1	A test of the size-constraint hypothesis for a limit to sexual
2	dimorphism in plants
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## 24 Abstract

25 In flowering plants, many dioecious species display a certain degree of sexual dimorphism in 26 non-reproductive traits, but this dimorphism tends to be much less striking than that found in 27 animals. Sexual size dimorphism in plants may be limited because competition for light in 28 crowded environments so strongly penalises small plants. The idea that competition for light 29 constrains the evolution of strong sexual size dimorphism in plants (the size-constraint 30 hypothesis) implies a strong dependency of the expression of sexual size dimorphism on the 31 neighbouring density as a result of the capacity of plants to adjust their reproductive effort 32 and investment in growth in response to their local environment. Here, we tested this 33 hypothesis by experimentally altering the context of competition for light among male-female 34 pairs of the light-demanding dioecious annual plant Mercurialis annua. We found that males 35 were smaller than females across all treatments, but sexual size dimorphism was diminished 36 for pairs grown at higher densities. This result is consistent with the size-constraint 37 hypothesis. We discuss our results in terms of the tension between selection on size acting in 38 opposite directions on males and females, which have different optima under sexual selection, 39 and stabilizing selection for similar sizes in males and females, which have similar optima 40 under viability selection for plasticity in size expression under different density 41 conditions.

43 In organisms with separate sexes, males and females often differ from one another in 44 morphological, life-history and physiological traits that are not directly related to primary sex 45 expression or gamete production (Darwin 1871). Such secondary sexual dimorphism is 46 common in animals, but is also known to occur in plants for a wide range of characters 47 (reviewed in Barrett and Hough 2013), including life-history (reviewed in Delph 1999, Obeso 48 2002), morphological (reviewed in Dawson and Geber 1999), physiological (reviewed in 49 Dawson and Geber 1999) and defence traits (reviewed in Ågren et al. 1999). For example, 50 males usually have larger flowers and/or floral displays than females (e.g. Silene latifolia, 51 Meagher 1992; Delph et al. 1996), the sexes may display different leaf shapes and sizes (e.g. 52 Simmondsia chinensis, Wallace and Rundel 1979), males may grow more quickly (e.g. Acer 53 negundo, Jing and Coley 1981; Dacryodes excelsa) or more slowly than females (e.g. Silene 54 latifolia, Gross and Soule 1981; Mercurialis annua, Hesse and Pannell 2011) and males tend 55 to be more susceptible to herbivores than females (e.g. Fragaria virginiana, Ashman et al. 2004; reviewed in Ågren et al. 1999; Obeso 2002). Although sexual secondary dimorphism is 56 57 often extreme in animals (Darwin 1871; Fairbairn 1997; Moore and Pannell 2011), it is rarely 58 so in plants, and it is seldom possible to tell males and females apart on the basis of vegetative 59 traits (Lloyd and Webb 1977; Delph et al. 1996; Moore and Pannell 2011; Barrett and Hough 60 2013). A striking exception is offered by the differences in leaf and branching traits between 61 the sexes in several species of the Cape fynbos genus *Leucadenron* (Bond and Midgely 1988; 62 Bond and Maze 1999; Harris and Pannell 2010).

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The fact that differences between males and females of dioecious plants tend often to be more
subtle than those found in many animal species probably reflects, in part, reduced
opportunities for sexual selection in plants (Skogsmyr and Lankinen 2002; Moore and Pannell

67 2011; Forrest 2014). The arena for female choice in plants is obviously much narrower than it

is in animals, being limited to the occasional influence females might exercise in which pollen 68 69 grains are allowed to fertilize their ovules (e.g. Lankinen and Madjidian 2011) and perhaps 70 aspects of the phenology of flowering (e.g. Brunet and Charlesworth 1995; Gleiser et al. 71 2008). Female choice would in any case likely affect floral rather than vegetative traits. Male-72 male competition is more likely to occur in plants, and many floral adaptations have probably 73 evolved in response to selection for increased siring success, both in hermaphrodites and 74 dioecious plants (reviewed in: Moore and Pannell 2011; Barrett and Hough 2013). Again, 75 selection for siring success will tend to shape floral and inflorescence traits, e.g., by 76 increasing the size or number of flowers or inflorescences or the amount of pollen produced 77 and dispersed, rather than vegetative traits directly. However, because competition for siring 78 success may bring about selection for increased absolute investment by individuals in 79 flowering (Bond and Maze 1999), specifically in their male function, we would expect 80 allocation trade-offs to have indirect effects on vegetative traits that might differ between 81 males and females (Delph et al. 1996; Bond and Maze 1999; Delph et al. 2004; Harris and 82 Pannell 2010; Moore and Pannell 2011; Barrett and Hough 2013). 83

84 Differences in trade-offs between investment in reproduction and growth and/or maintenance 85 between males and females are probably the main reason for many of the traits for which 86 dioecious species show secondary sexual dimorphism (Barrett and Hough 2013). Thus, for 87 example, the oft-observed increased susceptibility by males to herbivores or pathogens may be the outcome of steeper trade-offs between reproduction and defence in males than females 88 89 (reviewed in Ågren 1999). Similarly, different life-history schedules between males and 90 females (Delph 1999), including patterns of phenology and mortality, probably explain the 91 strong differences in secondary sex ratios in many dioecious plant populations, where the sex 92 making the heavier investment in reproduction flowers later or dies earlier (Pannell and Ojeda

2000; Field et al. 2013). This sex is often female (leading to male-biased sex ratios), but may
also be male (reviewed in Delph 1999; Field et al. 2013).

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96 The idea of trade-offs between sexual selection (and investment to enhance mating success) 97 and ecological (e.g., viability) selection lies at the heart of models for the evolution of sexual 98 dimorphism: the phenotype selected should be that for which marginal gains in fitness via one 99 component (e.g., through increased siring success) equal the marginal losses in another (e.g., 100 through increased mortality; Andersson 1994; Shuster and Wade 2003). In this sense, sexual 101 dimorphism in plants might thus be limited either by low benefits accruing to individuals with 102 phenotypes that diverge from an ecological optimum (because the scope for sexual selection 103 is limited) or by high costs of such divergence (because of elevated risks of mortality or 104 reproductive failure for other reasons), or by both factors together. In cases where these 105 benefits and costs vary with environmental conditions (Delph and Bell 2008), we would 106 expect the phenotypes expressed by males and females to vary as a result of phenotypic 107 plasticity, so that measures of sexual dimorphism might vary among environments, too. Here, 108 we test this idea with particular reference to sexual size dimorphism in a wind-pollinated 109 annual plant growing at different densities. We label our hypothesis the 'size-constraint 110 hypothesis'.

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The size-constraint hypothesis posits that ecological selection among plants under
competition for light will modulate the extent to which males and females can deviate from
one another in terms of their size. It is well known that plants often face strong asymmetrical
competition with both intraspecific and heterospecific neighbours for light, with the tallest
individuals gathering a disproportionate share of the resource (Schwinning and Weiner 1998).
In dioecious populations, intraspecific competition will occur both between plants of the same

118 sex and plants of the opposite sex. Plants possess a range of adaptations that allow them to 119 adjust their growth in response to their competitive environment. Preeminent among these is 120 their ability to increase growth in height in response to a shifted Red:Far-Red (R:FR) light 121 ratio, which serves as a signal for the presence of green neighbours (Ballare et al. 1990; 122 Schmitt and Wulff 1993; Ballare 1999). Responses to the R:FR light ratio allow plants avoid 123 competitive suppression that occurs when taller plants overshadow shorter ones (shade 124 avoidance response, Ruberti et al. 2012). A well-known consequence of competition for light 125 is the tendency for plants to reduce their reproductive effort in the face of competition for 126 light (Harper 1977), probably in response to life-history trade-offs associated with a higher 127 investment in growth. This trade-off partially explains the decrease in the seed yield of crops 128 per area planted with increases in plant density (Harper 1977, see also Sangoi et al. 2002). If 129 sexual dimorphism reflects a deviation by males and/or females from a common phenotype 130 that is optimal for success in competing for light, then the size-constraint hypothesis predicts 131 that sexual size dimorphism will be smaller in light-demanding dioecious plants at high 132 density (or, in general, under conditions where success at gaining access to light depends on 133 size); by contrast, plants growing at low density will be freer to deviate from the common 134 phenotype, and sexual size dimorphism will be more pronounced.

135

We tested the size-constraint hypothesis by manipulating the perceived density of plants of the wind-pollinated herb *Mercurialis annua*, a dioecious annual that shows substantial sexual size dimorphism, with males typically smaller than females in terms of biomass and often also height (Harris and Pannell 2008; Sanchez et al. 2011; Sanchez and Pannell 2011). Our experiment involved comparing sexual size dimorphism among plants expressing their phenotypes at different experimental densities, and in the middle versus at the edge of experiment plots. Because we were interested specifically in the response of males and

143 females in competition with one another for light, we grew plants in male-female pairs in 144 individual pots and varied the spacing between pots. Thus, all plants in our experiment were 145 subjected to the same competition for underground resources, but varied among treatment 146 densities in terms of their access to light. Because our focus was on sexual dimorphism, i.e., 147 the difference between male and female phenotypes, we used male-female pairs as the unit of 148 replication. We predicted that, in dense situations, males and females would be constrained to 149 grow to similar sizes, whereas, in sparse situations, they could afford to deviate from a size 150 optimal for competition for light and hence potentially express greater sexual size 151 dimorphism. Previous work (Harris and Pannell 2008) has shown that males tend to be 152 smaller than females of *M. annua*, partially because of their particularly heavy investment in 153 reproduction, presumably a behaviour that has evolved under intense scramble competition 154 for siring success. Sexual selection thus appears to favour males with high reproductive effort 155 and thus smaller sizes. However, small sizes are expected to be strongly penalised in dense 156 situations under asymmetrical competition for light.

157

### 158 Materials and methods

159 Study system

160 Mercurialis annua (Euphorbiaceae) is a wind-pollinated annual herb that occupies disturbed 161 habitats in central and Western Europe, as well as around the Mediterranean Basin (Durand 162 1963, Tutin et al. 1968). The species shows remarkable variation in its sexual systems, 163 including monoecious, dioecious and androdioecious populations (where males co-occur with 164 hermaphrodites) in different parts of its range (Pannell et al. 2004). Here, we focus on the 165 dioecious populations, which are diploid and occur throughout much of Europe. These 166 populations are sexually dimorphic, with male plants typically shorter and lighter than 167 females and possessing pedunculate inflorescences that are held erect above the plant whereas 168 females display sessile flowers at each leaf axil (Eppley and Pannell 2007). Reproduction 169 begins several weeks following germination, with males often commencing inflorescence 170 production a few days earlier than females (Harris and Pannell 2008). Flowering in *M. annua* 171 is indeterminate, with plants dispersing their pollen and seeds as they grow, meaning that 172 there is no clearly distinct vegetative and reproductive phases (Hesse and Pannell 2011). The 173 plants used in the study originated from seeds bulked from 25 naturally occurring diploid 174 dioecious populations, sampled across its native European range.

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## 176 Experimental design and measurements

177 We tested the size constraint hypothesis by experimentally altering the context of light 178 competition for pairs of male and female plants. We established 18 plots, each comprising 25 179 male-female pairs in single pots (i.e., each pot comprised a male and a female together); 180 henceforth, 'pot' refers to male-female pairs. Seeds randomly picked from the bulk pool were 181 individually sown in peat pellets (Jiffy Products International), and 900 young plants (450 of 182 each sex) were transplanted in male-female pairs into 15 cm diameter pots at the onset of 183 flowering, three weeks after germination. To reduce asymmetries in competition caused by 184 slight differences in the timing of germination, we matched males and females of 185 approximately similar height for each pair. Plants were then moved out-of-doors in a ploughed field on the campus of the University of Lausanne (Switzerland). Individuals were 186 187 supported with wooden stakes in their pots to prevent toppling in the wind.

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Within each plot, the randomly assigned 25 pots were arranged in a square lattice (5 x 5) with an inter-pot distance of 0 cm (pots touching), 5 cm, 15 cm, 25 cm, 35 cm, or 85 cm. These six density levels represent our competition treatment. Each density level was replicated three times. The lattice arrangement meant that there were nine internal pots and 16 edge pots in

193 each plot, and that competition for light experienced by the pots in the dense plots was 194 diminished for the pots placed at the edge. The experiment ran out-of-doors for eight weeks, 195 and plants were harvested when growth in size appeared to be levelling off and signs of 196 nitrogen limitation were being shown (a slight yellowing of the leaves). At harvest, we found 197 that pots were completely full of roots, pointing to the likelihood of strong belowground 198 competition. During growth, plants were watered daily, as necessary. There was very little 199 mortality during this period, with only two males and one female dying in plots of 200 intermediate density. At the end of week eight, all plants were measured for height, and 201 aboveground parts were then collected, bagged, dried and weighed.

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### 203 <u>Statistical analysis</u>

204 To test for the effect of interplant competition on sexual dimorphism in plant height and mass, 205 we ran two separate linear mixed-models (LMM), with sex, distance and position (edge 206 versus central placement) and their interactions as fixed variables and replicate pots nested 207 within plot replicate as random effects. As we found a significant effect on plant mass of the three-way interaction sex × distance × position ( $\gamma^2_5 > 15.6$ ; p =0.008), we ran two separate 208 209 LMMs on plants reared in pots in the centre and at the edge of the plot, respectively. To test 210 whether male and female mass were differently affected by their position in the plot, we also 211 ran two separate LMMs on males and females, respectively.

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To quantify more precisely the extent to which males and females differed within each pot, we calculated a sexual dimorphism index (SDI; Lovich and Gibbons 1992) per pot by subtracting the male trait value from the female trait value and dividing by the male trait value, excluding the three pots in which one of the plants had died. We used separate Wilcoxon tests for each density level to test whether mass SDI differed significantly from 0

(i.e., whether male and female masses were significantly different). We ran these tests
specifically on central pots where the effect of density was strongest. To compare SDI among
the different density levels, we used either a non-parametric Kruskal-Wallis test or a one-way
Welch's ANOVA, depending on the homogeneity of variances of SDI in the group of interest
(i.e. mass SDI or height SDI in the center or at the edge of the plot). Homogeneity of the
variances was tested with the non-parametric Flinger-Killeen test for homogeneity of

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We used backward elimination (successively removing factors with P > 0.10) until we

obtained a minimal adequate model, comparing models with log-likelihood ratio tests

228 (LLRT). When necessary, we normalised the response variable using the Boxcox

transformation. Data are given as mean  $\pm$  SE, unless specified. All analysis was conducted on

R 3.0.2 (R Core Team 2013), with LLMs implemented using the package lme4.

231

#### 232 Results

## 233 Effects of density on male and female plant mass and height

234 Male and female plants were lighter when they experienced more competition for light (Fig.

1a) and female plants experienced a steeper decrease in mass with increased plant density

than males (Fig 1a). Distance between the pots and position of the pot within the plot both

affected plant mass differently in male and female plants, as revealed by a significant three-

238 way interactions (LLRT sex × position × distance:  $\chi^2_5 > 15.6$ ; p = 0.008): while both male and

239 female plant mass decreased with increased competition for light, females placed in the center

- of the plot were significantly lighter than the females placed at the edge when density
- increased (Fig 1b; distance  $\times$  position interaction: p < 0.001; Table 1). In contrast, this effect

of the position of the plant within the plot was only marginally significant for males (Fig. 1b; p = 0.07; Table 1).

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Males and females were taller when they experienced more competition for light (Fig. 2a).
The position of the plant in the plot (edge vs. center) did not significantly affect the pattern of
height variation in female plants (Fig. 2b, Table 2), but it did affect males. At higher densities,
males from the centre of plots were significantly taller than the males at their edge (Fig. 2b;
Table 2).

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251 Effects of density on sexual dimorphism within pots

252 On average, mass SDI was positive, i.e., females were larger than males (Table 3). This 253 applied to total biomass, but also to vegetative parts after flowers and fruits had been 254 removed from females (seed and fruit mass accounted for about 10% of the female mass 255 in all densities, except for distance class 0 in which it accounted for 7.7% of the female 256 mass). However, because males were weighed with with their inflorescences, we 257 present mass SDI on the basis of a comparison between the sexes reproductive 258 structures intact. Mass SDI changed across the different density levels, with a tendency 259 to increase when the level of competition for light was relaxed (Fig.3). This was the case 260 especially in pots placed in the center of plots, though these differences were not 261 significantly different (Table 3). It is worth noting that variance was high among pots 262 placed in the center of the less dense plots (Table 3), partially explained by the presence 263 of two extreme outliers with Mass SDI>20 (Fig.3). Mass SDI in central pots was 264 significantly different from 0 for the distance classes 5, 15, 35 and 85 (with p < 0.03) and 265 fell short of significance for the distance class 0 and 25 (p = 0.08). Pots placed at the 266 edge of plots showed somewhat less variation in mass SDI among densities (Fig.3),

though these differences were not statistically different (Table 3). On average, the
highest mass SDI was often reached at the interplant distance 35 (Table 3), i.e., not the
sparsest plots.

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Height SDI increased significantly when density decreased for pots placed in the centerof the plots, but not for pots placed at the edge of plots (Table 4).

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## 274 Discussion

Notwithstanding high variation among lowest-density plots, we found that sexual dimorphism
in size tended to be lower in plots with high density than lower density, as well as for plants
reared in plot centers compared with their edges. Both these results largely conform to the
size-constraint hypothesis, which predicts that competition for access to light represents an
ecological component of selection that will strongly constrain responses to sexual selection.

281 It is somewhat puzzling that plants at the lowest-density plot (i.e., 85cm) showed lower sexual 282 size dimorphism than the plants at the density level 35cm, hence reversing the trend we 283 observed across the other treatments (i.e. reduced sexual size dimorphism when increasing 284 density levels from 35cm to 0cm). We grew our plants in pots that were perhaps more 285 susceptible to drying out in low-density situations that high-density situations, and this might 286 have affected growth rates at the lowest-density extreme of our experiment. While growing 287 plants in pots may poorly emulate natural conditions, it allowed us in our experiment to 288 ensure that all plants experienced the same belowground competitive situation. Although we 289 watered all pots similarly, differences in desiccation rates among pots might have introduced unintended variation among the treatments we imposed. We can offer no other explanation for 290 291 the seemingly anomalous finding at the lowest plot densities. As might be expected, the effect

of growing at the edge compared to the center of plants was most apparent at high-density
plots, where the competitive environment experienced by plants will have been greatest, i.e.,
growing at the edge of a plot released plants from competition more in high- than in lowdensity plots.

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297 In the case of *M. annua*, male investment in flowering and nitrogen-greedy pollen production 298 leads to a particularly steep trade-off with aboveground growth, so that males that invest in 299 nitrogen-harvesting roots tend to be smaller than females (Harris and Pannell 2008). Our 300 results indicate that, in dense situations, males and females respond plastically by maintaining 301 sizes more consistent with their opposite-sex neighbours. Interestingly, Conn and Blum 302 (1981) also found that differences in the size of males and females of the dioecious annual 303 *Rumex hastatulus* were smaller at high density, though the result was not interpreted in terms 304 of constraints on sexual selection. Similarly, Lovett Doust et al. (1987) noted that the much-305 described patterns of sexual dimorphism in Silene latifolia (e.g., higher number of stems in 306 female and higher number of flowers in males; Delph et al. 2002) disappeared in experimental 307 conditions under high densities.

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309 The experimental setups of Conn and Blum (1981) and Lovett Doust et al. (1987) affected 310 both below-ground and above-ground competition simultaneously, and the authors discussed 311 their results in terms of the calorific allocation to reproduction by males versus females. 312 Because all plants were exposed to the same level of below-ground competition, our 313 experiment teases apart the two components of competition and focuses on the importance of 314 above-ground competition only. Although competition for resources below ground will often 315 be important, especially where soil nutrients are limiting, above-ground competition for light 316 has the peculiarity of being highly asymmetrical between plants with different sizes

317 (Schwinning and Weiner 1998). Once plants are overtopped by their neighbours, their slower 318 growth will expose them to increasingly low light quality in a process with positive-feedback 319 on performance, ultimately leading to mortality – the cause of 'self-thinning' in dense plant 320 populations (Weller 1987; Lonsdale 1990; Vanderwerf et al. 1995). The essence of the size-321 constraint hypothesis is that selection will favour phenotypes that maintain similar sizes to 322 their neighbours when competition is intense but that males and females may attain different 323 sizes when released from the competitive constraint at lower densities. It is well known that 324 plants can respond in their growth to the presence of neighbours that they detect through their 325 sensitivity to the R:FR ratio: green neighbours transmit or reflect low R:FR ratios, which 326 induce altered growth patterns, including stem extension, internode elongation and decreased 327 branching (Schmitt and Wulff 1993). Sleeman et al. (2002) documented a phytochrome-328 mediated shade-avoidance response in androdioecious *M. annua*, which occupies similar open 329 habitat to dioecious populations we studied here. Our results here suggest that dioecious M. 330 annua also responds to R:FR ratio, and that this response does not differ much between the 331 sexes.

332

333 From the perspective of mating opportunities and reproduction, plant size may experience 334 different selective pressures/constraints in females and males that would explain sexual size 335 dimorphism. The outcome of allocation trade-offs between growth and reproduction would be 336 a good reason to expect males and females to differ in size. The sex with the greater 337 reproductive burden, i.e., the greater marginal cost of reproduction when measured in terms of 338 its negative effect on growth, is expected to be the smaller, particularly after allocation to 339 reproduction has commenced. Lloyd and Webb (1977) suggested that the result of such trade-340 offs would be evident only in perennial plants with iteroparous reproduction, but the 341 prediction should also apply to annuals with indeterminate reproduction, like *M. annua*,

342 because reproduction early in the season will affect ongoing growth and opportunities for 343 reproduction later in the same season (in this sense, such plants are 'iteroparous' annuals). In 344 iteroparous species, including annuals with indeterminate reproduction, the premium on plant 345 height for maintaining a positive carbon budget should strongly affect density-dependent 346 patterns of allocation, as has indeed been observed for *M. annua* (Pannell 1997). This 347 premium should apply especially in dense stands, so that both males and females will be 348 selected to maintain optimum (and similar) height by adjusting their allocation to 349 reproduction. In sparse stands where height will be less important in maintaining a positive 350 carbon budget, males and females should adopt a gender-dependent strategy in the trade-off 351 between growth and reproduction, leading to greater sexual size dimorphism, as largely 352 observed in our experiment.

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354 A second reason why we may expect males and females to differ in size could be the selection 355 on male height for pollen dispersal. In wind-pollinated species (especially herbs such as M. 356 annua), tall males will benefit from an outcross siring advantage over their smaller male 357 competitors by dispersing their pollen to the wind more effectively. This advantage has been 358 labelled a 'direct effect' of size (Klinkhamer et al. 1997). Eppley and Pannell (2007) found 359 that, in *M. annua*, the dispersal of pollen from erect inflorescences held above plants 360 increased siring success per pollen grain by approximately 60%, a result that is consistent 361 with a direct effect of size (though plant height per se was not estimated). In another revealing study, Pickup and Barrett (2012) found that although females were taller than males in wind-362 363 pollinated *R. hastatulus* late in the reproductive season, males were the taller sex when pollen 364 was being dispersed. This observation can be interpreted as an outcome of selection on height 365 in males when it matters for mating. Despite the mating advantage for taller males of M. 366 annua in the context of pollen dispersal, selection on height may be overridden by selection

367 on reproductive investment (Harris and Pannell 2008), which might explain why males are368 typically smaller than females in this species.

369 Our results illustrate the fact that the high levels of plasticity displayed by plants in their 370 response to the environment (Schlichting 1986; Sultan 2000) affect traits that can be sexually 371 dimorphic in dioecious plants, so that indices of dimorphism themselves become plastic. 372 Indeed, sex-specific plasticity in response to environmental conditions may be responsible for 373 intraspecific variation in sexual dimorphism, as has been hypothesized to explain sexual 374 dimorphism variation in insects (Stillwell et al. 2010). Similarly, sex-specific plasticity 375 probably explains the fact that, in the plant Simmondsia chinensis, sexual dimorphism in 376 several vegetative traits was observed only under drier conditions (for further examples, see 377 the review by Delph 1999).

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379 In discussing the size-constraint hypothesis, we have emphasised the flexibility that plants 380 have in responding to their local conditions, particularly in varying their allocation to growth 381 versus reproduction. The reduced sexual size dimorphism observed for dioecious species at 382 high density, consistent with the size-constraint hypothesis, is likely a response that allows 383 both males and females to compete with one another on a more equal footing than would be 384 the case if their allocation patterns reflected a response to selection on reproduction only. But 385 this buffering effect of plasticity is clearly only partial. There is ample evidence that sex ratios 386 in dioecious plant populations vary widely, often as a result of differential mortality under 387 different conditions (reviewed in Field et al. 2013). Such patterns reflect a kind of niche 388 partitioning between the sexes (Bierzychudek and Eckhart 1988; Eppley 2006), also manifest 389 in observed spatial separation of the sexes in a number of species (Bierzychudek and Eckhart 390 1988; Iglesias and Bell 1989; Korpelainen 1991; Shea et al. 1993; Lokker et al. 1994; Eppley 391 et al. 1998; Bertiller et al. 2002; Stark et al. 2005; Dudley 2006). They are likely due to the

392	fact that males and females ultimately still strike a different balance between the benefits of
393	gender specialisation and the risks of mortality. In the context of the paradigm that
394	phenotypes reflect a tension between ecological and sexual components of natural selection
395	(Lande 1980; Slatkin 1984; Shine 1989; Cornwallis and Uller 2010), it seems clear that
396	plasticity invoked by the size-constraint hypothesis for variation in sexual size dimorphism in
397	dioecious plants only partially resolves the tension: males and females still need to reproduce
398	to maintain a stake in the evolutionary game, and this necessity comes with gender-dependent
399	risks.

400

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409

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**Table 1.** Minimal adequate models (LMM; random factors: replicate/pot) for the effects of (a) distance between pots, plant sex and their interaction on plant mass of plant reared in the centre and at the edge of the population, and (b) distance between pots, position in the population and their interaction on the mass of female and male plants. Effect sizes given are untransformed. We present model comparisons (log likelihood ratio tests) by comparing the full model (for a: III: distance + sex + distance × sex; for b: III: distance + position + distance × position ), and simplified models II (for a: position + distance; for b: sex + distance), I (for a: sex; for b: position), I' (for a and b: distance).

### (a) By position

_									
		Centr	е						
		y= plaı	nt mass;	λ = 0.22	2	y= plant	mass;	λ = 0.34	1
	Explanatory variables	Effect size	SE	t	LLRT	Effect size	SE	t	LLRT
	(Intercept: female at distance 0)								
		1.53	0.04	35.37		2.28	0.048	48.02	I-II (Distance):
	Distance 5	0.06	0.06	0.97		0.05	0.062	0.78	lp(χ <sup>2</sup> <sub>5</sub> >42.25)<0.0001
	Distance 15	0.10	0.06	1.71		0.08	0.062	1.25	
	Distance 25	0.20	0.06	3.23		0.28	0.062	4.57	
	Distance 35	0.37	0.06	6.22		0.28	0.062	4.52	
	Distance 85	0.37	0.06	6.11		0.26	0.062	4.15	l'-ll (Sex) :
	Sex (M)	-0.05	0.06	-0.75		-0.23	0.036	-6.37	p(χ <sup>2</sup> <sub>1</sub> >39.21)<0.0001
	Dist. 5 : sex M	-0.02	0.09	-0.23	II-III (Distance x Sex):				
	Dist. 15 : sex M	-0.01	0.08	-0.12	p(χ <sup>2</sup> <sub>5</sub> >12.8)=0.03				
	Dist. 25 : sex M	0.08	0.09	0.99					-   : $p(x^2 > 7.75) = 0.17$
	Dist. 35 : sex M	-0.19	0.09	-2.25					
	Dist. 85 : sex M	-0.11	0.08	-1.26					

### (b) By sex

	Fema	le			Male			
	y= plai	nt mass;	; λ = 0.26	5	y= plant	mass;	λ = 0.30	0
Explanatory variables	Effect size	SE	t	LLRT	Effect size	SE	t	LLRT
(Intercept: Centre								
at distance 0)	1.65	0.05	31.13		1.79	0.05	37.37	I-II (Distance):
Distance 5	0.07	0.07	1.00		0.00	0.06	0.00	p(χ <sup>2</sup> <sub>5</sub> >64.84)<0.0001
Distance 15	0.13	0.07	1.83		0.05	0.06	0.85	
Distance 25	0.25	0.07	3.41		0.32	0.06	5.37	
Distance 35	0.49	0.07	6.61		0.27	0.06	4.55	
Distance 85	0.48	0.07	6.52		0.30	0.06	5.10	I'-II (Position):
Position (E)	0.22	0.07	3.29		0.06	0.04	1.84	p(χ <sup>2</sup> <sub>1</sub> >3.38)=0.07
Dist. 5 : Pos. (E)	0.01	0.09	0.15	II-III (Distance x Position):				
Dist. 15 : Pos. (E)	-0.04	0.09	-0.40	p(χ <sup>2</sup> <sub>5</sub> >31.53)<0.001				
Dist. 25 : Pos. (E)	-0.08	0.09	-0.90					II-III: $p(\chi^2 > 5.08) = 0.41$
Dist. 35 : Pos. (E)	-0.33	0.09	-3.66					
Dist. 85 : Pos. (E)	-0.34	0.09	-3.73					

**Table 2.** Minimal adequate models (LMM; random factors: Replicate/pot) for the effects of (a) distance between pots, plant sex and their interaction on plant height of plant reared in the centre and at the edge of the plots and (b) distance between pots, position in the plot and their interaction on plant mass of female and male plants. We present model comparisons (log likelihood ratio tests), comparing full model (for a: III: distance + sex + distance × sex; for b: III: distance + position + distance × position), and simplified models II (for a: sex + distance; for b: position + distance), I (for a: sex; for b: position), I' (for a and b: distance).

(a) By position

	Centro	e			Edge			
	y= plar	nt height			y= pla	nt heigh	ıt	
Explanatory variables	Effect size	SE	t	LLRT	Effect size	SE	t	LLRT
(Intercept: female at distance 0)	4146.2	123.49	33.57		437.14	8.57	51.01	
Distance 5 Distance 15 Distance 25	-875.3 -1524.7 -1403.0	139.41 139.41 139.79	-6.28 -10.94 -10.04	I-II (Distance): p(χ25>76.9)<0.0001	-71.75 -93.3 -110.65	10.25 10.25 10.27	-7 -9.1 -10.78	l-II(Distance): p(χ² <sub>5</sub> >138.07)<0.0001
Distance 35 Distance 85 Sex (M)	-1330.8 -1595.0 -385.0	139.79 139.41 52.46	-9.52 -11.44 -7.34	l'-II (Sex): p(χ² <sub>1</sub> >46.3)<0.0001	-88.58 -121.52 -39.36	10.25 10.25 3.85	-8.64 -11.85 -10.22	l'-II (Sex): p(χ² <sub>1</sub> >89.08)<0.0001
Dist. 5 : sex M Dist. 15 : sex M Dist. 25 : sex M Dist. 35 : sex M Dist. 85 : sex M				II-III: p(χ <sup>2</sup> <sub>5</sub> >8.19)=0.15				II-III: p(χ <sup>2</sup> <sub>5</sub> >3.16)=0.67

(b) By sex

	Fema	le			Male			
	y= plar	nt height			y= pla	nt heigh	t	
Explanatory variables	Effect size	SE	t	LLRT	Effect size	SE	t	LLRT
(Intercept:								
Centre at								
distance 0)	2387.24	54.50	43.80		513.683	14.91	34.44	
Distance 5	-483.71	62.76	-7.71	I-II (Distance):	-93.84	18.83	-4.98	I-II (Distance):
Distance 15	-708.07	62.76	-11.28	p(χ25>192.2)<0.0001	-161.75	18.83	-8.59	p( <b>x25&gt;193.84)&lt;0.0001</b>
Distance 25	-754.10	62.97	-11.98		-157.20	18.83	-8.35	
Distance 35	-612.04	62.76	-9.75		-166.47	19.01	-8.76	
Distance 85	-866.52	62.76	-13.81		-177.72	18.83	-9.44	l'-II (Position):
Position (E)				I'-II: p(χ <sup>2</sup> <sub>1</sub> >2.60)=0.11	-46.366	16.75	-2.77	p(χ <sup>2</sup> <sub>1</sub> >5.60)=0.02
Dist. 5 : Pos. (E)					7.207	23.51	0.31	
Dist. 15 : Pos. (E)					46.758	23.51	1.99	
Dist. 25 : Pos. (E)				II-III: $p(\chi^2 > 7.83) = 0.17$	25.258	23.55	1.07	II-III: $p(\chi^2 > 9.80) = 0.08$
Dist. 35 : Pos. (E)				1005	59.805	23.65	2.53	
Dist. 85 : Pos. (E)					41.468	23.51	1.76	

**Table 3.** Variation in mass SDI (a) with increasing distance between adjacent pots across the three replicates (A, B and C); mean  $\pm$  SD and for all pots grouped per distance level (Mean  $\pm$  SE and Mean  $\pm$  SD). We separated the data for pots placed either in the center of the plot or at the edge since this reflects differences in competition degree within plot. We present sexual dimorphism in mass without the two extreme outliers (SDI > 20) at the distances 35 and 85 cm (b). We performed a non-parametric Fligner Killeen test for homogeneity of variances, with 5 df for each test. We used non-parametric Kruskal-Wallis test to compare mean SDI when variances were homogeneous (5 df in each test) and Welch's ANOVA (one way) when variances were heterogeneous (df numerator: 5; df denominator: 69.2 and 68.5 for Mass SDI with outliers included and excluded, respectively).

a)	Mass SDI (Cente	er)			All replicates together	
	Distance	Α	В	С	Mean SE S	D
	0	0.27 + 0.72	0.34 + 0.82	0.97 + 1.80	0.52 + 0.24 + 1	.19
	5	$0.23 \pm 0.96$	$1.06 \pm 1.29$	$0.41 \pm 0.48$	$0.52 \pm 0.20 \pm 1$	.01
	15	$0.55 \pm 1.37$	0.71 ± 1.50	$0.81 \pm 1.00$	$0.69 \pm 0.24 \pm 1$	.26
	25	0.81 ± 1.13	0.53 ± 1.25	0.30 ± 1.06	0.54 ± 0.22 ± 1	.12
	35	0.54 ± 1.48	7.05 ± 14.61	3.19 ± 3.23	3.71 ± 1.75 ± 8	.92
	85	5.79 ± 11.80	1.36 ± 2.82	0.75 ± 1.72	2.63 ± 1.38 ± 7	.17
				Fligner Killeen test	y2=18.9; p=0.002	2
				Welch's ANOVA	F=1.09; p=0.37	_
	Mass SDI (Edge)					
	Distance	Α	В	С	Mean SE S	D
	0	0.46 ± 1.02	0.78 ± 1.17	1.05 ± 1.28	0.77 ± 0.17 ± 1	.16
	5	1.02 ± 1.17	1.80 ± 2.11	0.81 ± 1.36	1.20 ± 0.23 ± 1	.61
	15	1.21 ± 1.66	0.94 ± 1.49	1.37 ± 2.49	1.18 ± 0.28 ± 1	.90
	25	0.53 ± 0.69	0.90 ± 1.44	1.20 ± 3.04	0.88 ± 0.29 ± 1	.97
	35	1.07 ± 2.67	2.43 ± 7.28	2.16 ± 5.60	1.88 ± 0.79 ± 5	.39
	85	0.43 ± 1.03	1.16 ± 2.07	1.14 ± 2.16	0.91 ± 0.26 ± 1	.81
				Fligner Killeen test	v2=4 2 · n=0 44	
<b>)</b> )	Mass SDI (witho	out outliers; Cent	er)	Kruskal test	χ2=6.68; p=0.24	ļ
<b>)</b>	Mass SDI (witho	out outliers; Cent A	er) B	Kruskal test	χ2=6.68; p=0.24 Mean SE S	D
<b>)</b>	Mass SDI (witho Distance	out outliers; Cent A 0.27 ± 0.72	er) B 0.34 ± 0.82	C 0.97 ± 1.80	$\frac{\chi^2}{\chi^2=6.68; p=0.24}$ Mean SE Si 0.52 ± 0.24 ± 1	<b>D</b> 19
<b>)</b> )	Mass SDI (witho Distance 0 5	A 0.27 ± 0.72 0.23 ± 0.96	<b>B</b> 0.34 ± 0.82 1.06 ± 1.29	C 0.97 ± 1.80 0.41 ± 0.48	$\frac{1}{\chi^2 = 6.68; p = 0.24}$ $\frac{1}{0.52 \pm 0.24} \pm 1$ $\frac{1}{0.57 \pm 0.20} \pm 1$	<b>D</b> .19 .01
<b>)</b>	Mass SDI (witho Distance 0 5 15	A           0.27 ± 0.72           0.23 ± 0.96           0.55 ± 1.37	<b>B</b> 0.34 ± 0.82 1.06 ± 1.29 0.71 ± 1.50	C 0.97 ± 1.80 0.41 ± 0.48 0.81 ± 1.00	$\begin{array}{c c} Mean & SE & Si \\ \hline 0.52 \pm 0.24 \pm 1 \\ 0.57 \pm 0.20 \pm 1 \\ 0.69 \pm 0.24 \pm 1 \end{array}$	<b>D</b> .19 .01 .26
<b>)</b>	Mass SDI (witho Distance 0 5 15 25	A           0.27         ±         0.72           0.23         ±         0.96           0.55         ±         1.37           0.81         ±         1.13	<b>B</b> 0.34 ± 0.82 1.06 ± 1.29 0.71 ± 1.50 0.53 ± 1.25	C 0.97 ± 1.80 0.41 ± 0.48 0.81 ± 1.00 0.30 ± 1.06	$\begin{array}{c ccccc} Mean & SE & S \\ \hline \\ 0.52 \pm 0.24 \pm 1 \\ 0.57 \pm 0.20 \pm 1 \\ 0.69 \pm 0.24 \pm 1 \\ 0.54 \pm 0.22 \pm 1 \end{array}$	<b>D</b> .19 .01 .26 .12
<b>)</b> )	Mass SDI (witho Distance 0 5 15 25 35	A           0.27         ±         0.72           0.23         ±         0.96           0.55         ±         1.37           0.81         ±         1.13           0.54         ±         1.48	<b>B</b> 0.34 ± 0.82 1.06 ± 1.29 0.71 ± 1.50 0.53 ± 1.25 2.31 ± 3.65	C 0.97 ± 1.80 0.41 ± 0.48 0.81 ± 1.00 0.30 ± 1.06 3.19 ± 3.23	$\begin{array}{c ccccc} Mean & SE & Si \\ \hline 0.52 & \pm 0.24 & \pm 1 \\ 0.57 & \pm 0.20 & \pm 1 \\ 0.69 & \pm 0.24 & \pm 1 \\ 0.54 & \pm 0.22 & \pm 1 \\ 2.06 & \pm 0.61 & \pm 3 \end{array}$	<b>D</b> .19 .01 .26 .12 .04
<b>)</b>	Mass SDI (witho Distance 0 5 15 25 35 85	A           0.27         ±         0.72           0.23         ±         0.96           0.55         ±         1.37           0.81         ±         1.13           0.54         ±         1.48           1.94         ±         2.61	B           0.34         ±         0.82           1.06         ±         1.29           0.71         ±         1.50           0.53         ±         1.25           2.31         ±         3.65           1.36         ±         2.82	C 0.97 ± 1.80 0.41 ± 0.48 0.81 ± 1.00 0.30 ± 1.06 3.19 ± 3.23 0.75 ± 1.72	$\begin{array}{c cccc} Mean & SE & Si \\ \hline Mean & SE & Si \\ \hline 0.52 \pm 0.24 \pm 1 \\ \hline 0.57 \pm 0.20 \pm 1 \\ \hline 0.69 \pm 0.24 \pm 1 \\ \hline 0.54 \pm 0.22 \pm 1 \\ \hline 2.06 \pm 0.61 \pm 3 \\ \hline 1.33 \pm 0.47 \pm 2 \end{array}$	D .19 .26 .12 .04 .37
<b>)</b>	Mass SDI (witho Distance 0 5 15 25 35 85	A           0.27         ±         0.72           0.23         ±         0.96           0.55         ±         1.37           0.81         ±         1.13           0.54         ±         1.48           1.94         ±         2.61	<b>B</b> 0.34 ± 0.82 1.06 ± 1.29 0.71 ± 1.50 0.53 ± 1.25 2.31 ± 3.65 1.36 ± 2.82	C 0.97 ± 1.80 0.41 ± 0.48 0.81 ± 1.00 0.30 ± 1.06 3.19 ± 3.23 0.75 ± 1.72 Fligner Killeen test Weichels ANOVA	$\begin{array}{c c} Mean & SE & Si \\ \hline Mean & SE & Si \\ \hline 0.52 \pm 0.24 \pm 1 \\ \hline 0.57 \pm 0.20 \pm 1 \\ \hline 0.69 \pm 0.24 \pm 1 \\ \hline 0.54 \pm 0.22 \pm 1 \\ \hline 2.06 \pm 0.61 \pm 3 \\ \hline 1.33 \pm 0.47 \pm 2 \\ \chi 2 = 16; p = 0.006 \\ \hline 54 = 0.006 \\$	<b>D</b> .19 .01 .26 .12 .04 .37
<b>)</b>	Mass SDI (witho Distance 0 5 15 25 35 85	A           0.27 ± 0.72           0.23 ± 0.96           0.55 ± 1.37           0.81 ± 1.13           0.54 ± 1.48           1.94 ± 2.61	<b>B</b> 0.34 ± 0.82 1.06 ± 1.29 0.71 ± 1.50 0.53 ± 1.25 2.31 ± 3.65 1.36 ± 2.82	C 0.97 ± 1.80 0.41 ± 0.48 0.81 ± 1.00 0.30 ± 1.06 3.19 ± 3.23 0.75 ± 1.72 Fligner Killeen test Welch's ANOVA	$\begin{array}{c cccc} Mean & SE & Si \\ \hline Mean & SE & Si \\ \hline 0.52 \pm 0.24 \pm 1 \\ \hline 0.57 \pm 0.20 \pm 1 \\ \hline 0.69 \pm 0.24 \pm 1 \\ \hline 0.54 \pm 0.22 \pm 1 \\ \hline 2.06 \pm 0.61 \pm 3 \\ \hline 1.33 \pm 0.47 \pm 2 \\ \hline \chi 2 = 16; p = 0.006 \\ \hline F = 1.58; p = 0.18 \end{array}$	<b>D</b> .19 .01 .26 .12 .04 .37
<b>)</b>	Mass SDI (without of the second secon	A 0.27 ± 0.72 0.23 ± 0.96 0.55 ± 1.37 0.81 ± 1.13 0.54 ± 1.48 1.94 ± 2.61 Dut outliers; Edge	<b>B</b> 0.34 ± 0.82 1.06 ± 1.29 0.71 ± 1.50 0.53 ± 1.25 2.31 ± 3.65 1.36 ± 2.82	C 0.97 ± 1.80 0.41 ± 0.48 0.81 ± 1.00 0.30 ± 1.06 3.19 ± 3.23 0.75 ± 1.72 Fligner Killeen test Welch's ANOVA	$\begin{array}{c cccc} Mean & SE & Si \\ \hline 0.52 \pm 0.24 \pm 1 \\ 0.57 \pm 0.20 \pm 1 \\ 0.69 \pm 0.24 \pm 1 \\ 0.54 \pm 0.22 \pm 1 \\ 2.06 \pm 0.61 \pm 3 \\ 1.33 \pm 0.47 \pm 2 \\ \chi 2 = 16; p = 0.006 \\ F = 1.58; p = 0.18 \end{array}$	<b>D</b> .19 .01 .26 .12 .04 .37
<b>)</b>	Mass SDI (without of the second secon	A 0.27 ± 0.72 0.23 ± 0.96 0.55 ± 1.37 0.81 ± 1.13 0.54 ± 1.48 1.94 ± 2.61 out outliers; Edge A	B 0.34 ± 0.82 1.06 ± 1.29 0.71 ± 1.50 0.53 ± 1.25 2.31 ± 3.65 1.36 ± 2.82 B	C 0.97 ± 1.80 0.41 ± 0.48 0.81 ± 1.00 0.30 ± 1.06 3.19 ± 3.23 0.75 ± 1.72 Fligner Killeen test Welch's ANOVA	$\begin{array}{c ccccc} Mean & SE & Si \\ \hline Mean & SE & Si \\ \hline 0.52 \pm 0.24 \pm 1 \\ \hline 0.57 \pm 0.20 \pm 1 \\ \hline 0.69 \pm 0.24 \pm 1 \\ \hline 0.54 \pm 0.22 \pm 1 \\ \hline 2.06 \pm 0.61 \pm 3 \\ \hline 1.33 \pm 0.47 \pm 2 \\ \chi 2 = 16; p = 0.006 \\ \hline F = 1.58; p = 0.18 \\ \hline \end{array}$	D
<b>)</b>	Mass SDI (without of the second secon	A 0.27 ± 0.72 0.23 ± 0.96 0.55 ± 1.37 0.81 ± 1.13 0.54 ± 1.48 1.94 ± 2.61 A 0.46 ± 1.02	B 0.34 ± 0.82 1.06 ± 1.29 0.71 ± 1.50 0.53 ± 1.25 2.31 ± 3.65 1.36 ± 2.82 B 0.78 ± 1.17	C 0.97 ± 1.80 0.41 ± 0.48 0.81 ± 1.00 0.30 ± 1.06 3.19 ± 3.23 0.75 ± 1.72 Fligner Killeen test Welch's ANOVA C 1.05 ± 1.28	$\begin{array}{c ccccc} Mean & SE & Si \\ \hline 0.52 \pm 0.24 \pm 1 \\ 0.57 \pm 0.20 \pm 1 \\ 0.69 \pm 0.24 \pm 1 \\ 0.54 \pm 0.22 \pm 1 \\ 2.06 \pm 0.61 \pm 3 \\ 1.33 \pm 0.47 \pm 2 \\ \chi^2 = 16; p = 0.006 \\ F = 1.58; p = 0.18 \end{array}$	D .19 .01 .26 .12 .04 .37 D .16
<b>)</b>	Mass SDI (without on the second sec	A         0.27 ± 0.72         0.23 ± 0.96         0.55 ± 1.37         0.81 ± 1.13         0.54 ± 1.48         1.94 ± 2.61	B         0.34 ± 0.82         1.06 ± 1.29         0.71 ± 1.50         0.53 ± 1.25         2.31 ± 3.65         1.36 ± 2.82	C         0.97 ± 1.80         0.41 ± 0.48         0.81 ± 1.00         0.30 ± 1.06         3.19 ± 3.23         0.75 ± 1.72         Fligner Killeen test         Welch's ANOVA	Mean         SE         Si           0.52 $\pm$ 0.24 $\pm$ 1           0.57 $\pm$ 0.20 $\pm$ 1           0.69 $\pm$ 0.24 $\pm$ 1           0.54 $\pm$ 0.22 $\pm$ 1           0.54 $\pm$ 0.22 $\pm$ 1           2.06 $\pm$ 0.61 $\pm$ 3           1.33 $\pm$ 0.47 $\pm$ 2 $\chi$ 2=16; p=0.006         F=1.58; p=0.18           Mean         SE         Si           0.77 $\pm$ 0.17 $\pm$ 1           1.20 $\pm$ 0.23 $\pm$ 1	D .19 .01 .26 .12 .04 .37 D .16 .61
2)	Mass SDI (witho Distance 0 5 15 25 35 85 85 Mass SDI (witho Distance 0 5 15	A 0.27 ± 0.72 0.23 ± 0.96 0.55 ± 1.37 0.81 ± 1.13 0.54 ± 1.48 1.94 ± 2.61 A 0.46 ± 1.02 1.02 ± 1.17 1.21 ± 1.66	B 0.34 ± 0.82 1.06 ± 1.29 0.71 ± 1.50 0.53 ± 1.25 2.31 ± 3.65 1.36 ± 2.82 B 0.78 ± 1.17 1.80 ± 2.11 0.94 ± 1.49	C         0.97 ± 1.80         0.41 ± 0.48         0.81 ± 1.00         0.30 ± 1.06         3.19 ± 3.23         0.75 ± 1.72         Fligner Killeen test         Welch's ANOVA	Mean         SE         Si           0.52 $\pm$ 0.24 $\pm$ 1           0.57 $\pm$ 0.20 $\pm$ 1           0.69 $\pm$ 0.24 $\pm$ 1           0.54 $\pm$ 0.22 $\pm$ 1           0.54 $\pm$ 0.22 $\pm$ 1           0.54 $\pm$ 0.22 $\pm$ 1           2.06 $\pm$ 0.61 $\pm$ 3           1.33 $\pm$ 0.47 $\pm$ 2 $\chi$ 2=16; p=0.006         F=1.58; p=0.18           Mean         SE         Si           0.77 $\pm$ 0.17 $\pm$ 1           1.20 $\pm$ 0.23 $\pm$ 1           1.18 $\pm$ 0.28 $\pm$ 1	D .19 .01 .26 .12 .04 .37 D .16 .61 .90
<b>)</b>	Mass SDI (witho Distance 0 5 15 25 35 85 85 Mass SDI (witho Distance 0 5 15 25	A         0.27 ± 0.72         0.23 ± 0.96         0.55 ± 1.37         0.81 ± 1.13         0.54 ± 1.48         1.94 ± 2.61    out outliers; Edges          A         0.46 ± 1.02         1.02 ± 1.17         1.21 ± 1.66         0.53 ± 0.69	B 0.34 ± 0.82 1.06 ± 1.29 0.71 ± 1.50 0.53 ± 1.25 2.31 ± 3.65 1.36 ± 2.82 B 0.78 ± 1.17 1.80 ± 2.11 0.94 ± 1.49 0.90 ± 1.44	C 0.97 ± 1.80 0.41 ± 0.48 0.81 ± 1.00 0.30 ± 1.06 3.19 ± 3.23 0.75 ± 1.72 Fligner Killeen test Welch's ANOVA C 1.05 ± 1.28 0.81 ± 1.36 1.37 ± 2.49 1.20 ± 3.04	Mean         SE         Si           0.52 $\pm$ 0.24 $\pm$ 1           0.57 $\pm$ 0.20 $\pm$ 1           0.69 $\pm$ 0.24 $\pm$ 1           0.54 $\pm$ 0.22 $\pm$ 1           0.54 $\pm$ 0.22 $\pm$ 1           0.54 $\pm$ 0.22 $\pm$ 1           2.06 $\pm$ 0.61 $\pm$ 3           1.33 $\pm$ 0.47 $\pm$ 2 $\chi$ 2=16; p=0.006         F=1.58; p=0.18           Mean         SE         Si           0.77 $\pm$ 0.17 $\pm$ 1           1.20 $\pm$ 0.23 $\pm$ 1           0.88 $\pm$ 0.29 $\pm$ 1	D .19 .01 .26 .12 .04 .37 D .16 .61 .90 .97
<b>)</b>	Mass SDI (without of the second secon	A 0.27 ± 0.72 0.23 ± 0.96 0.55 ± 1.37 0.81 ± 1.13 0.54 ± 1.48 1.94 ± 2.61 A 0.46 ± 1.02 1.02 ± 1.17 1.21 ± 1.66 0.53 ± 0.69 1.07 ± 2.67	B 0.34 ± 0.82 1.06 ± 1.29 0.71 ± 1.50 0.53 ± 1.25 2.31 ± 3.65 1.36 ± 2.82 B 0.78 ± 1.17 1.80 ± 2.11 0.94 ± 1.49 0.90 ± 1.44 0.58 ± 1.30	C         0.97 ± 1.80         0.41 ± 0.48         0.81 ± 1.00         0.30 ± 1.06         3.19 ± 3.23         0.75 ± 1.72         Fligner Killeen test         Welch's ANOVA         C         1.05 ± 1.28         0.81 ± 1.36         1.37 ± 2.49         1.20 ± 3.04         0.83 ± 1.83	Mean         SE         Si           0.52 $\pm$ 0.24 $\pm$ 1           0.57 $\pm$ 0.20 $\pm$ 1           0.69 $\pm$ 0.24 $\pm$ 1           0.54 $\pm$ 0.22 $\pm$ 1           0.54 $\pm$ 0.22 $\pm$ 1           0.54 $\pm$ 0.22 $\pm$ 1           2.06 $\pm$ 0.61 $\pm$ 3           1.33 $\pm$ 0.47 $\pm$ 2 $\chi$ 2=16; p=0.006         F=1.58; p=0.18           Mean         SE         Si           0.77 $\pm$ 0.17 $\pm$ 1           1.20 $\pm$ 0.23 $\pm$ 1           1.18 $\pm$ 0.28 $\pm$ 1           0.88 $\pm$ 0.29 $\pm$ 1           0.84 $\pm$ 0.30 $\pm$ 2	D .19 .01 .26 .12 .04 .37 D .16 .61 .90 .97 .01
<b>b</b> )	Mass SDI (without of the second secon	A $0.27 \pm 0.72$ $0.23 \pm 0.96$ $0.55 \pm 1.37$ $0.81 \pm 1.13$ $0.54 \pm 1.48$ $1.94 \pm 2.61$ out outliers; Edges          A $0.46 \pm 1.02$ $1.02 \pm 1.17$ $1.21 \pm 1.66$ $0.53 \pm 0.69$ $1.07 \pm 2.67$ $0.43 \pm 1.03$	$B$ $(3.34 \pm 0.82)$ $1.06 \pm 1.29$ $0.71 \pm 1.50$ $0.53 \pm 1.25$ $2.31 \pm 3.65$ $1.36 \pm 2.82$ $(3.36)$ $B$ $(3.78) \pm 1.17$ $1.80 \pm 2.11$ $0.94 \pm 1.49$ $0.90 \pm 1.44$ $0.58 \pm 1.30$ $1.16 \pm 2.07$	C         0.97 ± 1.80         0.41 ± 0.48         0.81 ± 1.00         0.30 ± 1.06         3.19 ± 3.23         0.75 ± 1.72         Fligner Killeen test         Welch's ANOVA         C         1.05 ± 1.28         0.81 ± 1.36         1.37 ± 2.49         1.20 ± 3.04         0.83 ± 1.83         1.14 ± 2.16	Mean         SE         Si           0.52 $\pm$ 0.24 $\pm$ 1           0.57 $\pm$ 0.20 $\pm$ 1           0.57 $\pm$ 0.20 $\pm$ 1           0.69 $\pm$ 0.24 $\pm$ 1           0.54 $\pm$ 0.22 $\pm$ 1           2.06 $\pm$ 0.61 $\pm$ 3           1.33 $\pm$ 0.47 $\pm$ 2 $\chi$ 2=16; p=0.006         F=1.58; p=0.18           Mean         SE         Si           0.77 $\pm$ 0.17 $\pm$ 1           1.20 $\pm$ 0.23 $\pm$ 1           1.8 $\pm$ 0.28 $\pm$ 1           0.88 $\pm$ 0.29 $\pm$ 1           0.84 $\pm$ 0.30 $\pm$ 2           0.91 $\pm$ 0.26 $\pm$ 1	D .19 .01 .26 .12 .04 .37 .04 .37 .04 .37 .01 .90 .97 .01 .81
<b>D</b> )	Mass SDI (without           Distance           0           5           15           25           35           85           Mass SDI (without           Distance           0           5           15           25           35           85	A $0.27 \pm 0.72$ $0.23 \pm 0.96$ $0.55 \pm 1.37$ $0.81 \pm 1.13$ $0.54 \pm 1.48$ $1.94 \pm 2.61$ out outliers; Edge          A $0.46 \pm 1.02$ $1.02 \pm 1.17$ $1.21 \pm 1.66$ $0.53 \pm 0.69$ $1.07 \pm 2.67$ $0.43 \pm 1.03$	$B$ $(3.34 \pm 0.82)$ $1.06 \pm 1.29$ $0.71 \pm 1.50$ $0.53 \pm 1.25$ $2.31 \pm 3.65$ $1.36 \pm 2.82$ $(3.36)$ $($	C         0.97 ± 1.80         0.41 ± 0.48         0.81 ± 1.00         0.30 ± 1.06         3.19 ± 3.23         0.75 ± 1.72         Fligner Killeen test         Welch's ANOVA         C         1.05 ± 1.28         0.81 ± 1.36         1.37 ± 2.49         1.20 ± 3.04         0.83 ± 1.83         1.14 ± 2.16         Fligner Killeen test	Mean         SE         Si           0.52 $\pm$ 0.24 $\pm$ 1           0.57 $\pm$ 0.20 $\pm$ 1           0.57 $\pm$ 0.20 $\pm$ 1           0.69 $\pm$ 0.24 $\pm$ 1           0.54 $\pm$ 0.22 $\pm$ 1           2.06 $\pm$ 0.61 $\pm$ 3           1.33 $\pm$ 0.47 $\pm$ 2 $\chi$ 2=16; p=0.006         F=1.58; p=0.18           Mean         SE         Si           0.77 $\pm$ 0.17 $\pm$ 1           1.20 $\pm$ 0.23 $\pm$ 1           0.84 $\pm$ 0.29 $\pm$ 1           0.84 $\pm$ 0.30 $\pm$ 2           0.91 $\pm$ 0.26 $\pm$ 1	D .19 .01 .26 .12 .04 .37 D .16 .61 .90 .97 .01 .81

**Table 4.** Variation in height SDI with increasing distance between adjacent pots across the three replicates (A, B and C); Mean  $\pm$  SD and for all pots grouped per distance level (mean  $\pm$  SE and mean  $\pm$  SD). We separated the data for pots placed either in the center of the plot or at the edge, since this reflects differences in competition degree within plot. We performed a non-parametric Fligner Killeen test for homogeneity of variances with 5 df for each test. We used a non-parametric Kruskal-Wallis test to compare mean SDI when variances were homogeneous (5 df in each test), and Welch's ANOVA (one way) when variances were heterogeneoius (df numerator: 5; df denominator: 69.5)

Height SDI (Center)									All repli	cates tog	ether
Distance		Α			В			С	Mean	SE	SD
0	0.03	±	0.08	0.05	±	0.08 0.05	±	0.06	0.04	± 0.01	± 0.07
5	0.02	±	0.07	0.07	±	0.10 0.07	±	0.09	0.06	± 0.02	± 0.09
15	0.03	±	0.26	0.08	±	0.14 0.06	±	0.17	0.06	± 0.04	± 0.19
25	0.06	±	0.15	0.09	±	0.16 0.13	±	0.14	0.09	± 0.03	± 0.15
35	0.07	±	0.20	0.23	±	0.21 0.18	±	0.12	0.17	± 0.04	± 0.19
85	0.13	±	0.23	0.07	±	0.14 0.09	±	0.11	0.10	± 0.03	± 0.16
						Fligr	ner	Killeen test	χ2=13	8; p=0.00	3
						We	elch	's ANOVA	F=2.4	45; p=0.04	1
Height SDI (Edge)									All repli	cates tog	ether
Height SDI (Edge) Distance		A			В			с	All repli Mean	cates tog SE	ether SD
Height SDI (Edge) Distance 0	0.10	A ±	0.13	0.10	B ±	0.15 0.05	±	<b>c</b> 0.15	All repli Mean 0.08	cates tog SE ± 0.02	sether SD ± 0.14
Height SDI (Edge) Distance 0 5	0.10 0.11	<b>A</b> ± ±	0.13 0.14	0.10 0.14	<b>B</b> ± ±	0.15 0.05 0.14 0.06	± ±	<b>c</b> 0.15 0.14	All repli Mean 0.08 0.10	<b>SE</b> ± 0.02 ± 0.02	<b>SD</b> ± 0.14 ± 0.14
Height SDI (Edge) Distance 0 5 15	0.10 0.11 0.16	A ± ±	0.13 0.14 0.14	0.10 0.14 0.06	<b>B</b> ± ±	0.15 0.05 0.14 0.06 0.25 0.10	± ± ±	<b>c</b> 0.15 0.14 0.12	All repli Mean 0.08 0.10 0.11	<b>SE</b> ± 0.02 ± 0.02 ± 0.03	<b>sether</b> <b>SD</b> ± 0.14 ± 0.14 ± 0.18
Height SDI (Edge) Distance 0 5 15 25	0.10 0.11 0.16 0.10	A ± ± ±	0.13 0.14 0.14 0.16	0.10 0.14 0.06 0.08	<b>B</b> ± ± ±	0.15 0.05 0.14 0.06 0.25 0.10 0.12 0.11	± ± ±	<b>C</b> 0.15 0.14 0.12 0.17	All repli Mean 0.08 0.10 0.11 0.10	SE           ± 0.02           ± 0.03           ± 0.02	sether           SD           ± 0.14           ± 0.14           ± 0.18           ± 0.15
Height SDI (Edge) Distance 0 5 15 25 35	0.10 0.11 0.16 0.10 0.09	A ± ± ± ±	0.13 0.14 0.14 0.16 0.15	0.10 0.14 0.06 0.08 0.14	B ± ± ± ±	0.15 0.05 0.14 0.06 0.25 0.10 0.12 0.11 0.21 0.10	± ± ± ±	<b>C</b> 0.15 0.14 0.12 0.17 0.16	All repli Mean 0.08 0.10 0.11 0.10 0.11	set           ±         0.02           ±         0.03           ±         0.02	ether <u>SD</u> ± 0.14 ± 0.14 ± 0.18 ± 0.15 ± 0.17
Height SDI (Edge) Distance 0 5 15 25 35 85	0.10 0.11 0.16 0.10 0.09 0.04	A ± ± ± ± ±	0.13 0.14 0.14 0.16 0.15 0.14	0.10 0.14 0.06 0.08 0.14 0.09	B ± ± ± ±	0.15 0.05 0.14 0.06 0.25 0.10 0.12 0.11 0.21 0.10 0.15 0.08	± ± ± ±	C 0.15 0.14 0.12 0.17 0.16 0.13	All repli Mean 0.08 0.10 0.11 0.10 0.11 0.07	SE           ± 0.02           ± 0.03           ± 0.02           ± 0.03           ± 0.03           ± 0.03	sp           ± 0.14           ± 0.14           ± 0.14           ± 0.14           ± 0.14           ± 0.14           ± 0.14           ± 0.14           ± 0.14           ± 0.14           ± 0.15           ± 0.17           ± 0.14
Height SDI (Edge) Distance 0 5 15 25 35 85	0.10 0.11 0.16 0.10 0.09 0.04	A ± ± ± ± ±	0.13 0.14 0.14 0.16 0.15 0.14	0.10 0.14 0.06 0.08 0.14 0.09	<b>B</b> ± ± ± ±	0.15 0.05 0.14 0.06 0.25 0.10 0.12 0.11 0.21 0.10 0.15 0.08 Fligne	± ± ± ±	C 0.15 0.14 0.12 0.17 0.16 0.13 illeen test	All repli Mean 0.08 0.10 0.11 0.10 0.11 0.07 χ2=4	set           ±         0.02           ±         0.02           ±         0.03           ±         0.03           ±         0.02           ±         0.03           ±         0.02           ±         0.03           ±         0.02	sp           ±         0.14           ±         0.14           ±         0.14           ±         0.15           ±         0.17           ±         0.14
Height SDI (Edge) Distance 0 5 15 25 35 85	0.10 0.11 0.16 0.10 0.09 0.04	A ± ± ± ±	0.13 0.14 0.14 0.16 0.15 0.14	0.10 0.14 0.06 0.08 0.14 0.09	<b>B</b> ± ± ± ± ±	0.15 0.05 0.14 0.06 0.25 0.10 0.12 0.11 0.21 0.10 0.15 0.08 Fligne	± ± ± ± ± er K	C 0.15 0.14 0.12 0.17 0.16 0.13 illeen test skal test	All repli           Mean           0.08           0.10           0.11           0.10           0.11           0.10           0.11           0.10           0.11           0.10           0.12           0.13           0.14           0.07           χ2=4           χ2=3	SE           ± 0.02           ± 0.02           ± 0.03           ± 0.03           ± 0.02           ± 0.03           ± 0.03           ± 0.03           ± 0.05; p=0.45	$     ether     SD     \pm 0.14 \pm 0.14 \pm 0.18 \pm 0.15 \pm 0.17 \pm 0.14 5 7$

**Figure 1.** (a) Effect of interplant competition (distance between pots) on dry vegetative mass (g) of male and female *Mercurialis annua*; subsample of all male and female plants reared in the centre of the plot, N = 27 individuals per sex per distance between pots. (b) Effect of position in the plot (center vs. edge) and distance between pots (i.e. interplant competition levels) on plant dry mass separately for male and female plants. Data present mean  $\pm$  SE.

**Figure 2.** (a) Effect of interplant competition (distance between pots) on plant height after eight weeks growth (cm) of male and female *Mercurialis annua*; subsample of all male and female plants reared in the centre of the plot, N = 27 individuals per sex per distance between pots. (b) Effect of position in the plot (center vs. edge) and distance between pots (i.e. interplant competition levels) on plant height separately for male and female plants. Data present mean  $\pm$  SE.

**Figure 3.** Sexual dimorphism in mass (a) and height (b) within pot, for plants reared at the edge or in the centre of the plots. The index of sexual dimorphism (SDI) was calculated as follows: (female trait –male trait)/(male trait9. For mass SDI (a), we present mean  $\pm$  SE for both data with (left panel) and without (right panel) four extreme outliers occurring at the distance between pots of 35 cm (1 central pot, 2 edge pots) and 85 cm (1 central pot).

Figure 1.







Figure 3.

