with total and LDL-cholesterol, as well as with obesity traits studied in the GIANT study, however, the direction of the effect is consistent with increased blood lipids and obesity with each rs1572788 *C-allele* (Supplementary Table 5).

Discussions:

The transcriptional co-activator (CRTC) family is implicated with energy expenditure (CRTC1 and CRTC3)^{37, 38} and glucose metabolism (CRTC2)¹⁶ in animal models. In the present study, we aimed to study the effect of *CRTC2* SNPs on the development of diabetes in SOT recipients, a population at high risk of developing metabolic abnormalities, and in population-based samples. We also extended our analyses to investigate obesity and other metabolic traits. The *CRTC2 rs8450-AA* genotype was associated with increased risk of NODAT, increased FBG, and BMI in a sample of 156 patients with SOT. Results on NODAT were replicated only in the second STCS replication sample (n=438), while in the first and the combined STCS replication samples, the SNP only showed significant influence in the population at higher risk of developing NODAT, mainly in recipients having received a graft from a deceased donor and treated with TAC. We also observed a significant association for BMI in the

STCS replication sample; overweight and obesity were also more frequent in patients carrying the rs8450-AA genotype at 12 months post-SOT. CRTC2 rs8450G>A was also associated with increased risk of new-onset hyperlipidemia following SOT and with lower HDL-cholesterol blood levels. Furthermore, we also found the increased risk of the C-allele of rs1572788 (in complete LD with rs8450 A-allele) on diabetic and lipidemic traits in two large population-based samples (MAGIC and GWAS for Total Cholesterol, HDL-C, LDL-C and triglycerides, respectively), and a consistent direction even though non-significant between rs1572788T>C and obesity traits in the GIANT study. CRTC2 rs8450G>A was associated with NODAT in the total discovery sample and only in the second STCS replication sample, but not in the total STCS sample in which a significant association was observed only in the subgroup of patients at higher risk of developing NODAT. This could partly be explained by the difference of NODAT diagnostic criteria, which is only based on anti-diabetic treatment in the first STCS replication sample. Another explanation could be the high percentage of patients receiving an organ from a living donor in the first STCS replication sample (28%, Supplementary Table 2) compared to only 11.5% in the discovery sample (Supplementary Table 1). Deceased donor is a common risk factor of NODAT¹¹ and the impact of the SNP seems to be more prominent in the groups at higher risk of developing MetS traits. Additionally, when differentiating kidney and non-kidney transplant recipients (Table 3), we observed a significant association between the CRTC2 rs8450-AA genotype and NODAT in non-kidney transplant recipients. The percentage of non-kidney transplant recipients was higher in the second STCS replication sample compared to the first STCS sample (Supplementary Table 2) and it should be noted that kidney transplant recipients had higher percentages of living donors compared to non-kidney transplant recipients (43% vs. 2% respectively). Moreover, kidney transplant recipients usually have obesity and other metabolic problems already before transplantation compared to other solid organ recipients, which could explain the differential effect of the SNP depending on the type of the transplanted organ. Interestingly, in both the discovery and the combined STCS samples, the effect of the SNP was stronger in NODAT cases developed after the first year post-SOT (data not shown), It maybe that the effect of the CRTC2 SNP on NODAT is more important in the long-term, but this remains to be further studied.

319

320

321

322

323

324

325

326

327

328

329

330

331

332

333

334

335

336

337

338

339

340

341

342

343

344

In the GWAS literature, CRTC2 SNPs were not found to be associated with diabetes, obesity or hyperlipidemia. On the other hand, by testing the individual effect of CRTC2 rs1572788T>C SNP on diabetic and lipidemic traits in two population-based samples, a significant effect was observed, even though this effect is very weak and the variance explained by the SNP is very small (Supplementary Table 5) and could thus not be detected by GWAS. However, patients with SOT is a population at risk of developing MetS traits, because of the disease itself and/or the immunosuppressive medications, both can act as an important trigger, unmasking different genetic factors. So in this at risk population, the effect of CRTC2 SNP was more pronounced, even in much smaller sample sizes compared to population-based samples. Additionally, in the GWAS of NODAT14, CRTC2 SNPs did not reach genome-wide level of significance. Moreover, most of the SNPs retained in this GWAS were not replicated in another study with NODAT¹⁵. In fact, both GWAS and candidate gene studies are important gene association approaches and both are subject to the same artifacts of spurious association findings. Depending on the methodology of these studies (sample size, definition of the phenotypes, MAF of the SNPs, corrections for multiple testing, etc...), discordances between both approaches could be seen. GWAS relies on indirect association to locate a disease-causing variant while candidate gene approach relies on an a priori hypothesis to identify this disease-causing variant. For all of these reasons, replication of positive finding in either approach is of utmost importance to validate the results. The functional activity of this SNP, in the regulatory 3'UTR, is unknown and data from 1000

346

347

348

349

350

351

352

353

354

355

356

357

358

359

360

361

362

363

364

365

366

367

368

369

370

371

372

Genomes³⁹ showed that 78 SNPs in the CRTC2 gene region are in high LD (r^2 threshold ≥ 0.8) with rs8450G>A. We used the RegulomeDB database that annotates SNPs with known and predicted regulatory elements in the intergenic regions of the H. sapiens genome⁴⁰. This database reveals known and predicted regulatory DNA elements including regions of DNAase hypersensitivity, binding sites of transcription factors, and promoter regions that have been biochemically characterized to regulation transcription. Source of these data include public datasets from GEO, the ENCODE project, and published literature. Regarding CRTC2 rs8450G>A, several of these proxy SNPs (e.g. rs6680140 with $r^2=1$) show cis-eQTL effect on the expression of CRTC2 and fall into the autoimmune regulatory

(AIRE) motif, influence the binding of several proteins (*JUN*, *CREBBP* and *ELF1*), and has several histone marks (H3k09me3, H4k20me1, etc.). Altogether, these data suggest that the SNP might have a regulatory function.

Interestingly, among the analyzed clinical factors, treatment with corticosteroids was not associated with NODAT, neither in the discovery cohort nor in the STCS sample. However, in our samples most of the patients were treated with corticosteroids so it is difficult to draw major conclusions. On the other hand, the cumulative corticosteroids dosages and the duration of therapy rather than the simple corticosteroids administration could be the most influencing factors. In our discovery sample, the cumulative dosages of corticosteroids at the first year post-transplantation were not associated with NODAT outcome. In the literature, several studies did not find any influence of cumulative corticosteroid dosages on the appearance of NODAT⁴¹⁻⁴⁵. The current lower dosages of corticosteroids used in different immunosuppressive regimens could contribute to the negative association with NODAT appearance in our study.

This study has several limitations and strengths. Random and FBG levels were only available in the second STCS replication sample and the diagnosis of NODAT in the first STCS replication sample was only based on anti-diabetic treatments post-SOT. The genotype of the *rs8450G>A* SNP of the liver transplant donors is unknown, therefore the significant effect of the SNP on NODAT found in non-kidney transplant recipients, in which nearly half of them are liver transplant recipients, should be interpreted with caution. However, CRTC2 is expressed in different tissues and is implicated in glucose regulation through different mechanisms (gluconeogenesis, insulin signaling and beta-cell survival 16, 19, 22) which could highlight the importance of recipient *CRTC2* genotype even in liver transplant recipients. Additionally, by excluding patients with liver transplantation from the non-kidney transplant recipients' analyses, a significant association between the *CRTC2* SNP and NODAT is still observed (data not shown). The present results do not allow the determination of whether the *rs8450G>A* SNP is the causative variant or merely a proxy of one or more yet unidentified variants. Despite the possible regulatory functions proposed by the RegulomeDB database, further studies are needed to elucidate which precise mechanisms underlie the observed association. This study included

people of Caucasian origin and results cannot be generalized to other ethnic groups. On the other hand, the fact that the results were replicated in two independent samples with SOT and in large population-based samples, the latter used as a proof of concept of the effect of the polymorphism, strengthens the validity of our data. In conclusion, our results suggest that variations in CRTC2 play an important role in the high prevalence of MetS complications observed in patients with SOT. Besides, by studying CRTC2 SNPs in population-based samples, we still observed a weak yet significant association, thereby suggesting that the effect of CRTC2 variations is more important in population at risk of developing different MetS traits than in the general population. This is the first study showing the importance of CRTC2 variations with NODAT and other MetS traits in patients with SOT. The assessment of the major risk gene variants would allow predicting vulnerability for developing Mets phenotypes and thus adapt the immunosuppressive treatment from the beginning by genotyping the patients before transplantation. In the long-term, these patients could benefit from individualized immunosuppressive regimens adapted to their genetics and environment.

427	Funding
428	This work has been funded in part by the Swiss National Science Foundation (CBE: 324730_144064).
429	LQ and CBE received research support from the Roche Organ Transplantation Research Foundation
430	(#152358701) and the Swiss Transplant Cohort Study in the past 3 years. ZK was funded by the Swiss
431	National Science Foundation (31003A-143914) and the Leenaards Foundation.
432	This study has been conducted in the framework of the Swiss Transplant Cohort Study, supported by
433	the Swiss National Science Foundation and the Swiss University Hospitals (G15) and transplant
434	centers.
435	
436	Acknowledgements
437	Data on glycaemic traits have been contributed by MAGIC investigators and have been downloaded
438	from www.magicinvestigators.org
439	CBE takes full responsibility for the work as a whole, including the study design, access to data, and
440	the decision to submit and publish the manuscript.
441	
442	Declaration of interests:
443	CBE received honoraria for conferences or teaching CME courses from Advisis, Astra Zeneca,
444	Lundbeck, MSD, Sandoz, Servier and Vifor-Pharma in the past 3 years. He received an unrestricted
445	educational grant from Takeda in the past 3 years.
446	JFD Advisory committees: Bayer, BMS, Gilead Science, Janssen Cilag, Jennerex, Merck, Novartis,
447	Roche. Speaking and teaching: Bayer, Boehringer-Ingelheim, Novartis, Roche
448	SC received honoraria for teaching CME courses from Astra Zeneca and Lundbeck.
449 450	The authors of this manuscript have no conflicts of interest to disclose.

Footnote

451

452

453

454

455

456

457

458

459

460

461

462

463

464

465

466

467

468

The members of the Swiss Transplant Cohort Study are: Rita Achermann, John-David Aubert, Philippe Baumann, Guido Beldi, Christian Benden, Christoph Berger, Isabelle Binet, Pierre-Yves Bochud, Elsa Boely (Head of local data management), Heiner Bucher, Leo Bühler, Thierry Carell, Emmanuelle Catana, Yves Chalandon, Sabina de Geest, Olivier de Rougemont, Michael Dickenmann, Michel Duchosal, Thomas Fehr, Sylvie Ferrari-Lacraz, Christian Garzoni, Yvan Gasche, Paola Gasche Soccal, Emiliano Giostra, Déla Golshayan, Daniel Good, Karine Hadaya, Christoph Hess, Sven Hillinger, Hans Hirsch, Günther Hofbauer, Uyen Huynh-Do, Franz Immer, Richard Klaghofer, Michael Koller (Head of the data center), Thomas Kuntzen, Bettina Laesser, Roger Lehmann, Christian Lovis, Oriol Manuel, Hans-Peter Marti, Pierre Yves Martin, Pascal Meylan (Head, Biological samples management group), Paul Mohacsi, Isabelle Morard, Philippe Morel, Ulrike Mueller, Nicolas Mueller (Chairman Scientific Committee), Helen Mueller-McKenna, Thomas Müller, Beat Müllhaupt, David Nadal, Gayathri Nair, Manuel Pascual (Executive office), Jakob Passweg, Chantal Piot Ziegler, Juliane Rick, Eddy Roosnek, Anne Rosselet, Silvia Rothlin, Frank Ruschitzka, Urs Schanz, Stefan Schaub, Christian Seiler, Nasser Semmo, Susanne Stampf, Jürg Steiger (Head, Executive Office), Christian Toso, Dimitri Tsinalis, Christian Van Delden (Executive office), Jean-Pierre Venetz, Jean Villard, Madeleine Wick (STCS coordinator), Markus Wilhelm, Patrick Yerly.

469

470

471

Supplementary data:

Supplementary information is available at journal's website

472

473

474

References:

477	1.	Mayer AD, Dmitrewski J, Squifflet JP, Besse T, Grabensee B, Klein B, et al. Multicenter
478		randomized trial comparing tacrolimus (FK506) and cyclosporine in the prevention of renal
479		allograft rejection: a report of the European Tacrolimus Multicenter Renal Study Group.
480		Transplantation 1997; 64 (3): 436-443.
481		
482	2.	Hariharan S, Johnson CP, Bresnahan BA, Taranto SE, McIntosh MJ, Stablein D. Improved graft
483		survival after renal transplantation in the United States, 1988 to 1996. The New England

484 485

476

486 3. Ojo AO, Hanson JA, Wolfe RA, Leichtman AB, Agodoa LY, Port FK. Long-term survival in renal transplant recipients with graft function. *Kidney international* 2000; **57**(1): 307-313.

journal of medicine 2000; 342(9): 605-612.

488

489 4. Collins AJ, Foley RN, Herzog C, Chavers B, Gilbertson D, Ishani A, et al. United States Renal
490 Data System 2008 Annual Data Report. American journal of kidney diseases: the official
491 journal of the National Kidney Foundation 2009; 53(1 Suppl): S1-374.

492

Johnston SD, Morris JK, Cramb R, Gunson BK, Neuberger J. Cardiovascular morbidity and mortality after orthotopic liver transplantation. *Transplantation* 2002; **73**(6): 901-906.

495 496

497

498

6. Vogt DP, Henderson JM, Carey WD, Barnes D. The long-term survival and causes of death in patients who survive at least 1 year after liver transplantation. *Surgery* 2002; **132**(4): 775-780; discussion 780.

499

Opelz G, Dohler B, Collaborative Transplant S. Influence of immunosuppressive regimens on graft survival and secondary outcomes after kidney transplantation. *Transplantation* 2009;
 87(6): 795-802.

503

504 8. First MR, Dhadda S, Croy R, Holman J, Fitzsimmons WE. New-onset diabetes after transplantation (NODAT): an evaluation of definitions in clinical trials. *Transplantation* 2013; **96**(1): 58-64.

507

Kasiske BL, Snyder JJ, Gilbertson D, Matas AJ. Diabetes mellitus after kidney transplantation in the United States. American journal of transplantation: official journal of the American Society of Transplantation and the American Society of Transplant Surgeons 2003; 3(2): 178-185.

512

10. Israni AK, Snyder JJ, Skeans MA, Kasiske BL, Investigators P. Clinical diagnosis of metabolic syndrome: predicting new-onset diabetes, coronary heart disease, and allograft failure late after kidney transplant. *Transplant international : official journal of the European Society for Organ Transplantation* 2012; **25**(7): 748-757.

518 11. Pham PT, Pham PM, Pham SV, Pham PA, Pham PC. New onset diabetes after transplantation 519 (NODAT): an overview. Diabetes, metabolic syndrome and obesity: targets and therapy 520 2011; 4: 175-186. 521 522 12. Billings LK, Florez JC. The genetics of type 2 diabetes: what have we learned from GWAS? Ann 523 N Y Acad Sci 2010; 1212: 59-77. 524 525 13. Torres JM, Cox NJ, Philipson LH. Genome wide association studies for diabetes: perspective on results and challenges. Pediatr Diabetes 2013; 14(2): 90-96. 526 527 528 14. McCaughan JA, McKnight AJ, Maxwell AP. Genetics of new-onset diabetes after 529 transplantation. J Am Soc Nephrol 2014; **25**(5): 1037-1049. 530 531 15. Chand S, Shabir S, Chan W, McCaughan JA, McKnight AJ, Maxwell AP, et al. beta cell 532 glucotoxic-associated single nucleotide polymorphisms in impaired glucose tolerance and 533 new-onset diabetes after transplantation. Transplantation 2014; 98(3): e19-20. 534 535 16. Koo SH, Flechner L, Qi L, Zhang X, Screaton RA, Jeffries S, et al. The CREB coactivator TORC2 is 536 a key regulator of fasting glucose metabolism. Nature 2005; 437(7062): 1109-1111. 537 538 17. Conkright MD, Canettieri G, Screaton R, Guzman E, Miraglia L, Hogenesch JB, et al. TORCs: 539 transducers of regulated CREB activity. Mol Cell 2003; 12(2): 413-423. 540 541 18. Choong E, Quteineh L, Cardinaux JR, Gholam-Rezaee M, Vandenberghe F, Dobrinas M, et al. 542 Influence of CRTC1 Polymorphisms on Body Mass Index and Fat Mass in Psychiatric Patients 543 and the General Adult Population. JAMA psychiatry 2013; 70(10): 1011-1019. 544 545 19. Canettieri G, Koo SH, Berdeaux R, Heredia J, Hedrick S, Zhang X, et al. Dual role of the 546 coactivator TORC2 in modulating hepatic glucose output and insulin signaling. Cell 547 metabolism 2005; 2(5): 331-338. 548 549 20. Screaton RA, Conkright MD, Katoh Y, Best JL, Canettieri G, Jeffries S, et al. The CREB 550 coactivator TORC2 functions as a calcium- and cAMP-sensitive coincidence detector. Cell 551 2004; **119**(1): 61-74. 552 553 21. Bittinger MA, McWhinnie E, Meltzer J, Iourgenko V, Latario B, Liu X, et al. Activation of cAMP 554 response element-mediated gene expression by regulated nuclear transport of TORC 555 proteins. Current biology: CB 2004; 14(23): 2156-2161.

557 22. Eberhard CE, Fu A, Reeks C, Screaton RA. CRTC2 is required for beta-cell function and proliferation. *Endocrinology* 2013; **154**(7): 2308-2317.

556

Keshavarz P, Inoue H, Nakamura N, Yoshikawa T, Tanahashi T, Itakura M. Single nucleotide polymorphisms in genes encoding LKB1 (STK11), TORC2 (CRTC2) and AMPK alpha2-subunit (PRKAA2) and risk of type 2 diabetes. *Molecular genetics and metabolism* 2008; **93**(2): 200-209.

564

He Y, Li Y, Qiu Z, Zhou B, Shi S, Zhang K, et al. Identification and validation of PROM1 and CRTC2 mutations in lung cancer patients. *Molecular cancer* 2014; **13**(1): 19.

567

568 25. Crettol S, Venetz JP, Fontana M, Aubert JD, Pascual M, Eap CB. CYP3A7, CYP3A5, CYP3A4, and ABCB1 genetic polymorphisms, cyclosporine concentration, and dose requirement in transplant recipients. *Therapeutic drug monitoring* 2008; **30**(6): 689-699.

571

572 26. Crettol S, Venetz JP, Fontana M, Aubert JD, Ansermot N, Fathi M, et al. Influence of ABCB1 genetic polymorphisms on cyclosporine intracellular concentration in transplant recipients.

774 Pharmacogenetics and genomics 2008; **18**(4): 307-315.

575

576 27. Davidson J, Wilkinson A, Dantal J, Dotta F, Haller H, Hernandez D, et al. New-onset diabetes 577 after transplantation: 2003 International consensus guidelines. Proceedings of an 578 international expert panel meeting. Barcelona, Spain, 19 February 2003. *Transplantation* 579 2003; **75**(10 Suppl): SS3-24.

580

581 28. Koller MT, van Delden C, Muller NJ, Baumann P, Lovis C, Marti HP, et al. Design and methodology of the Swiss Transplant Cohort Study (STCS): a comprehensive prospective nationwide long-term follow-up cohort. *European journal of epidemiology* 2013; **28**(4): 347-355.

585

586 29. Manuel O, Kralidis G, Mueller NJ, Hirsch HH, Garzoni C, van Delden C, et al. Impact of antiviral preventive strategies on the incidence and outcomes of cytomegalovirus disease in solid organ transplant recipients. American journal of transplantation: official journal of the American Society of Transplantation and the American Society of Transplant Surgeons 2013; 13(9): 2402-2410.

591

592 30. Prokopenko I, Langenberg C, Florez JC, Saxena R, Soranzo N, Thorleifsson G, et al. Variants in MTNR1B influence fasting glucose levels. *Nature genetics* 2009; **41**(1): 77-81.

594

595 31. Lango Allen H, Estrada K, Lettre G, Berndt SI, Weedon MN, Rivadeneira F, et al. Hundreds of variants clustered in genomic loci and biological pathways affect human height. *Nature* 2010; 467(7317): 832-838.

598

599 32. Speliotes EK, Willer CJ, Berndt SI, Monda KL, Thorleifsson G, Jackson AU, et al. Association analyses of 249,796 individuals reveal 18 new loci associated with body mass index. *Nature genetics* 2010; **42**(11): 937-948.

603 33. Heid IM, Jackson AU, Randall JC, Winkler TW, Qi L, Steinthorsdottir V, et al. Meta-analysis 604 identifies 13 new loci associated with waist-hip ratio and reveals sexual dimorphism in the 605 genetic basis of fat distribution. Nature genetics 2010; 42(11): 949-960. 606 607 34. Genome Wide Associations Scans for Total Cholesterol, HDL-C, LDL-C and triglycerides. 608 609 35. Teslovich TM, Musunuru K, Smith AV, Edmondson AC, Stylianou IM, Koseki M, et al. 610 Biological, clinical and population relevance of 95 loci for blood lipids. Nature 2010; 466(7307): 707-713. 611 612 613 36. He C, Holme J, Anthony J. SNP genotyping: the KASP assay. Methods in molecular biology 614 2014; **1145**: 75-86. 615 616 37. Altarejos JY, Goebel N, Conkright MD, Inoue H, Xie J, Arias CM, et al. The Creb1 coactivator 617 Crtc1 is required for energy balance and fertility. Nature medicine 2008; 14(10): 1112-1117. 618 619 38. Song Y, Altarejos J, Goodarzi MO, Inoue H, Guo X, Berdeaux R, et al. CRTC3 links 620 catecholamine signalling to energy balance. Nature 2010; 468(7326): 933-939. 621 622 39. Johnson AD, Handsaker RE, Pulit SL, Nizzari MM, O'Donnell CJ, de Bakker PI. SNAP: a web-623 based tool for identification and annotation of proxy SNPs using HapMap. Bioinformatics 624 2008; 24(24): 2938-2939. 625 626 40. Boyle AP, Hong EL, Hariharan M, Cheng Y, Schaub MA, Kasowski M, et al. Annotation of functional variation in personal genomes using RegulomeDB. Genome research 2012; 22(9): 627 628 1790-1797. 629 630 41. Rodrigo E, Fernandez-Fresnedo G, Valero R, Ruiz JC, Pinera C, Palomar R, et al. New-onset diabetes after kidney transplantation: risk factors. Journal of the American Society of 631 632 Nephrology: JASN 2006; 17(12 Suppl 3): S291-295. 633 634 42. Numakura K, Satoh S, Tsuchiya N, Horikawa Y, Inoue T, Kakinuma H, et al. Clinical and genetic 635 risk factors for posttransplant diabetes mellitus in adult renal transplant recipients treated 636 with tacrolimus. *Transplantation* 2005; **80**(10): 1419-1424. 637 638 43. Gourishankar S, Jhangri GS, Tonelli M, Wales LH, Cockfield SM. Development of diabetes 639 mellitus following kidney transplantation: a Canadian experience. American journal of 640 transplantation: official journal of the American Society of Transplantation and the American 641 Society of Transplant Surgeons 2004; **4**(11): 1876-1882. 642 643 44. Sulanc E, Lane JT, Puumala SE, Groggel GC, Wrenshall LE, Stevens RB. New-onset diabetes

after kidney transplantation: an application of 2003 International Guidelines. Transplantation

644

645

2005; **80**(7): 945-952.

45. Maes BD, Kuypers D, Messiaen T, Evenepoel P, Mathieu C, Coosemans W, et al. Posttransplantation diabetes mellitus in FK-506-treated renal transplant recipients: analysis of incidence and risk factors. *Transplantation* 2001; **72**(10): 1655-1661.

Table 1 Association between *CRTC2* SNPs and NODAT, blood glucose levels and BMI in patients with solid organ transplantation from the discovery study sample:

	NODAT (n=156)		FBG (n=156)		BMI (n=156)	
SNP	OR (95% CI)	P corrected	Difference (mmol/L) compared to wild-type (ref) (95% CI)	P corrected	Difference (kg/m2) compared to wild-type (ref) (95% CI)	P corrected
CRTC2 rs84	150					
GG	ref		ref		ref	
GA	1.81 (0.71 - 4.61)	0.46	0.24 (-0.00 - 0.49)	0.06	1.36 (0.28- 2.42)	0.02
AA	6.91 (1.52 - 31.36)	0.02	0.47 (0.07 - 0.86)	0.02	0.37 (-1.38 - 2.06)	0.76
Dominant	2.32 (0.96 - 5.63)	0.12	0.28 (0.05 - 0.51)	0.02	1.17 (0.17 - 2.35)	0.04
Recessive	5.11 (1.24 - 21.03)	0.04	0.36 (-0.03 - 0.74)	0.06	-0.24 (-2.33 - 1.68)	0.80
CRTC2 rs12	2117078					
GG	ref		ref		ref	
GC & CC	0.36 (0.11 - 1.16)	0.18	-0.07 (-0.40 - 0.24)	0.70	0.24 (-1.41 - 1.62)	0.78

NODAT: New-onset diabetes after transplantation, FBG: Fasting blood glucose, OR: odd ratio, CI: confidence interval, BMI: body mass index.

Regarding NODAT and blood glucose levels analyses: P values were adjusted for age of recipient at transplantation, sex of the recipient and baseline BMI and type of calcineurin inhibitor.

Regarding BMI analyses: P values were adjusted for age of recipient at transplantation, sex of the recipient and type of calcineurin inhibitor.

Table 2: Association between *CRTC2 rs8450* SNP in a recessive model and NODAT in the first, second and combined population of STCS and in different subgroups at risk of NODAT:

CRTC2 rs8450		GG&GA (n)	AA	
	N total		NODAT OR (95% CI)	P value
First STCS replication sample [§] :				
Total	711	Ref (n=623)	1.02 (0.62 - 1.70)	0.93
Treatment with TAC & deceased donor	281	Ref (n=244)	2.36 (1.16 - 4.78)	0.01
Second STCS replication sample [§] :				
Total	438	Ref (n=385)	2.01 (1.03 - 3.91)	0.04
Combined STCS replication sample [§] :				
Total	1149	Ref (n=1008)	1.25 (0.84 - 1.87)	0.26
Treatment with TAC & deceased donor	395	Ref (n=340)	2.08 (1.15 - 3.74)	0.02

[§]Excluding patients with previous history of diabetes before transplantation

NODAT: New-onset diabetes after transplantation, TAC: tacrolimus, OR: odd ratio, CI: confidence interval P values were adjusted (when appropriate) for age of recipient at transplantation, sex of the recipient, hepatitis C status, baseline BMI, type of calcineurin inhibitor and type of donor.

Table 3: Association between CRTC2 rs8450 SNP and MetS traits in all kidney and non-kidney transplant recipients in the combined STCS sample

	Kie	dney transplant recip	ients	Non-kidney transplant recipients			
	n	OR	p-value	n	OR	p-value	
	(95	5% CI) for rs8450-AA			(95% CI) for rs8450-AA	١	
NODAT	740	0.87 (0.50 - 1.52)	0.62	409	2.09 (1.13 - 3.86)	0.02	
BMI≥25 kg/m2 at 12months post SOT	761	1.59 (0.98 - 2.58)	0.06	454	1.52 (0.84 - 2.75)	0.17	
New-onset hyperlipidemia	573	1.95 (1.14 - 3.15)	0.01	434	1.73 (0.84 - 3.56)	0.14	
New-onset hypertension	146	1.64 (0.43 - 6.30)	0.47	345	1.44 (0.75 - 2.78)	0.27	

OR: odd ratio, CI: confidence interval

NODAT: New-onset diabetes after transplantation, BMI: body mass index

For NODAT analyses: P values were adjusted for age of recipient at transplantation, sex of the recipient, baseline BMI, type of calcineurin inhibitor and type of donor.

For BMI analyses: P values were adjusted for age of recipient at transplantation and sex of the recipient.

For new-onset hyperlipidemia analyses: P values were adjusted for age of recipient at transplantation, sex of the recipient, baseline BMI, the type of calcineurin inhibitor and treatment with corticosteroids.

For new-onset hypertension analyses: P values were adjusted for age of recipient at transplantation and sex of the recipient, treatment with corticosteroids, the type of calcineurin inhibitor and type of donor.

1	Supplementary data:
2	Materials and Methods
3	- Population based samples:
4	> The Meta-Analyses of Glucose and Insulin-related traits Consortium (MAGIC)
5	➤ Genetic Investigation of ANtropometric Traits (GIANT) consortium
6	> Genome Wide Associations Scans for Total Cholesterol, HDL-C, LDL-C and
7	triglycerides
8	- Polymorphism selection and genotyping
9	- Statistical analysis
10	
11	Results:
12	- General characteristics of the discovery sample
13	- General characteristics of the replication sample (STCS):
14 15	- CRTC2 SNP and new-onset hypertension in the STCS
16 17	- CRTC2 SNP and new-onset hyperlipidemia in the STCS
18	
19	Supplementary Tables 1-5
20	
21	
22	
23	
24	
25	
26	
20	

Population based samples: The Meta-Analyses of Glucose and Insulin-related traits Consortium (MAGIC) MAGIC is a large-scale meta-analyses of genome-wide data for continuous diabetes-related traits in participants without diabetes ¹. Meta-analyses of ~2.5 million directly genotyped or imputed autosomal single nucleotide polymorphisms (SNPs) were performed from genome-wide association studies (GWAS). These cohorts include up to 46'186 non-diabetic participants of European descent informative for FBG, the surrogate estimates of beta-cell function (HOMA-B) and insulin resistance (HOMA-IR) derived from fasting variables by homeostasis model assessment and up to 15'234 non-diabetic individuals informative for 2 hour oral glucose tolerance test (OGTT) ². Genetic Investigation of ANtropometric Traits (GIANT) consortium The GIANT consortium performed a meta-analysis of GWAS data with a discovery set of 123'865 individuals of European ancestry from 46 studies for height ³, BMI⁴ and waist-to hip ratio ⁵. Genome Wide Associations Scans for Total Cholesterol, HDL-C, LDL-C and triglycerides Data on lipid traits have been downloaded from "Genome Wide Associations Scans for Total Cholesterol, HDL-C, LDL-C and triglycerides" website 6, 7 which is a meta-analysis of 46 lipid GWASs. These studies together comprise >100,000 individuals of European descent (maximum sample size 100,184 for Total Cholesterol, 95,454 for LDL-C, 99,900 for HDL-C and 96,598 for triglycerides), ascertained in the United States, Europe or Australia.

Materials and Methods

Polymorphism selection and genotyping:

55

80

81

82

Polymorphisms within CRTC2 were selected using HapMap Genome Browser (release 28)⁸. Two 56 57 tagging SNPs were found by limiting the search to SNPs within CRTC2 gene ± 1kb with a minor allele frequency > 5% in the Caucasian population and r^2 cutoff of 0.8: rs8450G>A within the 3'UTR 58 and the intronic rs12117078G>C SNP. 59 Genotyping of these 2 SNPS for the discovery study sample was done using Taqman allelic 60 61 discrimination assays (ABI PRISM 7000 Sequence Detection System; Applied Biosystems, Rotkreuz, Switzerland) after genomic DNA extraction from whole blood. Life technologies Taqman genotyping 62 assays C_8722376_10 and C_30997814_10 were used for CRTC2 rs8450G>A and 63 rs12117078G>C SNPs, respectively. Genotyping were performed in the Unit of Pharmacogenetics 64 and Clinical Psychopharmacology, in Lausanne, Switzerland. 65 Genetic analyses for the first STCS replication sample were performed by KBioscience Institute in 66 United Kingdom using the novel fluorescence-based competitive allele-specific PCR technology 67 (KASPTM). Details available 68 about this technology are at 69 http://www.lgcgenomics.com/genotyping/kasp-genotyping-chemistry/. Genetic analyses for the second STCS replication sample were also performed using the KASPTM 70 71 technology on ABI 7500 Fast real-time thermocycler (Applied Biosystems). Genotyping were 72 performed in the service of infectious disease in Lausanne University Hospital, Lausanne, 73 Switzerland, according to manufacturer's protocols (LGC Genomics, UK). The KASP primers were designed by KrakenTM assay design and workflow management software and further validated by 74 manufacturer (LGC Genomics, UK). Automated allele calling was performed using SDS software 75 76 according to standard protocols (Applied Biosystems). The following primers were used for CRTC2 77 rs8450G>A genotyping in this sample: primer allele FAM: ACCCCTAGGCATCCGGAAAAG, 78 primer allele HEX: CACCCCTAGGCATCCGGAAAAA and common primer 79 GGGTAGAGGGGAGCCCTGGAA.

Statistical analysis:

Quantitative data are presented as median and range unless otherwise mentioned, while qualitative data are presented as frequency and percentage. For association studies, the chi-square (Chi²) test or the Fisher exact test for binomial variables were used. Differences in allele and genotype frequencies as well as deviation from Hardy-Weinberg equilibrium were assessed using Chi² test.

Discovery study sample:

For NODAT analyses, logistic regression was applied adjusted for recipient's age at transplantation, sex, BMI at baseline and type of CNIs. Due to nonlinearity of our models and the absence of any linear transformation, the association between *CRTC2* SNPs with BMI and FBG levels was assessed by fitting a Generalized Additive Mixed Model (GAMM) ^{9,10} to allow a smooth trend for the response in time based on multiple observations for each patient (using a thin plate regression spline basis) adjusting for recipient's age at transplantation, sex, and type of CNIs (antidiabetic treatments for FBG analyses). GAMMs were fitted using the mgcv package of R (settings were fixed at package defaults). In order to be more conservative, the uncertainty of estimated parameters were assessed by 1000 bootstraps ¹¹ at the subject level and results were similar with those gained by 10'000 bootstraps.

Due to the small number of subjects being homozygous for the variant allele of *CRTC2*

Replication samples (STCS):

rs12117078G>C, this SNP was analyzed in a dominant model.

For the statistical analyses of NODAT, new-onset hypertension, new-onset hyperlipidemia, and overweight or obese, logistic regression models were applied adjusted for recipient's age at transplantation and sex. Other variables identified through the univariate analysis (P<0.10) were also added as covariates in these models. GAMM models were applied for BMI and HDL-cholesterol analyses in the STCS sample, as both models were not linear. Linear mixed models were used for systolic and diastolic blood pressure analyses and for blood total cholesterol and LDL-cholesterol analyses. In addition to recipient's age at transplantation, sex, BMI at baseline and type of CNIs, blood

pressure models and lipid models were also adjusted for antihypertensive and hypolipidemic drugs intake, respectively.

Population-based samples:

The associations of *CRTC2* SNP with different MetS traits were analyzed using multivariate linear regression with allele dosage in which potential confounding factors such as age, sex and smoking status were added as covariates. FBG, OGTT, the surrogate estimates of beta-cell function (HOMA-B) and insulin resistance (HOMA-IR) were analyzed in the MAGIC study. BMI, waist circumference and waist-to hip ratio were the only adiposity traits analyzed by the GIANT Consortium. Triglycerides, total cholesterol, HDL-cholesterol and LDL-cholesterol were analyzed in the "Genome Wide Associations Scans for Total Cholesterol, HDL-C, LDL-C and triglycerides" study.

Results:

General characteristics of the discovery sample

Among the 197 patients included in the initial study, 17 patients were excluded because they were diagnosed with diabetes before transplantation, 11 patients had no follow-up data in their medical files, 2 patients did not sign consents for MetS-related genetic analysis, 4 patients dropped-out from the study and 7 were non-Caucasians. Finally, 156 Caucasian patients were included in the current study, 65% were kidney transplant recipients. The mean age at transplantation was 47.8 ± 11 years, 61% were male and the mean BMI pre-transplant was 24 ± 4 kg/m². During the five years of follow-up post-transplantation, NODAT occurred in 45 (28.9%) patients. Most NODAT cases (67%) occurred during the 1st year after transplantation. Most of the patients were treated with CSA (n=102, 65%), while 35% (n=54) were treated with TAC. As expected, NODAT was higher in the TAC-treated group compared to CSA¹² (46.3 vs. 19.6 % respectively). Full characteristics of our population and their distribution between NODAT and non-NODAT patients are presented in Supplementary Table 1. Recipient age (p=0.002), male sex (p=0.05), donor age (p=0.04), baseline BMI (p=0.0003) and the type of CNIs (p<0.001) were significantly different between patients with and without NODAT (Supplementary Table 1).

General characteristics of the replication sample (STCS):

Overall, 1294 patients from the first STCS replication sample received a SOT from May 2008 until May 2011, among them 1219 patients had both the clinical data and the genetic material available and were included in the current study. By excluding patients younger than 18 years old and patients with multiple organ transplantation, 958 patients remained and included in the analysis. The mean age at transplantation was 52.3 ± 13 years, 66% were male and pre-transplant BMI was 24.6 kg/m² (range:13.7-41.2). During the follow-up period post-transplantation, NODAT, new-onset HTN and new-onset hyperlipidemia occurred in 27.4%, 67.6% and 33.1% of the patients, respectively. Among the patients included in the second STCS replication sample (n=759) with both clinical and genetic data and by excluding patients younger than 18 years old and patients with multiple organ transplantation, 667 remained and were included in the analysis. Lower incidence of NODAT, new-onset HTN and new-onset hyperlipidemia were observed in this sample, which could be explained by the shorter follow-up duration compared to the first STCS replication sample (Supplementary Table 2). Full description of the first, second and combined STCS samples are presented in Supplementary Table 2. All patients from both STCS samples were of Caucasian origin.

CRTC2 SNP and new-onset hypertension in the STCS:

Among the non-genetic factors, type of calcineurin inhibitor (p<0.0001), type of transplanted organ (p<0.0001), treatment with corticosteroids (p<0.0001) and type of donor (p=0.02) were associated with increased risk of new-onset hypertension after SOT in the univariate analyses and were used as covariates in the genetic models. A non-significant association was observed between *CRTC2 rs8450G>A* and new-onset hypertension, except for patients carrying the *rs8450-GA* genotype which showed a protective effect against new-onset hypertension (n=491, OR=0.62, 95%CI: 0.42 - 0.94, p=0.02) compared to wild type genotype.

shown).

CRTC2 SNP and new-onset hyperlipidemia in the STCS: Among the non-genetic factors, increased recipient's age (p=0.003), baseline BMI (p=0.006), type of calcineurin inhibitor (p=0.005) and treatment with corticosteroids (p<0.0001) were associated with increased risk of new-onset hyperlipidemia after SOT in the univariate analyses and were used as covariates in the genetic models.

191 Supplementary Table 1: Clinical characteristics of the discovery study sample:

Characteristic	Total	Non-NODAT	NODAT	p-value
Recipient age at transplantation (years), median (range)	48 (22-68)	46 (22-68)	53 (28-68)	0.002
Recipients sex (Males) [%]	60.9	55.9	73.3	0.05
Living donor [%]	11.5	11.7	11.1	0.92
Donor age (years), median (range)	43.5 (10-73)	41 (10-73)	48 (13-69)	0.04
Donor sex (Males) [%]	56.8	63.2	54.8	0.06
Organ, n [%]				
lung	17 (10.9)	10 (9.0)	7 (15.6)	
kidney	102(65.4)	75 (67.6)	27(60.0)	0.46
liver	37(23.7)	26 (23.4)	11 (24.4)	
BMI pre-transplant (kg/m²), median (range)	23.5 (15.8-37.3)	22.3 (15.8-36.2)	26.4 (18.8-37.3)	0.0003
BMI 5 year follow-up (kg/m²), median (range)	26.0 (16.7-43.6)	25.5 (16.7-43.6)	27.1 (19.9-42.3)	0.10
Recipient CMV infection (R ⁺) [%]	49.3	52.5	42.2	0.25
Donor CMV infection (D ⁺) [%]	61.5	59.2	66.7	0.39
Recipient and Donor CMV infection (R^+/D^+) [%]	27.6	27.0	28.9	0.81
Calcineurin inhibitors, n [%]				
TAC	54 (34.6)	29 (26.1)	25 (55.6)	<0.001
CSA	102 (65.4)	82 (73.9)	20 (44.4)	-0.001
First transplantation [%]	80.1	77.5	86.7	0.19
Acute rejection episode during first year post-transplant [%]	47.4	47.2	47.7	0.96

NODAT: New-onset diabetes after transplantation, BMI: body mass index, CMV: cytomegalovirus, R: recipient, D: Donor, TAC: Tacrolimus, CSA: Cyclosporine.

196 Supplementary Table 2: Clinical characteristics of STCS§

197

198

	First STCS	Second STCS	combined STCS replication
Characteristic	replication sample	replication sample	sample
	(n=958)	(n=667)	(n=1625)
Recipient age at transplantation (years),	FF (19.70)	FF (70)	FF (19.70)
median (range)	55 (18-79)	55 (79)	55 (18-79)
Recipients sex (Males) [%]	66.2	64.6	65.5
Period of follow-up (months), median (range)	12 (0-60)	6 (0-48)	6 (0-60)
Living donor [%]	28.7	24.0	26.7
Donor age (years)	53 (1-86)	55 (1-88)	53 (1-88)
Donor sex (Males) [%]	53.5	53.4	53.4
Organ, n [%]			
Kidney	624 (65.1)	360 (54.0)	984 (60.5)
Liver	158 (16.5)	142 (21.3)	300 (18.5)
Lung	94 (9.8)	97 (14.5)	191 (11.8)
Heart	65 (6.8)	57 (8.6)	122 (7.5)
Islets and pancreas [£]	17 (1.8)	11 (1.6)	28 (1.7)
BMI pre-transplant (kg/m²), median (range)	24.6 (13.7-41.2)	24.9 (13.4-43.5)	24.8 (13.4-43.5)
BMI 1 year follow-up (kg/m²), median (range)	25.3 (15.4-44.6)	24.8 (13.7-41.7)	25.2 (13.7-44.6)
Recipient CMV infection (R ⁺) [%]	57.5	59.5	58.2
Donor CMV infection (D ⁺) [%]	53.4	58.4	56.2
Recipient and Donor CMV infection (R^{+}/D^{+}) [%]	33.0	36.3	34.3
Calcineurin inhibitors, [%]			
TAC	65.5	67.1	66.3
CSA	26.7	26.1	26.4
None	7.8	6.8	7.3
Incidence of NODAT [%]	27.4	18.7	24.4
Incidence of New hypertension [%]	67.6	32.8	53.6
Incidence of New hyperlipidemia [%]	33.1	13.8	26.6

[§]patients included in this table represent the global STCS sample with the clinical data available for the analyses without the exclusion of patients with diabetes, hypertension or hyperlipidemia before transplantation.

[£]These patients had type I diabetes mellitus as an indication for transplantation and were excluded from NODAT analysis. Abbreviations: BMI: body mass index, CMV: cytomegalovirus, R: recipient, D: Donor, TAC: Tacrolimus, CSA: Cyclosporine, NODAT: New-onset diabetes after transplantation, New hypertension: new onset hypertension after transplantation, New hyperlipidemia: new onset hyperlipidemia after transplantation.

Supplementary Table 3: Genotype frequencies in the discovery sample and in the first, the second and the combined STCS samples. Hardy-Weinberg equilibrium and a comparison between the observed minor allele frequency and HapMap minor allele frequency in the Caucasian population.

	CRTC2 SNPs Frequencies n (%)			Hardy-Weinberg	MAF	
SNP		,	(1.7)	Equilibrium (p-value)	observed	НарМар
Discovery sam	ple (n=156)					
rs8450	GG	GA	AA	0.87	0.30	0.30
	78 (50.0)	64 (41.0)	14 (9.0)			
rs12117078	GG	GC	CC	0.75	0.10	0.10
	126 (80.8)	28 (17.9)	2 (1.3)			
STCS						
rs8450	GG	GA	AA			0.30
First STCS sample	408 (43.0)	429 (44.2)	112 (11.8)	0.96	0.34	
(n=949) [§]						
second STCS sample	281 (42.1)	302 (45.3)	84 (12.6)	0.84	0.35	
(n=667)						
Combined	689 (42.7)	731 (45.2)	196 (12.1)	0.92	0.35	
STCS sample (n=1616)	. ,	, ,	130 (12.1)	0.32	0.00	

^{§9} missing genotypes for this sample.

MAF: minor allele frequency.

Supplementary Table 4: Association between *CRTC2 rs8450* SNP and NODAT in total population of STCS in different subgroups at risk of NODAT:

	Additive model							Dominant model		
CRTC2 rs8450		GG	GA		AA		GG	GA&AA		
	N		NODAT OR	p-value	NODAT OR	p-value		NODAT OR	p-value	
			(95% CI)		(95% CI)			(95% CI)		
First STCS replication sa	mple:									
Total	711	ref	0.97 (0.68 - 1.38)	0.85	1.01 (0.59 - 1.73)	0.98	ref	0.97 (0.70 - 1.37)	0.88	
Treatment with TAC & deceased donor	281	ref	1.08 (0.63 - 1.85)	0.79	2.45 (1.14 - 5.26)	0.02	ref	1.31 (0.79 - 2.17)	0.30	
Second STCS replication	sample:									
Total	438	Ref	0.81 (0.47 - 1.40)	0.46	1.80 (0.88 - 3.71)	0.11	ref	0.99 (0.60 - 1.62)	0.96	
Combined STCS replicati	ion samp	ole:								
Total	1149	ref	0.89 (0.66 - 1.20)	0.44	1.18 (0.76 - 1.81)	0.46	ref	0.95 (0.72 - 1.25)	0.70	
Treatment with TAC & deceased donor	395	ref	0.81 (0.51 - 1.30)	0.39	1.86 (0.99 - 3.52)	0.06	ref	1.00 (0.65 - 1.54)	0.99	

TAC: tacrolimus, OR: odd ratio, CI: confidence interval

P values were adjusted (when appropriate) for age of recipient at transplantation, sex of the recipient, hepatitis C status, baseline BMI, type of calcineurin inhibitor and type of donor.

Supplementary Table 5: Associations between $CRTC2\ rs1572788\ T>C$ (in complete linkage disequilibrium (LD) with $rs8450\ G>A$) and glycemic, lipidemic and obesity traits in several

oopulation-based samples:	Effect	of each <i>rs1572788</i>	C-allele					
	(complete LD with <i>rs8450 A-allele</i>)							
phenotype	n	beta	p-value	Explained variance*				
Glucose [£]	46186	0.01	0.004	0.0002				
2h glucose tolerance test [£]	15234	-0.02	0.26					
HOMA-β [£]	46186	-0.008	0.03	0.0001				
HOMA-IR [£]	46186	0.0002	0.96					
Total cholesterol [§]	97148	0.005	0.16					
HDL-Cholesterol [§]	96908	-0.008	0.013	0.0001				
LDL-Cholesterol [§]	92503	0.006	0.06					
Triglycerides [§]	93562	0.009	0.004	0.0001				
BMI ^{\$}	113955	0.004	0.37					
Waist-hip ratio ^{\$}	55282	0.002	0.71					
Waist circumference ^{\$}								
Male	38305	0.008	0.32					
Female	47320	0.01	0.19					

explained variance by the polymorphism (only calculated for p<0.05).

[£] This clinical variable was analyzed in the MAGIC study.

§ This clinical variable was analyzed in the "Genome Wide Associations Scans for Total Cholesterol, HDL-C, LDL-C and triglycerides".

§ This clinical variable was analyzed in the GIANT study.

²h: 2 hours, HOMA: homeostatic model assessment, β: beta-cell function, IR: insulin resistance, BMI: Body mass index.

References:

- 1. Prokopenko I, Langenberg C, Florez JC, Saxena R, Soranzo N, Thorleifsson G, et al. Variants in MTNR1B influence fasting glucose levels. *Nature genetics* 2009; **41**(1): 77-81.
- 2. Dupuis J, Langenberg C, Prokopenko I, Saxena R, Soranzo N, Jackson AU, et al. New genetic loci implicated in fasting glucose homeostasis and their impact on type 2 diabetes risk. *Nature genetics* 2010; **42**(2): 105-116.
- 3. Lango Allen H, Estrada K, Lettre G, Berndt SI, Weedon MN, Rivadeneira F, et al. Hundreds of variants clustered in genomic loci and biological pathways affect human height. *Nature* 2010; 467(7317): 832-838.
- 4. Speliotes EK, Willer CJ, Berndt SI, Monda KL, Thorleifsson G, Jackson AU, et al. Association analyses of 249,796 individuals reveal 18 new loci associated with body mass index. *Nature genetics* 2010; **42**(11): 937-948.
- 5. Heid IM, Jackson AU, Randall JC, Winkler TW, Qi L, Steinthorsdottir V, et al. Meta-analysis identifies 13 new loci associated with waist-hip ratio and reveals sexual dimorphism in the genetic basis of fat distribution. *Nature genetics* 2010; **42**(11): 949-960.
- 6. Genome Wide Associations Scans for Total Cholesterol, HDL-C, LDL-C and triglycerides.
- 7. Teslovich TM, Musunuru K, Smith AV, Edmondson AC, Stylianou IM, Koseki M, et al. Biological, clinical and population relevance of 95 loci for blood lipids. *Nature* 2010; **466**(7307): 707-713.
- 8. Barnes MR. Navigating the HapMap. Briefings in bioinformatics 2006; 7(3): 211-224.
- 9. Wood SN. GAMMs with R. In: Carlin BP, Chatfield C, Tanner M, Zidek J (eds). *Generalized Additive Models An Introduction with R*. Chapman & Hall/CRC: Broken, 2006, pp 319-324.
- 10. Lin X, Zhang D. Inference in generalized additive mixed models by using smoothing splines. *J R Statist Soc B* 1999; **61**(2): 381-400.
- 11. Davison AC, Hinkley DV. *Bootstrap Methods and their Application*. Cambridge University Press: Cambridge, New York, 1997, 1-592pp.
- 12. Opelz G, Dohler B, Collaborative Transplant S. Influence of immunosuppressive regimens on graft survival and secondary outcomes after kidney transplantation. *Transplantation* 2009; **87**(6): 795-802.