JOURNAL OF Evolutionary Biology



doi: 10.1111/jeb.12596

Sex-linked inheritance, genetic correlations and sexual dimorphism in three melanin-based colour traits in the barn owl

A. ROULIN* & H. JENSEN†

*Department of Ecology and Evolution, University of Lausanne, Lausanne, Switzerland †Department of Biology, Centre for Biodiversity Dynamics, Trondheim, Norway

Keywords:

birds; quantitative genetics; sexual selection & conflicts.

Abstract

Theory states that genes on the sex chromosomes have stronger effects on sexual dimorphism than genes on the autosomes. Although empirical data are not necessarily consistent with this theory, this situation may prevail because the relative role of sex-linked and autosomally inherited genes on sexual dimorphism has rarely been evaluated. We estimated the quantitative genetics of three sexually dimorphic melanin-based traits in the barn owl (Tyto alba), in which females are on average darker reddish pheomelanic and display more and larger black eumelanic feather spots than males. The plumage traits with higher sex-linked inheritance showed lower heritability and genetic correlations, but contrary to prediction, these traits showed less pronounced sexual dimorphism. Strong offspring sexual dimorphism primarily resulted from daughters not expressing malelike melanin-based traits and from sons expressing femalelike traits to similar degrees as their sisters. We conclude that in the barn owl, polymorphism at autosomal genes rather than at sex-linked genes generate variation in sexual dimorphism in melanin-based traits.

Introduction

Sexual dimorphism refers to traits that are differentially expressed in the two sexes, which often results from selection favouring one sex to express a trait to larger values than the other sex. For instance, sexual selection can promote the evolution of showy male ornaments that confer mating benefits, whereas natural selection can favour camouflage in females (Lande, 1980; Andersson, 1994; Cuervo & Møller, 2000). Males can thus be positively selected to express an ornament to larger values and females selected to display the same trait but to reduced values (Bonduriansky & Chenoweth, 2009). Examples of so-called sexually antagonistic selection can also be found in other, nonornamental traits and across different taxa (e.g. sexual size dimorphism in birds, mammals, insects; Preziosi & Fairbairn, 2000; Lindenfors, 2002; Kruger, 2005). However, in

Correspondence: Alexandre Roulin, Department of Ecology and Evolution, University of Lausanne, Biophore Building, 1015 Lausanne, Switzerland.

Tel.: +41 21 692 4189; fax: +41 21 692 4165; e-mail: Alexandre.Roulin@unil.ch

scenarios where selection acts differently in males and females, evolutionary change in trait expression is largely constrained because both sexes share most of their genome. When the genetic correlation between the sexes for an ornament is strong, selection exerted on one sex to be ornamented will affect the evolution of the ornament not only in this sex, but also in the other sex (Lande, 1987; Rhen, 2000; Poissant et al., 2009). Selection exerted on genetically correlated phenotypic traits will have similar constraining evolutionary effects. Indeed, counter-selected alleles encoding a sexually dimorphic trait can be maintained in a population owing to their positive effects on genetically correlated traits (Lande, 1980; Chenoweth et al., 2008; Kirkpatrick, 2009; Gosden et al., 2012). For instance, in the European kestrel (Falco tinnunculus) the degree of melanin-based coloration, a sexually dimorphic trait, is genetically correlated with body mass (Kim et al., 2013), implying that selection on body mass can affect the evolution of sexual dimorphism. The situation can be even more complex when selection exerted on traits in females constrains the evolution of the sexually dimorphic trait in males due to the genetic correlations among homologous and nonhomologous traits across

sexes (Jensen *et al.*, 2008; Poissant *et al.*, 2009; Harano *et al.*, 2010). The allocation of genes on the sexual chromosomes may have evolved to alleviate this situation, as it is expected to decrease genetic correlations between the sexes and thereby allow each sex to express phenotypes to their optimal value. However, the relative role of polymorphisms at autosomally and sex-linked genes on sexual dimorphism (Reinhold, 1998) has rarely been considered in natural populations (Roulin *et al.*, 2010; Husby *et al.*, 2012; Larsen *et al.*, 2014). This is a critical issue because, for instance, sex-linked and autosomally inherited components of phenotypic variation are not similarly sensitive to selection (Rice, 1984).

Sex-linked genes control the expression of secondary sexual characters because these genes can be expressed to higher levels in the homogametic than heterogametic sex (e.g. in birds, males are homogametic ZZ and females heterogametic ZW) (Naurin *et al.*, 2009). The exact role played by sex chromosomes and autosomes

on sexual dimorphism is currently debated (Dean & Mank, 2014), further justifying studies on the quantitative genetics of sexual dimorphism. Depending on the genetic architecture, offspring sexual dimorphism can take several forms. The simplest situation is when only males express a secondary sexual character. In that case, the extent of offspring sexual dimorphism can depend on whether sons inherit alleles encoding an exaggerated or a small version of the sexually dimorphic trait (Fig. 1a). When the two sexes express the secondary sexual trait, the extent of offspring sexual dimorphism may vary depending on the degree of resemblance between parents and their sons and daughters. When the resemblance between parents and sons is identical to the resemblance between parents and daughters (Fig. 1b), the degree of sexual dimorphism (i.e. the lower trait expression in female offspring compared to their brothers) is of similar magnitude across families (value α in Fig. 1b). When the lower trait expression in females differs between

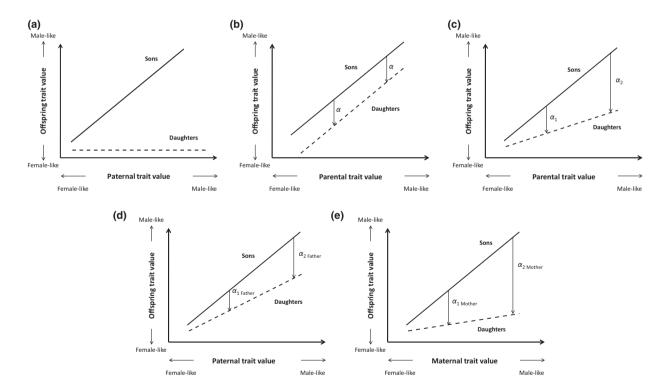


Fig. 1 Sons' and daughters' expression of a secondary sexual character in relation to parental trait values. In these examples, males express the sex trait to larger mean values than females, and hence, large values are denoted 'malelike' and low values 'femalelike'. (a) Complete sexual dimorphism (or sex-limited dimorphism): only males express the secondary sexual character, and in this case, males can express different values of the trait, whereas all females display the same 'femalelike trait'. (b–d) The dimorphism is not sex limited, and offspring resemble each of their parents to varying degrees. (b) Sons and daughters similarly resemble their parents, but females express the trait to lower values than males (by a value α). (c) Sons and daughters resemble their parents but to different values; females express the trait to lower values than males, but the extent of the decreased expression (values α_1 and α_2) is correlated with parental trait value. (d, e) The values α_1 and α_2 can be different when comparing offspring phenotype with paternal (d) and maternal (e) phenotype. These phenotypic models cannot make any assumptions or predictions regarding the underlying genetic basis.

families, there might be specific situations where male offspring closely resemble their sisters (in Fig. 1c when parents display a femalelike version of the sexually dimorphic trait) and other situations where brothers and sisters are clearly different (in Fig. 1c when parents display a malelike version of the trait). These effects may, furthermore, differ if we consider mothers and fathers (Fig. 1d,e), for instance if polymorphic genes encoding sexually dimorphic traits are located on the sex chromosomes. In birds, for example, mothers pass on their single Z sex chromosome to sons but not to daughters, whereas males transmit a copy of the Z sex chromosome in both sons and daughters. Because genes located on sex chromosomes are predicted to result in higher phenotypic differences between males and females, we can expect a correlation between the degree to which phenotypic traits are coded by sexlinked genes and the degree of sexual dimorphism. Surprisingly, the current evidence for this prediction is rather limited (Dean & Mank, 2014).

Recently, quantitative genetic tools have been developed to estimate the extent to which polymorphism at genes located on the sex chromosomes and autosomes explain variation in phenotypic traits (Roulin et al., 2010; Husby et al., 2012; Larsen et al., 2014; Evans et al., 2014). This offers the possibility to examine the prediction that sexual dimorphism is positively associated with the extent to which polymorphism at sexlinked genes participate in the expression of the sexually dimorphic trait (Dean & Mank, 2014). To this end, we considered the barn owl (Tyto alba) because this bird displays three sexually dimorphic melanin-based plumage traits, with at least one trait showing sex-linked inheritance (Roulin et al., 2010; Larsen et al., 2014). Although members of the two sexes can express any phenotype, females are on average darker reddish than males (pheomelanin-based colour trait) and display more and larger black spots located on the feather tips (two eumelanin-based colour traits) (Roulin et al., 2001). These three melanin-based colour traits are genetically correlated to a different extent in the two sexes (darker birds display more and larger black spots, particularly males) (Roulin et al., 2001; Roulin & Dijkstra, 2003). Because the three colour traits are already expressed in nestlings, we can perform powerful quantitative genetic analyses and compare the level of offspring sexual dimorphism with parental plumage traits. Our aim is therefore to measure the degree of offspring sexual dimorphism, estimate Z-linked and autosomal components of phenotypic variation, measure the phenotypic and genotypic correlations among traits within males and females, and measure the phenotypic and genotypic correlation between homologous and nonhomologous traits among sexes. In traits for which sexlinked genes have a strong effect on trait expression, we predict heritabilities and genetic correlations to be lower (because females have only one copy of the Z sex chromosome) but sexual dimorphism to be more pronounced than in traits for which the Z-linked component of phenotypic variation is lower.

Materials and methods

Study organism

The worldwide distributed barn owl shows pronounced variation in the expression of the three melanin-based colour traits (reddish pheomelanic coloration and number and size of eumelanic black spots; Roulin et al., 2009). Although members of the two sexes can express these three heritable phenotypes in the range of any possible values, females are on average darker reddish and display on average more and larger black spots than males (Roulin et al., 2001). Spot size is positively selected in females and negatively selected in males, the magnitude of sexually antagonistic selection being population specific (Roulin et al., 2010, 2011). Spot size is associated, particularly in females, with numerous phenotypic attributes such as growth, appetite and resistance to various stressful factors including free radicals, pathogens and predators (Roulin & Ducrest, 2011; Van den Brink et al., 2012). The number of spots is associated with thermoregulation and sibling competition (A. Roulin, unpublished) and pheomelanin-based coloration is involved in foraging, with differently coloured individuals being adapted to different ecological conditions (Roulin, 2004a; Charter et al., 2012; Dreiss et al., 2012). Extra-pair paternity is rare in this species (Henry et al., 2013).

General method

Between 1996 and 2010, we studied barn owls in western Switzerland (46°49′N/06°56′E) in an area of 190 km² where 196 nest boxes were available. Nestling sex was identified using sex-specific molecular markers (Roulin *et al.*, 1999), whereas breeding females were distinguished from breeding males by the presence of a brood patch. Age of the breeding birds was known precisely if ringed as nestlings in previous years. For other individuals, we estimated age based on moult pattern (Taylor, 1993).

Melanin-based traits were reliably recorded (Roulin, 1999, 2004b). As feathers of each body part are similarly coloured, a single person (AR) compared pheomelanin-based coloration of the breast, belly, flank and underside of the wings with eight chips ranging from –VIII for white to –I for reddish. A mean value over the four body parts was calculated. A 60 × 40 mm² frame was then placed on the breast and black spots were counted, and their diameter measured to the nearest 0.1 mm. Mean spot diameter was used in the statistical analyses. At the age of 45 days, we can already record pheomelanin-based coloration, but not

yet count and measure black spots, which were measured at 50–55 days. Thus, in a few cases, we could record only one plumage trait if these individuals disappeared from their nest (either because they died or left the nest prematurely) before we could record the other plumage traits. This explains discrepancies in sample sizes between plumage traits.

Offspring sexual dimorphism

From 1996 to 2010, plumage traits were measured in the two parents and in at least one daughter and one son in 431 families. This represents a sample of 1099 female nestlings and 1121 male nestlings, 259 different breeding males and 309 different breeding females. Because environmental effects on the expression of melanin-based traits are very weak in the barn owl (Roulin & Dijkstra, 2003; Roulin et al., 2010), we pooled all offspring produced by a given pair in several broods produced in the same (barn owls can produce two annual broods) or different years giving a total of 400 families. We thus calculated mean daughters' and mean sons' pheomelanin-based reddish coloration, number of black spots and spot size in pooled families of 400 different breeding pairs. To compare the relative degree of sexual dimorphism in the three melaninbased plumage traits, we standardized plumage traits across the two sexes and the 400 different families (i.e. to calculate mean and standard deviations, we had 800 values for each plumage trait). For each plumage trait and family, offspring sexual dimorphism was defined as 'standardized daughter value - standardized son value'. Thus, larger values of offspring sexual dimorphism indicate that daughters are darker reddish than sons, or that daughters display more or larger black spots than sons. Note that if offspring sexual dimorphism is defined as 'daughters' value/sons' value', results are qualitatively similar. However, dimorphism defined as a difference rather than a ratio is more appropriate because ratios obtained with very small denominators tend towards infinity.

Statistical procedure

Animal model to derive quantitative genetic parameters We used an estimate of the 1-year size of traits for all birds (fledglings and adults) because in a previous study, we showed that melanin-based plumage traits change with age in a sex-specific way (Dreiss & Roulin, 2010). This was performed by estimating the relationship between age, in years, and trait size for all birds. In these models, we allowed for differences between sexes, effects of age, age², interactions sex*age and sex*age². Each of these parameters was included in the final model if *P*-values were smaller than 0.10. We then reran the model when also identity was included as fixed factor to derive individual intercepts because

we often captured each individual in more than 1 year. The relationships found between age and plumage traits in these models were used to estimate 1-year trait size for all birds by summing the overall intercept of the model, the individual's intercept and the sex-specific effect. To this value, we then added the (sex-specific) change in trait size from age 0 (i.e. intercept trait size) to age 1 as given by the slope(s) for age, age², sex*age and/or sex*age². For birds measured as fledglings, the hatch year was known. For other birds, we used estimated age at first breeding to calculate their year of hatching.

Animal models are general mixed models, which utilize information from individuals with different levels of relatedness (i.e. not only parent-offspring) in a pedigree to estimate quantitative genetic quantities and various environmental effects (e.g. Lynch & Walsh, 1998; Kruuk & Hill, 2008; Charmantier et al., 2014). In the animal models, we estimated the proportion of variance due to differences among years (because this may explain some proportion of the trait variance; Roulin et al., 2010), and environmental maternal effects by including year and maternal identity as random factors, respectively. Birds from the same brood were assigned the same unique dummy mother ID's if their mother was missing, and birds without nest information were also assigned unique dummy mother ID's. Maternal environmental effects may be particularly important to control for in analyses of sex-chromosomal inheritance (Fairbairn & Roff, 2006). To avoid any bias due to differences in means and/or variance of traits between males (at least one plumage trait was measured in 1703 individuals) and females (at least one trait measured in 1922 individuals), trait sizes were standardized within each sex to have a phenotypic mean of 0 and phenotypic variance of 1 when estimating quantitative genetic parameters within and across sexes. The pedigree consisted of 4343 individuals, 2065 males and 2278 females. Both parents were known for 70.4% of the individuals in the pedigree, only the mother for 5.1% and only the father for 0.3%. 1047 individuals (i.e. 24.1%) in the pedigree had no known parents. Among the 334 individuals born before 1996 only 6 had one or two known parents. In contrast, among every cohort born 1996-2011 at least one parent was known for on average 80% of the individuals (range: 54-100%).

Additive genetic (co)variances, heritabilities and genetic correlations of reddish pheomelanin-based coloration, number and diameter of black spots were estimated by implementing a restricted maximum-likelihood animal model using the VCE6 software (Neumaier & Groeneveld, 1998; Groeneveld *et al.*, 2010). To estimate intra- and intersexual additive genetic (co) variances, and corresponding heritabilities and genetic correlations, we assumed that males and females represented two different environments and that each

homologous trait in the two sexes in reality consists of two separate traits, one of which is expressed only in males and one of which is expressed only in females. Hence, male traits are missing in females and vice versa. This is equivalent to estimating the additive genetic variances and covariances within and across two environments, which in our case are the two sexes (Roff, 1997).

To test whether heritability estimates were significantly different from zero and whether estimates of genetic correlations between traits were significantly different from each other, or from zero or one, we calculated *z*-scores that were tested against a large sample standard normal distribution, following the procedure outlined in Jensen *et al.* (2003). Unfortunately, a likelihood ratio test cannot be carried out in VCE because the likelihood value calculated by VCE is different from the real likelihood as only the part of the likelihood required for optimization is computed (Groeneveld *et al.*, 2010).

To examine whether any of the phenotypic variance observed in the reddish pheomelanin-based coloration, number and diameter of black spots was due to genes located on the Z-chromosomes (see Roulin et al., 2010; Larsen et al., 2014), we estimated autosomal and Zchromosomal additive genetic variances using Bayesian animal models and the INLA framework (Steinsland & Jensen, 2010; Holand et al., 2013; Larsen et al., 2014). These Bayesian animal models can currently only be used for single-trait models and could hence not be used to estimate additive genetic covariances within and across sexes. In the Bayesian animal models, we followed the recommendations of Larsen et al. (2014): we standardized the data across both sexes instead of within each sex as above, we present the Z-chromosomal additive genetic variance for males, and we regard Z-chromosomal additive genetic variance to be present if the model with both Z- and autosomal additive genetic variance has a DIC which is at least 10 units lower than the model with only autosomal inheritance.

Relationship between offspring sexual dimorphism and parental phenotypes

We examined whether the degree of offspring sexual dimorphism is associated more strongly with maternal or paternal phenotypes. Offspring sexual dimorphism in pheomelanin-based coloration, number of spots and spot diameter were entered as dependent variables in separate linear mixed models, where the identities of the two biological parents were entered as random variables. Six independent variables were simultaneously introduced in the models, that is standardized (mean = 0, SD = 1) maternal and paternal pheomelanin-based coloration and number and size of black spots. Nonsignificant variables were backward removed starting with the least significant ones (threshold level was 0.05). However, all significant variables in the reduced model were already significant in the initial model.

Analyses on sexual dimorphisms were carried out with the software JMP (version no. 8; SAS software, Inc., Cary, NC, USA). All tests were two tailed and *P*-values smaller than 0.05 considered significant.

Results

Quantitative genetics

For sexes combined, pheomelanin-based coloration was more strongly heritable ($h^2 = 0.836$) than spot diameter ($h^2 = 0.668$) and number of spots ($h^2 = 0.570$) (Table 1). Hatch year explained approximately 8% and 6% of the variation in number and diameter of spots, respectively, but explained none of the colour variation (Table 1). Mother identity explained between 1% and 5% of phenotypic variation (Table 1). When sexes were analysed separately, we found a tendency that the heritability was higher in males than in females (Table 1); this difference was, however, not significant for any of the three traits (P = 0.65, P = 0.13, P = 0.09 for pheomelanin-based colour, number and diameter of spots, respectively). These patterns nevertheless support

Table 1 Quantitative genetics of plumage traits in the barn owl. Estimates of the proportion of variation in pheomelanin-based coloration, number and diameter of black spots due to additive genetic effects (i.e. the heritability, h^2), maternal environmental effects and year of hatching. Estimates \pm SE are from a model where both sexes and all three traits were included. We first assumed sexes to be of the same environment, estimates for sexes combined. We also report heritability of plumage traits of male and female barn owls separately. These estimates are from a model where males and females were assumed to be two different environments, that is that females had missing values for male traits and vice versa. Both male and female traits were included in the same model. Variation in trait sizes due to any effects of year of hatching and maternal environmental effects were accounted for.

	Variance components – sexes combined			h^2 – sexes separately		
Trait	Hatch year	Mother ID	h ²	Males	Females	
Pheomelanin-based colour Number of black spots Diameter of black spots	0.005 ± 0.003 0.077 ± 0.021 0.055 ± 0.016	0.012 ± 0.011 0.040 ± 0.012 0.052 ± 0.013	0.836 ± 0.021 0.570 ± 0.030 0.668 ± 0.025	0.872 ± 0.124 0.716 ± 0.136 0.805 ± 0.032	0.793 ± 0.125 0.500 ± 0.040 0.576 ± 0.132	

Bayesian single-trait animal models showing significant Z-chromosomal additive genetic variance for both number and diameter of spots but not for pheomelanin-based coloration (Table 2).

The three plumage traits were positively correlated within males and females, both phenotypically and genetically (Table 3a). The genetic correlations were more strongly positive within males than within females (P < 0.0001, P = 0.06, P < 0.0001 for pheomelanin-based colour, number and diameter of spots, respectively; Table 3a). Although phenotypic correlations were less strong than their associated genetic

correlation, they were highly correlated (Spearman's correlation: $r_s = 1$, n = 6, P < 0.0001) (Table 3a), which is not surprising given that heritabilities were very high (Table 1, see also Hadfield *et al.*, 2007).

The genetic correlations between sexes for homologous traits were very strong and only significantly lower than 1 for number of spots (P = 0.002, values in the diagonal of Table 3b). Genetic correlations between males and females for nonhomologous traits ranged from 0.145 for male pheomelanin-based colour and female number of spots up to 0.841 for male number of spots and female diameter of spots, and

Table 2 Autosomal and Z-chromosomal inheritances of three melanin-based plumage traits in the barn owl. Additive genetic variances were calculated in one model assuming autosomal and Z-linked inheritances of the traits and another model assuming autosomal inheritance only. Z-chromosomal additive genetic variance is given for males (i.e. approximately twice that of females). Additive genetic variance estimates (Va) are posterior means, and credible intervals are given in parentheses.

	Autosomal Va	Z-chromosomal Va	Hatch year Vy	Maternal identity Vm	Residual Vr	DIC
Pheomelanin coloration	0.524 (0.453; 0.607) 0.651 (0.593; 0.715)	0.169 (0.104; 0.250) -	0.003 (0.001; 0.008) 0.003 (0.001; 0.009)	0.003 (0.000; 0.011) 0.003 (0.000; 0.011)	0.136 (0.110; 0.167) 0.135 (0.108; 0.165)	5618.161 5586.007
Number of black spots	0.154 (0.081; 0.272) 0.548 (0.476; 0.628)	0.511 (0.411; 0.625) -	0.053 (0.025; 0.101) 0.059 (0.028; 0.116)	0.042 (0.021; 0.074) 0.035 (0.014; 0.065)	0.320 (0.278; 0.365) 0.306 (0.262; 0.355)	7845.770 7895.389
Diameter of black spots	0.224 (0.161; 0.300) 0.544 (0.481; 0.615)	0.454 (0.365; 0.563) -	0.041 (0.018; 0.084) 0.046 (0.020; 0.093)	0.030 (0.012; 0.059) 0.035 (0.014; 0.068)	0.206 (0.171; 0.248) 0.208 (0.169; 0.254)	6664.644 6758.495

The model with both autosomal and Z-chromosomal inheritances is regarded as significantly better if its DIC is > 10 lower than DIC for the autosomal model only (Larsen *et al.*, 2014). Hatch year and maternal identity were included in the models as random effects, and sex was included as fixed effect. The best model for each trait is given in bold.

Table 3 Phenotypic and genetic correlations between plumage traits (pheomelanin-based coloration, number and diameter of black spots) within males (above diagonal, in bold) and females (below diagonal) (a), and across-sex genetic correlations between homologous and nonhomologous plumage traits in the barn owl (b). All phenotypic correlations are significant at P < 0.001. Estimates of genetic correlations (\pm SE) are from a model where males and females were assumed to be two different environments, that is females had missing values for male traits and vice versa. All male and female traits were included in the same model. Variation in trait sizes due to any effects of year of hatching and maternal identity was accounted for.

	Phenotypic correlation			Genetic correlation		
	Pheomelanin colour	Number of black spots	Diameter of black spots	Pheomelanin colour	Number of black spots	Diameter of black spots
Pheomelanin colour	_	0.369	0.418	-	0.501 ± 0.023	0.557 ± 0.067
Number of black spots	0.182	_	0.713	0.194 ± 0.041	_	0.926 ± 0.028
Diameter of black spots	0.332	0.527	_	0.380 ± 0.068	0.661 ± 0.037	_

(b) Genetic correlations across sexes in homologous and nonhomologous traits

Female trait

		Pheomelanin colour	Number of black spots	Diameter of black spots
Male trait	Pheomelanin colour Number of black spots Diameter of black spots	0.996 ± 0.014 0.543 ± 0.026 0.591 ± 0.025	0.145 ± 0.035 0.903 ± 0.031 0.710 ± 0.035	0.346 ± 0.041 0.841 ± 0.053 0.963 ± 0.019

Estimates in the diagonal refer to the genetic correlations between homologous plumage traits in related males and females (e.g. pheomelanin-based coloration in females vs. males [0.996 \pm 0.014]). Off-diagonal elements refer to the genetic correlations between nonhomologous plumage traits in males and females (e.g. pheomelanin-based coloration in females vs. number of black spots in males [0.543 \pm 0.026]).

were significantly higher than 0 and lower than 1 (all *P*-values < 0.0027, values off the diagonal in Table 3b).

Offspring sexual dimorphism

Offspring sexual dimorphism was more pronounced in pheomelanin-based coloration than in the size of eumelanic spots (mean standardized values \pm SE: 1.03 \pm 0.04 vs. 0.74 ± 0.05 ; paired *t*-test: $t_{399} = 6.07$, P < 0.0001), which was itself more pronounced than offspring sexual dimorphism in the number of eumelanic spots $(0.54 \pm 0.05; paired t-test: t_{339} = 4.70, P < 0.0001)$. In 367 of the 400 different breeding pairs (91.8%), sisters were on average darker pheomelanic than their brothers (sign test comparing sexual dimorphism with 0, M = 158.50, P < 0.0001); in 313 families (78.3%), sisters displayed on average larger black spots than their brothers (M = 113.50, P < 0.0001); and in 267 families (66.8%), sisters displayed on average more black spots than their brothers (M = 67.50, P < 0.0001). In 229 families (57.3%), sisters were simultaneously on average darker pheomelanic and displayed more and larger eumelanic spots than their brothers (note that in some families, females can be darker reddish than their brothers but not necessarily more spotted). Offspring sexual dimorphism in pheomelanin-based coloration was strongly correlated with offspring sexual dimorphism in both number and size of black spots, that is when sisters were darker reddish than their brothers, they also displayed more black spots (Pearson's correlations: r = 0.48, n = 400 families, P < 0.0001) and larger spots (r = 0.45, P < 0.0001). The correlation between offspring sexual dimorphism in both eumelanic traits (i.e. number and size of spots) was stronger than the relationships with offspring sexual dimorphism in the degree of pheomelanism (r = 0.64, P < 0.0001; when sisters displayed more black spots than their brothers, these spots were also larger). This is consistent with the genetic correlation analyses (Table 3).

Relationship between offspring sexual dimorphism and parental phenotypes

Standardized offspring sexual dimorphism was more often and more strongly associated with standardized maternal than paternal plumage traits (Table 4, Fig. 2). Furthermore, offspring sexual dimorphism in pheomelanin-based coloration was more strongly related to the homologous trait (i.e. pheomelanin-based coloration) of their parents than it was for number and size of black spots (Table 4, Fig. 2).

Offspring sexual dimorphism in pheomelanin-based coloration increased (i.e. daughters were darker reddish than sons) particularly when their mother and father were pale rather than dark reddish (Figs 2 and 3a) and when their mother displayed small rather than large black spots (Table 4). Similarly, daughters displayed more black spots than sons particularly when their mother was pale rather than dark reddish and when the mother and father displayed small rather than large black spots (Table 4; Figs 2 and 3b). Finally, daughters displayed larger black spots than sons when their mother was light rather than dark reddish and when the mother exhibited small rather than large black spots (Table 4; Figs 2 and 3c).

Table 4 Offspring sexual dimorphism in melanin-based traits (pheomelanin-based coloration, number and size of eumelanic spots) in relation to parental plumage phenotypes in the barn owl. Linear mixed models included the identity of the two biological parents as random variables and the three standardized paternal and maternal plumage traits as six independent variables. Nonsignificant terms are eliminated starting with the least significant ones. Significant terms are written in bold and estimates (\pm SE) are given. Sample size is 400 different breeding pairs. Offspring sexual dimorphism was defined as the difference between standardized daughters' and sons' plumage trait values. Negative estimates indicate a reduced offspring sexual dimorphism when parents are more melanic.

	Offspring sexual dimorphism in standardized melanin-based plumage traits			
	Pheomelanin-based colour	Number of black spots	Diameter of black spots	
Standardized paternal plumag	ge traits			
Pheomelanin colour	$F_{1,184.5}$ = 13.72, P = 0.0003 (-0.171 \pm 0.046)	$F_{1,230.8}$ = 0.007, $P = 0.93$	$F_{1,201.4} = 0.005, P = 0.94$	
Number of black spots	$F_{1,203} = 0.58, P = 0.45$	$F_{1.136} = 0.48, P = 0.49$	$F_{1,242,7} = 0.11, P = 0.74$	
Diameter of black spots	$F_{1,164.6}$ = 0.03, P = 0.86	$F_{1,158}$ = 4.19, P = 0.04 (-0.081 \pm 0.039)	$F_{1,170.3}$ = 0.09, P = 0.76	
Standardized maternal plumage	ge traits			
Pheomelanin colour	$F_{1,285.3}$ = 24.16, P < 0.0001 (-0.238 \pm 0.048)	$F_{1,274.6}$ = 24.49, P < 0.0001 (-0.223 \pm 0.045)	$F_{1,310.2}$ = 17.49, P < 0.0001 (-0.203 \pm 0.048)	
Number of black spots	$F_{1,305}$ = 0.90, P = 0.34	$F_{1,293}$ = 0.88, P = 0.35	$F_{1,319}$ = 1.23, $P = 0.27$	
Diameter of black spots	$F_{1,175.3}$ = 14.05, P = 0.0002 (-0.183 \pm 0.049)	$F_{1,187.1}$ = 86.53, P < 0.0001 (-0.429 \pm 0.046)	$F_{1,251.6}$ = 54.45, P < 0.0001 (-0.370 \pm 0.050)	

For instance, the degree of offspring sexual dimorphism in pheomelanin-based coloration is negatively correlated with paternal and maternal pheomelanin-based coloration and the size of maternal black spots.

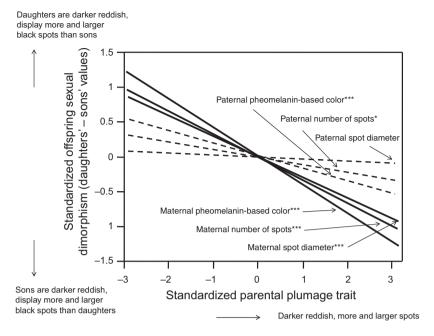


Fig. 2 Relationship between sexual dimorphism in offspring plumage traits in relation to parental plumage traits in the barn owl. Offspring' mean plumage trait values were standardized across the 400 different breeding pairs, and for each pair, daughters' standardized value was subtracted from the sons' standardized value so that sexual dimorphism in the three plumage traits had the same scale and could be compared. Parental plumage traits were also standardized within each sex. For each of the three plumage traits (i.e. pheomelanin-based coloration, number and size of black spots), standardized offspring sexual dimorphism was regressed on the standardized homologous trait value of their mother and father, separately. For instance, the line 'Maternal spot diameter' is for the relationship between standardized offspring sexual dimorphism in spot diameter and maternal spot diameter. Significant regressions are indicated with the symbols ***P < 0.0001 and *P < 0.05.

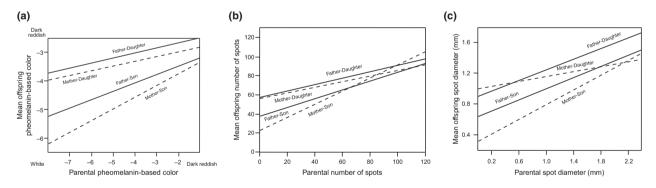


Fig. 3 Relationship between mean offspring and parental pheomelanin-based coloration (a), mean offspring and parental number of spots (b), and offspring and parental spot diameter (c) in the barn owl. Regression lines are based on 400 different breeding pairs.

Discussion

We identified the determinants of offspring sexual dimorphism in three heritable melanin-based traits in the barn owl. Quantitative genetic analyses showed that the three traits are heritable and genetically correlated within and across sexes (Tables 1–3; Figs 2 and 3). As expected, the trait with the lowest Z-linked component, reddish pheomelanin-based coloration, showed the highest heritability and genetic correlation between the sexes (Tables 1–3). In contrast to prediction, this trait was a more strongly sexually dimorphic trait compared to

eumelanin-based plumage traits (i.e. number and size of black spots), which showed a significant Z-linked component (Table 2; Fig. 3). Another important result is that the degree to which daughters are more pigmented (i.e. display a darker reddish coloration, more or larger black spots) than their brothers increases when their parents are less pigmented (Table 4, Fig. 2).

Heritability

Heritabilities tended to be smaller in female than in male barn owls (Table 1; Fig. 3), a situation that seems to contrast with other organisms (e.g. Jensen et al., 2003). The cause is probably that two of the plumage traits are partially encoded by polymorphic genes located on the Z sex chromosome (Table 2). As females do not pass on their Z to daughters, this might reduce the magnitude of the heritability. Up to date, few researchers have demonstrated that genes located on sex chromosomes are responsible for variation in colour traits using Mendelian genetics (Southern, 1946; Munro et al., 1968; Zann, 1996), parent-offspring regression (Potti & Canal, 2011) or animal models (Roulin et al., 2010; Husby et al., 2012; Evans et al., 2014; Larsen et al., 2014). A recent study in collared flycatchers (Ficedula albicollis) and zebra finches (Taenipygia guttata) has suggested that sexually selected traits are not more often encoded by polymorphic genes located on sex chromosomes than other morphological traits (Husby et al., 2012), but rather the expression of genes underlying sex-specific phenotypes could be controlled by sex hormones (something that is not taken into account in animal models). Although theory postulates that sex chromosomes play an important role in the evolution of sexual dimorphism, as their transmission is sex biased or sex limited (Rice, 1984), many sexually dimorphic traits are encoded by polymorphic genes located on autosomes (Mank, 2009; Mank & Ellegren, 2009).

In the barn owl, variation in the most sexually dimorphic plumage trait (pheomelanin-based coloration) did not show any significant Z-component in contrast to the two least sexually dimorphic traits (number and size of black spots) (Table 2). This suggests that sex-linked inheritance is not a prerequisite for sexual dimorphism. For both number and size of black spots, the slopes of the resemblance between father and sons and between father and daughters were of similar magnitude but not the intercepts (Fig. 3b,c). In contrast, the resemblance mother-sons was more pronounced than mother-daughters for the three plumage traits (Fig. 3) probably because the maternal Z chromosome is transmitted only to sons. As a consequence, the intensity of offspring sexual dimorphism was reduced when mothers displayed many and large black spots and when they were darker reddish (Table 4; Fig. 2), as they produced similarly spotted sons and daughters (absence of sexual dimorphism). This suggests that, in contrast to intuition (Rice, 1984; Mank, 2009), polymorphism at genes located on sex chromosomes does not necessarily increase the degree of offspring sexual dimorphism because at some specific parental trait values, parents produce similarly plumaged sons and daughters (Fig. 3).

Genetic correlations between plumage traits within sexes

The evolution of a given phenotype can result from selection being exerted on it directly but also on

genetically correlated traits, as shown for the specific case of secondary sexual characters (Brooks & Endler, 2001; Jensen et al., 2008; Poissant et al., 2008). Genetic correlations between traits can evolve because selection exerted on the different traits is correlated (McGlothlin et al., 2005; Roff & Fairbairn, 2012). Correlational selection implies that functionally related traits should not be expressed independently from each other because individuals derive more fitness benefits from some specific trait combinations than other combinations. For instance, alternative foraging modes may require different combinations of characters (Sinervo & Svensson, 2002), for example foraging upon a given type of prey may necessitate a particular coloration to be cryptic which in turn requires specific morphological structures (Roulin & Wink, 2004). Mechanistically, genetic correlations can arise if a given gene regulates different traits (pleiotropy), if several genes encoding a given phenotype are physically linked (Johnston et al., 2010) or if these genes are in linkage disequilibrium due to, for example, nonrandom mating (Lynch & Walsh, 1998).

In the barn owl, we studied three melanin-based colour traits that necessarily share part of the melanogenic biochemical cascade. As could be predicted, the strongest genetic correlations were between the two eumelanin-based plumage traits (number and diameter of black spots), mean of male and female genetic correlations being 0.794, twice as strong as the genetic correlation between number of eumelanic spots and pheomelanin-based coloration (0.348). Interestingly, genetic correlations were on average 1.6 times stronger in males than in females (Table 3), possibly as a consequence of Z-linked genes (Table 2), implying that the evolution of a given plumage trait should be particularly constrained by the evolution of the two other traits in males. This finding suggests that the three melanin-based traits may have a more redundant function in males than in females. Accordingly, in females (and to a lower extent in males) the size of black spots is related to behaviour and physiology (Roulin & Ducrest, 2011), and pheomelanin-based coloration plays a role in the adaptation to local conditions (Dreiss et al., 2012), probably associated with predator-prey relationships (Charter et al., 2015).

Genetic correlations between the three plumage traits were all positive (Table 3), that is darker pheomelanic individuals displayed more and larger black spots. This suggests that the production of pheomelanin and eumelanin pigments is not traded off against each other. Therefore, the expression of the melanogenic genes that allow the production of the precursors of both melanin types may have an overwhelming effect on plumage traits compared to genes that trigger the expression of pheomelanin pigments at the expanse of eumelanin pigments. To test these scenarios, measurement of gene expression is needed (e.g. Emaresi *et al.*, 2013).

Genetic correlations between homologous traits in the

Each sex can evolve more rapidly towards its phenotypic optimum if the genetic correlation between the sexes is low rather than high (e.g. Chenoweth et al., 2007) as shown in a review of the literature (Poissant et al., 2009). This is particularly relevant in species in which a trait is the target of sexually antagonistic selection where males are positively selected and females negatively selected (or vice versa). When the genetic correlation between the sexes is high, positively selected males will not only produce sons having a selective advantage but also counter-selected daughters, and the other way round with positively selected females (Foerster et al., 2007; Mills et al., 2012). In such a situation, selection should favour the breakdown of the intersexual genetic correlation, a process that may, however, take many generations (Lande, 1987). As a consequence, males who inherit the counter-selected femalelike version of a sexually antagonistically selected trait (and females who inherit a counter-selected malelike trait) may evolve compensatory strategies to reduce the cost of sexually antagonistic selection (e.g. Abbott et al., 2013).

In the barn owl, we found evidence for the hypothesis that spot size is sexually antagonistically selected (positive selection in females and negative selection in males; Roulin et al., 2010, 2011). Although some of the underlying genes are located on sex chromosomes (Roulin et al., 2010; Larsen et al., 2014; Table 2), the genetic correlation between males and females for spot size is very strong (0.963; Table 3b), implying that small-spotted fathers will produce counter-selected daughters. Furthermore, if the absolute strength of negative selection exerted in males is weaker than positive selection in females, males will evolve away from their phenotypic optimum, as we could demonstrate in Switzerland (Roulin et al., 2010). This may explain why offspring sex ratio is correlated with parental spot diameter, the probability of producing sons being higher when both parents displayed a malelike plumage (i.e. small spots) and lowest when at least one of the parent displayed a femalelike plumage (i.e. large spots). Furthermore, malelike females and femalelike males produced sons and daughters with a high survival prospect, respectively (Roulin et al., 2010). These two compensatory mechanisms may have evolved as a consequence of the very strong genetic correlation between the sexes for spot size.

Interestingly, the degree of offspring sexual dimorphism was related to parental phenotype (Table 4, Fig. 2), implying that some parents produce daughters and sons that closely resemble each other, whereas other parents produce very distinct daughters and sons. For the three plumage traits, the degree of sexual dimorphism decreased with parental melanism, that is

when parents (particularly mother) were darker reddish or displayed more and larger black spots, sons resembled their daughters to a larger degree than when their parents were pale reddish or lightly spotted (Table 4). From a proximate of view, a potential explanation is that a gene of major phenotypic effect (such as MCIR) determines the amount of pigments produced in the two sexes and another gene of minor phenotypic effect (such as those of the melanocortin system) is responsible for the slight overexpression of melanin in females compared to males. In that case, when parents possess a mutation of the gene of major phenotypic effect that triggers the expression of a large amount of melanin pigments, feathers become saturated in melanin, implying that the gene of minor phenotypic effect will have hardly any effect on sexual dimorphism.

With respect to spot size, sons and daughters differed the most when their mother displayed small rather than large black spots. Because this trait is sexually antagonistically selected (Roulin et al., 2010), the resulting intralocus genetic conflict may be particularly strong when mothers are large spotted, as they will produce counter-selected large-spotted sons. As can be seen in Fig. 3c, large-spotted mothers produce sons and daughters who are similarly spotted, whereas smallspotted mothers produce offspring displaying spots at the size that is typical for their sex (i.e. large-spotted daughters and small-spotted sons). This finding is particularly interesting as it suggests that positive selection on female spot size is associated with more intense intralocus genetic conflict, whereas negative selection on male spot size is associated with a reduced genetic conflict. This further suggests that mutations that increase the expression of larger spots are associated with more intense genetic conflict, whereas mutations that suppress the expression of large spots are related to a reduced conflict. It would be particularly interesting to identify the genes involved in the expression of large black spots but also to determine whether they are ancestral or derived (i.e. if barn owls were originally large- or small-spotted).

Genetic correlations between nonhomologous traits within and across the sexes

Genetic correlations between nonhomologous traits have been shown to reduce the rate of adaptation (Teplitsky *et al.*, 2014a, b). In contrast, a study in the house sparrow (*Passer domesticus*) showed that indirect selection had a larger contribution to the predicted evolution of a melanin-based trait in males than direct selection (Jensen *et al.*, 2008). Of particular interest was the finding that selection exerted on female morphology can affect the evolution of male ornamentation. This reinforces the idea that a particular trait cannot be considered independently from other phenotypic characters. This statement is not trivial because the impor-

tance of natural selection on the evolution of sexually selected traits may be considerable, but the role played by indirect selection exerted on genetically correlated traits in males and females is rarely studied or even discussed (see reviews in Jensen *et al.*, 2008 and Teplitsky *et al.*, 2014a, b).

Acknowledgments

We thank A.M. Holand for help with running the INLA analyses. The Swiss National Science Foundation (grants no. PPOA-102913 and 31003A_120517), Foundation De Giacomi and Herbette, and the Research Council of Norway (grants no. 221956 and 223257 to HJ) supported this study financially. We are grateful to two anonymous reviewers for useful comments.

References

- Abbott, J.K., Innocenti, P., Chippindale, A.K. & Morrow, E.H. 2013. Epigenetics and sex-specific fitness: an experimental test using male-limited evolution in *Drosophila melanogaster*. *PLoS One* **8**: e70493.
- Andersson, M. 1994. Sexual selection. Princeton University Press, Princeton, NJ.
- Bonduriansky, R. & Chenoweth, S.F. 2009. Intralocus sexual conflict. *Trends Ecol. Evol.* **24**: 280–288.
- Brooks, R. & Endler, J.A. 2001. Direct and indirect sexual selection and quantitative genetics of male traits in guppies (*Poecilia reticulata*). *Evolution* **55**: 1002–1015.
- Charmantier, A., Garant, D. & Kruuk, L.E.B. 2014. *Quantitative Genetics in the Wild*. Oxford University Press, Oxford, UK.
- Charter, M., Peleg, O., Leshem, Y. & Roulin, A. 2012. Similar patterns of local barn owl adaptation in the Middle East and Europe with respect to melanic coloration. *Biol. J. Linn. Soc.* **106**: 447–454.
- Charter, M., Leshem, Y., Izhaki, I. & Roulin, A. 2015. Pheomelanin-based colouration is correlated with indices of flying strategies in the barn owl. *J. Ornithol.* **156**: 309–312.
- Chenoweth, S.F., Petfield, D., Doughty, P. & Blows, M.W. 2007. Male choice generates stabilizing sexual selection on a female fecundity correlate. *J. Evol. Biol.* **20**: 1745–1750.
- Chenoweth, S.F., Rundle, H.D. & Blows, M.W. 2008. Genetic constraints and the evolution of display trait sexual dimorphism by natural and sexual selection. *Am. Nat.* **171**: 22–34.
- Cuervo, J.J. & Møller, A.P. 2000. Sex-limited expression of ornamental feathers in birds. *Behav. Ecol.* 11: 246–259.
- Dean, R. & Mank, J.E. 2014. The role of sex chromosomes in sexual dimorphism: discordance between molecular and phenotypic data. *J. Evol. Biol.* 27: 1443–1453.
- Dreiss, A.N. & Roulin, A. 2010. Age-related change in melanin-based coloration: females that become more female-like and males more male-like with age perform better in barn owls (*Tyto alba*). *Biol. J. Linn. Soc.* 101: 689–704.
- Dreiss, A.N., Antoniazza, S., Burri, R., Fumagalli, L., Sonnay, C., Frey, C. *et al.* 2012. Local adaptation and matching habitat choice in female barn owls with respect to melanic coloration. *J. Evol. Biol.* **25**: 103–114.
- Emaresi, G., Ducrest, A.-L., Bize, P., Richter, H., Simon, C. & Roulin, A. 2013. Pleiotropy in the melanocortin system:

- expression levels of this system are associated with melanogenesis and pigmentation in the tawny owl (*Strix aluco*). *Mol. Ecol.* **22**: 4915–4930.
- Evans, S.R., Schielzeth, H., Forstmeier, W., Sheldon, B.C. & Husby, A. 2014. Nonautosomal genetic variation in carotenoid coloration. *Am. Nat.* **184**: 374–383.
- Fairbairn, D.J. & Roff, D.A. 2006. The quantitative genetics of sexual dimorphism: assessing the importance of sex-linkage. *Heredity* **97**: 319–328.
- Foerster, K., Coulson, T., Sheldon, B.C., Pemberton, J.M., Clutton-Brock, T.H. & Kruuk, L.E.B. 2007. Sexually antagonistic genetic variation for fitness in red deer. *Nature* **447**: 1107–1110.
- Gosden, T.P., Shastri, K.-L., Innocenti, P. & Chenoweth, S.F. 2012. The B-matrix harbours significant and sex-specific constraints on the evolution of multi-character sexual dimorphism. *Evolution* **66**: 2106–2116.
- Groeneveld, E., Kovac, M. & Mielenz, N. 2010. VCE User's Guide and Reference Manual Version 6.0. ftp://ftp.tzv.fal.de/pub/vce6/doc/vce6-manual-3.1-A4.pdf.
- Hadfield, J.D., Nutall, A., Osorio, D. & Owens, I.P.F. 2007.
 Testing the phenotypic gambit: phenotypic, genetic and environmental correlations of colour. *J. Evol. Biol.* 20: 549–557
- Harano, T., Okada, K., Nakayama, S., Miyatake, T. & Hosken, D.J. 2010. Intralocus sexual conflict unresolved by sex-limited trait expression. *Curr. Biol.* 20: 2036–2039.
- Henry, I., Antoniazza, S., Dubey, S., Simon, C., Waldvogel, C., Burri, R. et al. 2013. Multiple paternity in polyandrous barn owls (Tyto alba). PLoS One 8: e80112.
- Holand, A.M., Steinsland, I., Martino, S. & Jensen, H. 2013. Animal models and integrated nested Laplace approximations. *G3* 3: 1241–1251.
- Husby, A., Schielzeth, H., Forstmeier, W., Gustafsson, L. & Qvarnström, A. 2012. Sex chromosome linked genetic variance and the evolution of sexual dimorphism of quantitative traits. *Evolution* 67: 609–619.
- Jensen, H., Sæther, B.-E., Ringsby, T.H., Tufto, J., Griffith, S.C. & Ellegren, H. 2003. Sexual variation in heritability and genetic correlations of morphological traits in house sparrow (*Passer domesticus*). J. Evol. Biol. 16: 1296–1307.
- Jensen, H., Steinsland, I., Ringsby, T.H. & Sæther, B.-E. 2008. Evolutionary dynamics of a sexual ornament in the house sparrow (*Passer domesticus*): the role of indirect selection within and between sexes. *Evolution* **62**: 1275–1293.
- Johnston, S.E., Beraldi, D., McRae, A.F., Pemberton, J.M. & Slate, J. 2010. Horn type and horn length genes map to the same chromosomal region in Soay sheep. *Heredity* **104**: 196–205
- Kim, S.-Y., Fargallo, J.A., Vergara, P. & Martínez-Padilla, J. 2013. Multivariate heredity of melanin-based coloration, body mass and immunity. *Heredity* 111: 139–146.
- Kirkpatrick, M. 2009. Patterns of quantitative genetic variation in multiple dimensions. *Genetica* **136**: 271–284.
- Kruger, O. 2005. The evolution of reversed sexual size dimorphism in hawks, falcons and owls: a comparative study. *Evol. Ecol.* 19: 467–486.
- Kruuk, L.E.B. & Hill, W.G. 2008. Introduction. Evolutionary dynamics of wild populations: the use of long-term pedigree data. Proc. R. Soc. Lond. B Biol. Sci. 275: 593–596.
- Lande, R. 1980. Sexual dimorphism, sexual selection, and adaptation in polygenic characters. *Evolution* **34**: 292–305.

- Lande, R. 1987. Genetic correlations between the sexes in the evolution of sexual dimorphism and mating preferences. In: *Sexual Selection: Testing the Alternatives* (J.W. Bradbury & M. Andersson, eds), pp. 83–94. Dahlem Konferensen, Wiley Press, Chichester, UK.
- Larsen, C.T., Holand, A.M., Jensen, H., Steinsland, I. & Roulin, A. 2014. On estimation and identifiability issues of sex-linked inheritance with a case study of pigmentation in Swiss barn owls (*Tyto alba*). *Ecol. Evol.* **4**: 1555–1566.
- Lindenfors, P. 2002. Sexually antagonistic selection on primate size. J. Evol. Biol. 15: 595–607.
- Lynch, L. & Walsh, B. 1998. *Genetics and Analysis of Quantitative Traits*. Sinauer Associates Inc, Sunderland, MA.
- Mank, J.E. 2009. Sex chromosomes and the evolution of sexual dimorphism: lessons from the genome. *Am. Nat.* **173**: 141–150.
- Mank, J.E. & Ellegren, H. 2009. Sex-linkage of sexually antagonistic gene is predicted by female, but not male, effects in birds. *Evolution* **63**: 1464–1472.
- McGlothlin, J.W., Parker, P.G., Nolan, V. & Ketterson, E.D. 2005. Correlation selection leads to genetic integration of body size and an attractive plumage trait in dark-eyed juncos. *Evolution* **59**: 658–671.
- Mills, S.C., Koskela, E. & Mappes, T. 2012. Intralocus sexual conflict for fitness: sexually antagonistic alleles for testosterone. Proc. R. Soc. Lond. B Biol. Sci. 279: 1889–1895.
- Munro, R.E., Smith, L.T. & Kupa, J.J. 1968. The genetic basis of color differences observed in the mute swan (*Cygnus olor*). *Auk* **85**: 504–505.
- Naurin, S., Hansson, B., Bensch, S. & Hasselquist, D. 2009. Why does dosage compensation differ between XY and ZW taxa. Trends Genet. 26: 15–20.
- Neumaier, A. & Groeneveld, E. 1998. Restricted maximum likelihood estimation of covariances in sparse linear models. *Genet. Sel. Evol.* 1: 3–26.
- Poissant, J., Wilson, A.J., Festa-Bianchet, M., Hogg, J.T. & Coltman, D.W. 2008. Quantitative genetics and sex-specific selection on sexually dimorphic traits in bighorn sheep. *Proc. R. Soc. Lond. B Biol. Sci.* 275: 623–628.
- Poissant, J., Wilson, A.J. & Coltman, D.W. 2009. Sex-specific genetic variance and the evolution of sexual dimorphism: a systematic review of cross-sex genetic correlations. *Am. Nat.* 173: 176–187.
- Potti, J. & Canal, D. 2011. Heritability and genetic correlation between the sexes in a songbird sexual ornament. *Heredity* **106**: 945–954.
- Preziosi, R.F. & Fairbairn, D.J. 2000. Lifetime selection in adult body size and components of body size in a waterstrider: opposing selection and maintenance of sexual size dimorphism. *Evolution* **54**: 558–566.
- Reinhold, K. 1998. Sex-linkage among genes controlling sexually selected traits. *Behav. Ecol. Sociobiol.* **44**: 1–7.
- Rhen, T. 2000. Sex-limited mutations and the evolution of sexual dimorphism. *Evolution* **54**: 37–43.
- Rice, W.R. 1984. Sex-chromosomes and the evolution of sexual dimorphism. *Evolution* **38**: 735–742.
- Roff, D.A. 1997. Evolutionary Quantitative Genetics. Chapman and Hall, New York.
- Roff, D.A. & Fairbairn, D.J. 2012. A test of the hypothesis that correlational selection generates genetic correlations. *Evolution* 66: 2953–2960.

- Roulin, A. 1999. Nonrandom pairing by male barn owls (*Tyto alba*) with respect to a female plumage trait. *Behav. Ecol.* **10**: 688–695.
- Roulin, A. 2004a. Covariation between plumage colour polymorphism and diet in the barn owl *Tyto alba*. *The Ibis* **146**: 509–517.
- Roulin, A. 2004b. Proximate basis of the covariation between a melanin-based female ornament and offspring quality. *Oecologia* **140**: 668–675.
- Roulin, A. & Dijkstra, C. 2003. Genetic and environmental components of variation in eumelanin and phaeomelanin sex-traits in the barn owl. *Heredity* **90**: 359–364.
- Roulin, A. & Ducrest, A.-L. 2011. Association between melanism, physiology and behaviour: a role for the melanocortin system. *Eur. J. Pharmacol.* **660**: 226–233.
- Roulin, A. & Wink, M. 2004. Predator-prey relationships and the evolution of genetic colour polymorphism: a comparative analysis in diurnal raptors. *Biol. J. Linn. Soc.* **81**: 565–578.
- Roulin, A., Ducrest, A.-L. & Dijkstra, C. 1999. Effect of brood size manipulations on parents and offspring in the barn owl *Tyto alba. Ardea* **87**: 91–100.
- Roulin, A., Riols, C., Dijkstra, C. & Ducrest, A.-L. 2001. Female- and male-specific signals of quality in the barn owl. *J. Evol. Biol.* **14**: 255–267.
- Roulin, A., Wink, M. & Salamin, N. 2009. Selection on a eumelanic ornament is stronger in the tropics than in temperate zones in the worldwide-distributed barn owl. *J. Evol. Biol.* **22**: 345–354.
- Roulin, A., Altwegg, R., Jensen, H., Steinsland, I. & Schaub, M. 2010. Sex-dependent selection on an autosomal melanic female ornament promotes the evolution of sex ratio bias. *Ecol. Lett.* **13**: 616–626.
- Roulin, A., Antoniazza, S. & Burri, R. 2011. Spatial variation in the temporal change of male and female melanic ornamentation in the barn owl. *J. Evol. Biol.* **24**: 1403–1409.
- Sinervo, B. & Svensson, E. 2002. Correlation selection and the evolution of genomic architecture. *Heredity* **89**: 329–338.
- Southern, H.N. 1946. Polymorphism in *Poephila gouldiae* Gould. J. Genet. 47: 51–57.
- Steinsland, I. & Jensen, H. 2010. Utilizing Gaussian Markov random field properties of Bayesian animal models. *Biometrics* **66**: 763–771.
- Taylor, I.R. 1993. Age and sex determination of barn owls Tyto alba alba. Ring. Migr. 14: 94–102.
- Teplitsky, C., Tarka, M., Møller, A.P., Nakagawa, S., Balbontin, J., Burke, T.A. *et al.* 2014a. Assessing multivariate constraints to evolution across ten long-term avian studies. *PLoS One* 9: e90444.
- Teplitsky, C., Robinson, M.R. & Merilä, J. 2014b. Evolutionary potential and constraints in wild populations. In: *Quantitative Genetics in the Wild* (A. Charmantier, D. Garant & L.E.B. Kruuk, eds), pp. 190–208. Oxford University Press, Oxford, UK.
- Van den Brink, V., Dolivo, V., Falourd, X., Dreiss, A. & Roulin, A. 2012. Melanic color-dependent anti-predator behavior strategies in barn owl nestlings. *Behav. Ecol.* 23: 473–480.
- Zann, R.A. 1996. The Zebra Finch, a Synthesis of Field and Laboratory Studies. Oxford University Press, Oxford, UK.

Received 12 December 2014; revised 23 January 2015; accepted 26 January 2015