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Endothelial nitric oxide synthase gene transfer restores endothelium-dependent relaxations and attenuates lesion formation in carotid arteries in apolipoprotein E-deficient mice

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Introduction

Endothelial nitric oxide (NO) synthase (eNOS) constitutively produces NO, a potent vasodilator that mediates pleiotropic vasoprotective effects such as inhibition of endothelial leukocyte adhesion, suppression of platelet aggregation, and negative regulation of vascular smooth muscle cell (VSMC) proliferation [10, 31, 51]. Endothe-

■ **Abstract** Nitric oxide (NO) and monocyte chemoattractant protein-1 (MCP-1) exert partly opposing effects in vascular biology. NO plays pleiotropic vasoprotective roles including vasodilation and inhibition of platelet aggregation, smooth muscle cell proliferation, and endothelial monocyte adhesion, the last effect being mediated by MCP-1 downregulation. Early stages of arteriosclerosis are associated with reduced NO bioactivity and enhanced MCP-1 expression. We have evaluated adenovirus-mediated gene transfer of human endothelial NO synthase (eNOS) and of a *N*-terminal deletion (8ND) mutant of the MCP-1 gene that acts as a MCP-1 inhibitor in arteriosclerosis-prone, apolipoprotein E-deficient (ApoE^{-/-}) mice. Endothelium-dependent relaxations were impaired in carotid arteries instilled with a noncoding adenoviral vector but were restored by eNOS gene transfer ($p < 0.01$). A perivascular collar was placed around the common carotid artery to accelerate lesion formation. eNOS gene transfer reduced lesion surface areas, intima/media ratios, and macrophage contents in the media at 5-week follow-up ($p < 0.05$). In contrast, 8ND-MCP-1 gene transfer did not prevent lesion formation. In conclusion, eNOS gene transfer restores endothelium-dependent vasodilation and inhibits lesion formation in ApoE^{-/-} mouse carotids. Further studies are needed to assess whether vasoprotection is maintained at later disease stages and to evaluate the long-term efficacy of eNOS gene therapy for primary arteriosclerosis.

■ **Key words** NO synthase – MCP-1 – arteriosclerosis – endothelium – apolipoprotein-E – mouse – gene therapy

lium-dependent vasodilation is impaired in hypercholesterolemic animals and humans, suggesting that NO bioactivity is decreased at early stages of vascular disease [12, 43]. Administration of the NO precursor L-arginine has been shown to correct endothelial dysfunction in hypercholesterolemic patients [11]. Chronic dietary administration of L-arginine induces regression of early intimal lesions and attenuates progression of atherosclerotic plaques in hypercholesterolemic mice and rabbits

[1, 3]. Conversely, chronic administration of NO antagonists promotes lesion formation in hypercholesterolemic rabbits [7]. There is experimental and clinical evidence to show that eNOS protein levels are decreased in the endothelium overlying severe atherosclerotic lesions in arteries [13, 27, 47] and diseased bypass grafts [5]. However, reduced NO bioactivity has also been described in association with normal or even increased eNOS protein levels [2, 12]. These findings illustrate the complexity of the mechanisms underlying the observed decrease in NO bioactivity in diseased arteries.

Mice deficient in the apolipoprotein E gene (ApoE^{-/-}) provide a useful model for studying atherosclerosis in that complex arterial lesions in this model mimic those observed in humans [33]. Lesion formation is further accelerated in ApoE/eNOS double-knockout mice compared with ApoE^{-/-} mice [20]. Conversely, reduced lesion formation in a model of vascular remodeling [19], as well as hypotension and reduced NO-elicited vasorelaxation [28] have been reported in ApoE^{-/-}/eNOS transgenic mice that overexpress eNOS in the vascular endothelium, as compared with ApoE^{-/-} mice. In contrast, a recent study [30] showed accelerated lesion formation in ApoE^{-/-}/eNOS transgenic mice, which was reversed by supplementation with tetrahydrobiopterin, an important NO cofactor.

Previous studies have demonstrated that eNOS gene transfer improves vasodilation in endothelial dysfunction, while reducing neointimal hyperplasia in mechanically injured arteries [9]. *In vitro* eNOS gene transfer improved vasorelaxation in canine coronary arteries during hypoxia [6] and carotid arteries from diabetic rabbits [23] and eNOS^{-/-} mice [21]. *In vivo* eNOS gene transfer attenuated acute hypoxic pulmonary vasoconstriction in rats [16] and improved carotid artery dilatation in hypercholesterolemic rabbits [8, 35]. Furthermore, eNOS gene transfer inhibited hyperplastic neointima formation in balloon-injured rat carotid arteries [9, 17], pig coronary arteries [40], and atherosclerotic ApoE^{-/-} mouse carotid arteries [45]. However, eNOS gene transfer has not been investigated in models of primary arteriosclerosis that do not rely on mechanical endovascular injury to promote lesion formation.

NO and monocyte chemoattractant protein-1 (MCP-1) play partly opposing roles in vascular biology. MCP-1 is expressed in endothelial cells, VSMC, monocytes, and adipocytes in response to inflammatory stimuli such as TNF- α , IL-1 β , and IL-4 [36]. MCP-1 expression is detectable in aortas from hypercholesterolemic, but not from normal, rabbits [48]. Downregulation of MCP-1 expression is a mechanism by which NO inhibits monocyte adhesion to the endothelium [26, 37, 38, 49]. MCP-1 expressed by human aortic endothelial cells and VSMC in response to minimally modified low-density lipoprotein (LDL) cholesterol accounts for virtually all of the monocyte chemotactic activity under these conditions

[32]. Mice deficient in MCP-1 show abnormal monocyte recruitment and cytokine expression [22]. Mice deficient in the C-C chemokine receptor-2 (CCR2), the main receptor for MCP-1, display reduced arterial lesion formation with less macrophage infiltrates [4]. These findings establish a central role for MCP-1 in atherogenesis, while providing a rationale for anti-MCP-1 therapy. Among MCP-1 inhibitors, N-terminally truncated MCP-1 peptides have been shown to block MCP-1-mediated monocyte chemotaxis with high efficacy both *in vitro* and *in vivo* [14, 50]. Gene transfer of the 7 N-terminal deletion (ND) mutant of the human MCP-1 gene inhibited post-angioplasty neointima formation in rat and monkey arteries [39]. Injection of the same gene construct into remote muscles attenuated aortic lesion formation and progression in ApoE^{-/-} mice [15, 25]. However, arterial gene transfer of a MCP-1 inhibitor has not been studied in the absence of angioplasty injury. Therefore, we have evaluated arterial eNOS and anti-MCP-1 gene transfer-based approaches in carotid arteries in ApoE^{-/-} mice.

Methods

■ Adenoviral vectors

The expression plasmid encoding human eNOS (pUC18-eNOS) was kindly provided by Daniel Hayoz (Lausanne, Switzerland). Generation of recombinant adenovirus expressing Xpress-tagged human eNOS driven by the cytomegalovirus (CMV) promoter was carried out by homologous recombination between co-transformed virusmid and the corresponding adenoviral transfer plasmid in *E. coli*, as described [24]. The eNOS cDNA was first subcloned into the pcDNA3.1/His-A vector (Invitrogen) allowing the insertion of the transgene into the E1 region of a cloned $\Delta E1, \Delta E3$ adenoviral backbone (vmAdcDNA3) bearing an empty expression cassette homologous to the one of pcDNA3.1/His. The transfer plasmid pcDNA3.1/His-eNOS was digested with *SalI*, and co-transformed with *SwaI*-linearized vmAdcDNA3 into BJ5183. Recombinants were screened by colony PCR followed by re-transformation of positive clones into strain HB101, allowing for higher plasmid yields. Recombinant virusmids were digested with *PacI* and transfected into HER911 cells to generate recombinant adenovirus AdCMV-eNOS. The cDNA sequence of human MCP-1 was kindly provided by Etienne Régulier (Lausanne, Switzerland). The cDNA sequence corresponding to the 8ND mutant of the MCP-1 gene encoding a truncated MCP-1 peptide lacking amino acid residues 1-8 was obtained by PCR using site-directed deletion by overlap extension. 8ND-MCP-1 cDNA was sequenced and sub-

cloned into the pcDNA3.1/A vector. Recombinant adenovirus Ad.CMV-8ND-MCP-1 was generated by homologous recombination with the co-transfected, *Cla*I-digested large fragment of a $\Delta E1, \Delta E3$ -adenoviral vector genome in human 293 cells. Recombinant adenovirus Ad-BHG3 containing no transgene (Ad-Null) was used as a control. Recombinant adenovirus Ad.CMV-EGFP containing an enhanced green fluorescent protein (EGFP) expression cassette [42] was used to assess arterial gene transfer efficiency. Concentrated virus stocks were prepared using two CsCl_2 ultracentrifugation gradients and stored in storage buffer (10 mmol/L Tris-HCl, pH 7.4, 1 mmol/L MgCl_2 , 10% glycerol). Virus titers of Ad.CMV-eNOS, Ad.CMV-8ND-MCP-1, Ad-Null, and Ad.CMV-EGFP stock solutions determined by plaque titration assay were 2×10^{11} , 6×10^{10} , 1×10^{11} , and 3×10^{11} plaque forming units (PFU)/ml, respectively (virus particles/PFU ratios ≈ 5 -30).

■ Mice

Animal work was approved by the local regulatory authority and was performed in compliance with guidelines on animal experimentation at our institution. Male C57BL/6J ApoE^{-/-} mice [33] were purchased from Transgenic Alliance (France) and maintained on a 12-hour light-dark cycle. They were fed standard chow until 18-20 weeks of age when they underwent gene transfer and were placed on a Western-type diet (Harlan TD88137, 21.2% total fat, 0.2% cholesterol) for 2 weeks, followed by standard chow for 3 weeks.

■ *In vivo* gene transfer

Gene transfer into mouse carotid arteries was performed essentially as described [42]. Briefly, mice ($n = 7/\text{group}$) were anesthetized by i.p. ketamine/xylazine. The left carotid artery was surgically exposed, and a segment of the common carotid artery was isolated by placement of a vascular clamp on the proximal part of the artery, while the internal carotid artery was ligated with a polypropylene monofilament suture (7-0 Surgilene; Geck and Davis) immediately after the bifurcation. The external carotid artery was ligated 2-3 mm distally of the bifurcation. A silicone catheter (.012" I.D., .025 O.D.; Helix Medical) was introduced into the common carotid artery through an arteriotomy of the external carotid artery, and blood was flushed out the common carotid artery by injecting 0.3-0.4 ml M-199 medium (Gibco BRL). Virus vector stocks were thawed and diluted with M-199 medium supplemented with 1 mg/ml purified mouse albumin fraction V (Sigma) to the final concentration of 2×10^{10} PFU/ml. This concentration was chosen based on our previous dose-response study [42]. A second catheter

was reintroduced, and vector containing medium was instilled into the isolated common carotid segment. The duration of the virus incubation period was 20 minutes. Vector containing medium was then withdrawn and blood flow was re-established through the common and internal carotid arteries. To accelerate lesion formation, a perivascular collar prepared from Silastic tubing (inside diameter ≈ 0.3 mm; length ≈ 3 mm) was placed around the central portion of the left common carotid artery, as described by von der Thüsen et al. [44]. The collar results in a modest reduction in vessel lumen ($\approx 30\%$) that does not limit blood flow but causes flow disturbances that accelerate lesion formation within 3-6 weeks in ApoE^{-/-} mice fed a Western-type diet [44, 45]. The axial edges of the collar were approximated by placement of 3 circumferential silk ties. Subsequently, the cervical wound was closed.

■ Detection of human eNOS and 8ND-MCP-1

Human eNOS protein in ApoE^{-/-} carotid arteries instilled *in vivo* with Ad.CMV-eNOS or Ad-Null ($n = 3$ each) was detected by Western blotting. Briefly, crude protein extracts were obtained by homogenizing snap-frozen arteries in protein extraction buffer, as described [24]. Protein concentrations were measured by the Bradford method. Protein samples (30 μg) were subjected to SDS-PAGE and transferred to a PVDF membrane (Millipore). Mouse anti-eNOS mAb (Transduction Lab.) and anti-mouse alkaline phosphatase-conjugated secondary Ab were used and visualized with Western Blue Stabilized Substrate (Uptima). Signals were quantified densitometrically by the NIH Image-1.62 software. NOS levels were normalized against total actin levels. Human 8ND-MCP-1 protein was measured by a commercially available human MCP-1 ELISA kit (Biosource) that does not react with endogenous mouse MCP-1 [15].

■ Vascular reactivity studies

Vessel rings (≈ 3 mm in length) with intact endothelium were cut from carotid arteries isolated from ApoE^{-/-} mice (18-20 weeks of age) fed a Western-type diet at day 5 after *in vivo* gene transfer with Ad.CMV-eNOS or Ad-Null ($n = 4$ each). They were suspended in a modified Krebs-Ringer bicarbonate solution (in mmol/L, NaCl 118, KCl 4.7, CaCl_2 2.5, MgSO_4 1.2, KH_2PO_4 1.2, NaHCO_3 25, edetate calcium disodium 0.026, and glucose 11.1) in a Multi-Myograph System (Model 610M, Danish Myo Technology A/S, Denmark). The carotid rings were contracted with norepinephrine (0.3 $\mu\text{mol/L}$). Endothelium-dependent and -independent relaxations were examined with acetylcholine and sodium nitroprusside, respectively.

■ Tissue harvesting and preparation

For histological analysis of arterial lesions, mice were sacrificed by a lethal pentobarbital dose at 5 weeks after gene transfer. *In situ* perfusion-fixation through the left cardiac ventricle was performed by PBS instillation for 5 minutes, followed by constant-pressure (80 mmHg) infusion of 4% neutral buffered formalin for 10 minutes. Subsequently, the perivascular collar was removed and the carotid artery was harvested and kept in 4% formalin for 24 hours. The specimens were embedded in OCT compound (Tissue-Tek; Sakura Finetek) and snap-frozen in liquid nitrogen. Five series of carotid cross-sections (n = 10 each) at 7 μ m of thickness were cut at ~0.3 mm step intervals throughout the vessel segment subjected to gene transfer.

■ Histological and immunohistochemical analysis

Two series of arterial cross-sections were stained with hematoxylin and eosin (H&E) and elastic/van Gieson for optimal visualization of the internal elastic lamina. Immunostaining for macrophages was performed on additional sections using anti-mouse Mac-2 mAb (Cedarlane) and a biotin-conjugated rabbit-anti-rat IgG secondary Ab, followed by StreptABComplex/HRP and DAB/H₂O₂ (Dako). Sections were counter-stained with eosin. Images were acquired using a Hyper-HAD-Axiocscope microscope and an Axiocam-MRcD camera (Zeiss). Morphometric analysis of intima and media surface areas was performed by the NIH Image-1.62 software. The intima surface area was calculated by subtracting the patent lumen area from the area encompassed by the internal elastic lamina. The media surface area was measured as the area between the external and the internal

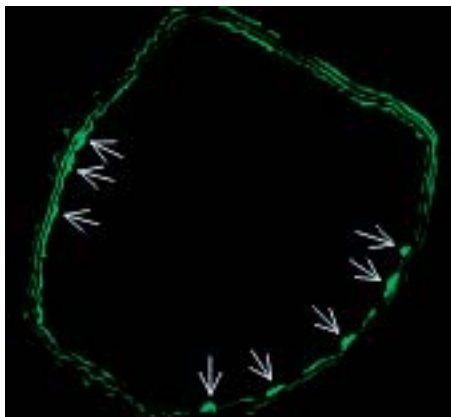


Fig. 1 Fluorescence photomicrograph of a mouse carotid artery 3 days after *in vivo* gene transfer with Ad.CMV-EGFP. Fluorescent, EGFP expressing endothelial cells lining the vessel lumen are shown (arrows). Three layers of elastic fibers in the media exhibit auto-fluorescence (200x original magnification)

elastic lamina. The intima/media ratio was calculated by dividing the intima surface area by the media surface area. Macrophages in arterial lesions and in the media were counted on Mac-2-immunostained sections, and normalized by the respective surface areas.

■ Measurement of plasma lipid levels

Serum total cholesterol and HDL-cholesterol (HDL-C) were measured using commercially available kits (Cholesterol RTU and HDL-Cholesterol Direct, BioMerieux).

■ Statistical analysis

Statistical analysis of arterial relaxations was performed by one-way ANOVA with Bonferroni post-test. Arterial intima and media surface areas, intima/media ratios, and macrophage contents are median values (range); they were compared between either the eNOS or the 8ND-MCP-1 gene transfer group and the Ad-Null group by the Mann-Whitney U test. Plasma lipid levels are mean values + SEM; they were compared between groups by ANOVA. A level of $p < 0.05$ was considered to be significant.

Results

■ Gene transfer efficiency

The efficiency of arterial gene transfer was assessed using an EGFP expressing vector. Arteries (n=3) were harvested at 3 days after gene transfer, and EGFP expressing cells were visualized on arterial sections under a fluorescence microscope. Approximately 10% of the endothelial cells lining the vessel lumen, but virtually no VSMC in the media, expressed EGFP at a histologically detectable level (Fig. 1). No fluorescent cells were observed in arteries instilled with Ad-Null.

■ eNOS and 8ND-MCP-1 expression

Western blot analysis for eNOS protein was performed on tissue extracts from mouse carotids harvested at 5 days after *in vivo* gene transfer with Ad.CMV-eNOS or Ad-Null. The eNOS signal in eNOS gene-transduced arteries was increased 2.3-fold compared with endogenous eNOS expression in arteries instilled with Ad-Null (Fig. 2). A human MCP-1 ELISA that recognizes human 8ND-MCP-1 but not mouse MCP-1 (15) detected 360 pg/gram of tissue after 8ND-MCP-1 gene transfer (Ad-Null, <30 pg/gram of tissue).

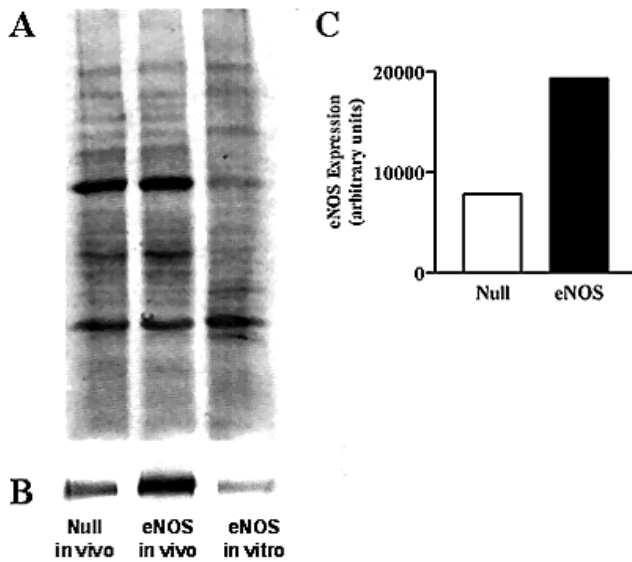


Fig. 2 Western blot analysis of eNOS gene expression in mouse carotids. **A** Ponceau staining showing equal amounts of loaded protein in each lane. **B** eNOS signal; left lane: *in vivo* Ad-Null administration (endogenous eNOS expression); middle lane: *in vivo* Ad.CMV-eNOS administration; right lane: *in vitro* positive control consisting of endothelial cells (HUVECs) infected with Ad.CMV-eNOS (not for quantitative comparison). **C** Quantitative analysis of eNOS protein levels in ApoE^{-/-} carotids *in vivo*; Ad-Null (white column), Ad.CMV-eNOS (black column; n = 3 each)

■ eNOS gene transfer improves endothelium-dependent relaxations

In carotid rings transduced with Ad-eNOS *in vivo*, endothelium-dependent relaxations to acetylcholine were markedly enhanced as compared to those instilled with Ad-Null (Fig. 3, $p < 0.01$), while the responses to sodium nitroprusside did not show significant differences between the two groups. These data demonstrate that

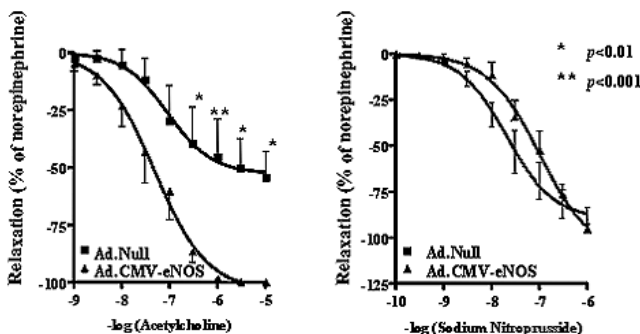


Fig. 3 Endothelium-dependent and -independent relaxation responses to acetylcholine (left) and sodium nitroprusside (right), respectively, in ApoE^{-/-} carotids infused with Ad.CMV-eNOS (triangles) or Ad-Null (square symbols). Endothelium-dependent relaxation is reduced in control arteries but is improved in eNOS gene-transduced arteries

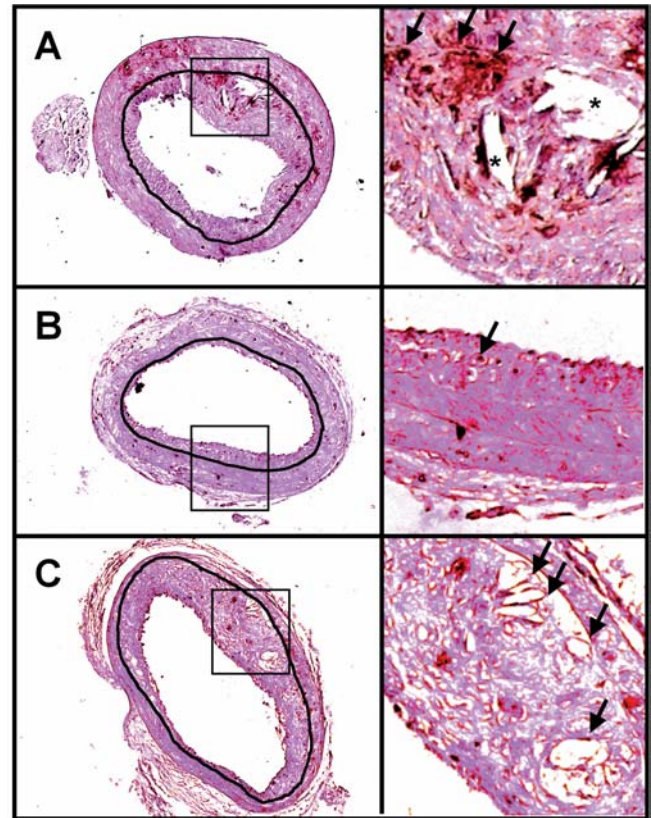


Fig. 4 Representative Mac-2 immunostained cross-sections of ApoE^{-/-} carotids 5 weeks after gene transfer. Right panels are high-magnification fields from left panels. Internal elastic laminae encompassing the lesions are drawn in black for better visualisation; counter-staining with eosin. **A** Ad-Null; large, macrophage-rich (brown; arrows) lesion containing cholesterol crystal clefts (*). **B** Ad.CMV-eNOS; smaller lesion with less abundant macrophages (arrow). **C** Ad.CMV-8ND-MCP-1; large lesion containing abundant foam cells (arrows)

eNOS gene transfer improves endothelium-dependent relaxations in ApoE^{-/-} mouse carotid arteries.

■ Morphometric analysis of gene-transduced arteries

Five weeks after gene transfer, arterial lesions had appeared in all experimental groups (Fig. 4). Lesions ranged from small intimal lesions, which were more frequent in the Ad.CMV-eNOS group (n = 6; one additional mouse died before completion of follow-up; Fig. 4B), to atheromatous plaques rich in foam cells, which were more frequent in the Ad-Null and Ad.CMV-8ND-MCP-1 groups (n = 7 each; Fig. 4 A and C, respectively). Cholesterol crystal clefts were seen occasionally in the last two groups. In the Ad.CMV-eNOS, Ad.CMV-8ND-MCP-1, and Ad-Null groups, intima surface area was 0.031 (0.002–0.068) mm² ($p < 0.05$), 0.053 (0.015–0.152) mm² (NS), and 0.073 (0.013–0.126) mm², respectively; media surface area was 0.058 (0.053–0.075) mm², 0.053

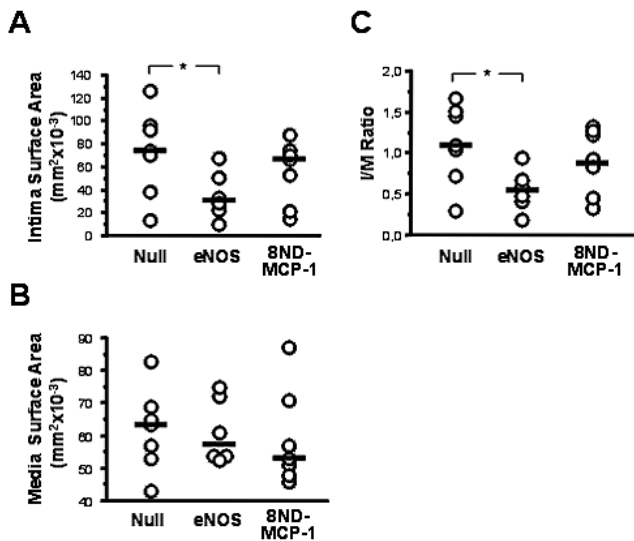


Fig. 5 Quantitative analysis of lesion size in ApoE^{-/-} mouse carotids. **A** Intima surface area; **B** Media surface area. **C** Intima/media ratio. Null, eNOS, and 8ND-MCP-1 refer to arteries instilled with the respective adenoviral vectors. Circles and horizontal bars are individual and median values, respectively. *p*-values for differences between each treatment group and the Ad-Null control group are shown

(0.044–0.089) mm² (both NS), and 0.065 (0.043–0.095) mm²; the intima/media ratio was 0.59 (0.17–0.95; *p* < 0.05), 1.07 (0.30–1.80; NS) and 1.22 (0.30–1.87), respectively (Fig. 5).

■ Macrophage content in eNOS gene-transduced arteries

Macrophage contents in the intima and media normalized by the respective surface areas are shown in Fig. 6. In the Ad.CMV-eNOS and Ad-Null groups, macrophages in intimal lesions were 54 (2–101) and 146 (26–165; NS), respectively. Macrophages in the media were less abundant than in the intima and decreased in the Ad.CMV-

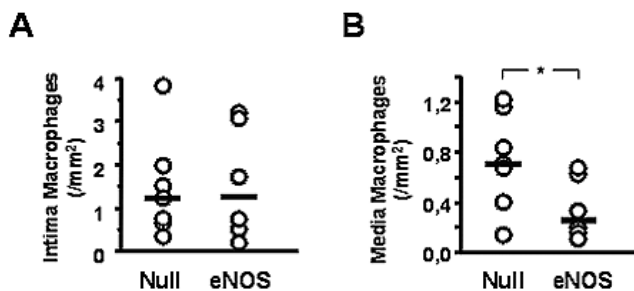


Fig. 6 Quantitative analysis of macrophage densities in ApoE^{-/-} mouse carotids instilled with Ad.CMV-eNOS or Ad-Null. **A** Intimal lesions. **B** Tunica media. Macrophages are normalized by the respective surface areas. Circles and horizontal bars are individual and median values, respectively. *p*-values for differences between each treatment group and the Ad-Null control group are shown

eNOS compared with the Ad-Null group: 19 (6–36) vs. 45 (28–102; *p* < 0.01), respectively.

■ Plasma lipid levels

Differences in plasma total cholesterol and HDL-C among groups were not statistically significant. One week after gene transfer and onset of the Western-type diet, total cholesterol in the Ad.CMV-eNOS, Ad.CMV-8ND-MCP-1, and Ad-Null groups was 463 ± 28, 418 ± 34, and 424 ± 84 mg/dl, while HDL-C was 64 ± 9, 54 ± 11, and 66 ± 11 mg/dl, respectively. Four weeks after gene transfer, when rats were fed standard chow, total cholesterol in the same groups was 248 ± 31, 323 ± 26 and 316 ± 47 mg/dl, while HDL-C was 40 ± 8, 37 ± 4, and 51 ± 3 mg/dl, respectively.

Discussion

Many gene therapy approaches to arteriosclerosis have focused on the systemic delivery of genes that encode proteins involved in lipid metabolism. Alternatively, genes that encode vasoprotective factors have been delivered directly to the vessel wall. Systemic and localized gene transfer approaches are associated with distinct advantages and disadvantages. In analogy to conventional lipid-lowering drugs, systemic gene transfer of a protein involved in lipid metabolism may affect atherosclerotic lesions spread throughout the arterial system. However, systemic gene overexpression may also be associated with systemic side effects, as exemplified by hypotension as a result of systemic eNOS overexpression [28]. Localized gene transfer into targeted vessels aims to protect vessel segments where arteriosclerosis is most likely to develop, or where its complications are most likely to occur. Using this approach, we have evaluated adenovirus-mediated *in vivo* eNOS and anti-MCP-1 gene therapy in carotid arteries in arteriosclerosis-prone ApoE^{-/-} mice. Experimental and clinical evidence indicates that the balance between eNOS and MCP-1 bioactivities plays an important role in atherogenesis. Endothelial NO activity is decreased at early stages of atherosclerosis, frequently associated with decreased levels of eNOS expression [13, 27, 47]. However, decreased NO bioactivity has been observed also in diseased arteries that show normal or even increased eNOS protein levels [2, 12]. These findings illustrate the complexity of the relationships between eNOS expression, NO bioactivity, and atherogenesis.

The main findings of the present study are that eNOS gene transfer improves endothelium-dependent relaxations and attenuates lesion formation in carotid arteries in ApoE^{-/-} mice. Studies from other groups showed

impaired endothelium-dependent vasodilation in carotid arteries in ApoE^{-/-} mice fed a Western-type diet [12]. In the present study, we showed that *in vivo* eNOS gene transfer into ApoE^{-/-} mouse carotid arteries resulted in increased eNOS expression levels and normalized relaxations to acetylcholine, suggesting exogenous eNOS was functional. To the best of our knowledge, this is the first report showing improved vasodilation by *in vivo* eNOS gene transfer into ApoE^{-/-} mouse arteries. These results are in general agreement with previous data showing improved endothelium-dependent relaxations by eNOS gene transfer into pulmonary arteries in hypoxic rats [16] and into carotid arteries in hypercholesterolemic rabbits [8, 35].

Carotid arteries transduced with the eNOS gene exhibited smaller lesions and lower macrophage contents compared with control arteries. These findings are consistent with previous data showing reduced vascular adhesion molecule expression and decreased inflammatory cell infiltration in eNOS gene-transduced arteries in hypercholesterolemic rabbits [8, 35]. In addition, several reports showed that eNOS gene transfer can reduce post-angioplasty neointima formation in rat, pig and ApoE^{-/-} mouse arteries [9, 17, 40, 45]. However, these models are characterized by mechanical endovascular injury resulting in endothelium denudation, medial cell necrosis, and extensive VSMC proliferation. To the contrary, the model used in the present study is characterized by preserved endothelial integrity, as evidenced by the histologically intact endothelial layer and the observed relaxation responses to acetylcholine after eNOS gene transfer.

eNOS gene transfer may mediate vasoprotection through different biological mechanisms in different vascular disease conditions. NO-mediated inhibition of VSMC proliferation and platelet aggregation may play central roles in reducing neointima formation post-angioplasty, while NO-mediated regulation of VSMC differentiation may be important in vascular remodelling of venous bypass grafts [29, 46]. Regulation of vascular adhesion molecules, of MCP-1 expression, and of vascular inflammation are primary mechanisms by which NO protects vessels in the absence of mechanical endothelial injury [10, 35]. However, eNOS overexpression may not be invariably beneficial in diseased vessels. It is worth noting that eNOS expression in the endothelium overlying arterial lesions is reduced in the experimental model used in the present study [44]. Moreover, common carotid arteries (unlike the aorta and its branching sites) did not show histological lesions at the time of gene transfer. Under these conditions, eNOS gene transfer may restore, at least in part, eNOS expression in this model. Gene dosage is another important issue. While moderate eNOS overexpression (≈ 2.3 -fold increase) was beneficial in the present study, exaggerated eNOS expression might be detrimental due to increased superoxide

and peroxynitrite formation, especially in disease conditions that are associated with reduced bioavailability of NO cofactors (e.g., tetrahydrobiopterin) [30, 41]. "Uncoupling" of the NO synthase also involves peroxynitrite oxidation of tetrahydrobiopterin [37]. Our findings of improved endothelium-dependent relaxations in eNOS gene-transduced arteries indicates restored NO bioactivity, thus suggesting that the bioavailability of NO cofactors was not limiting in this model (although this has not been directly measured). It is worth re-emphasizing that our results must be interpreted in the framework to the experimental model used. As such, they are not necessarily applicable to other models or conditions, especially to more advanced stages of the disease.

The second gene construct tested was 8ND-MCP-1. A biological link between MCP-1 and eNOS is supported by observations that NO downregulates MCP-1 expression in endothelial cells, thereby inhibiting monocyte adhesion [26, 37, 38, 49]. Purified 8ND-MCP-1 peptide inhibits with high efficacy MCP-1 binding and signalling through the CCR2 receptor *in vitro* and MCP-1-mediated monocyte accumulation in inflamed joints in a mouse model of monocyte-rich arthritis *in vivo* [14]. In the present study, 8ND-MCP-1 gene transfer did not prevent lesion formation in carotid arteries in ApoE^{-/-} mice. Recently, two reports from Takeshita's group showed that systemic gene transfer of 7ND-MCP-1 (which differs from 8ND-MCP-1 by a single residue) [50] reduces lesion formation and progression in the aortic root in ApoE^{-/-} mice [15, 25]. A direct comparison of these data with our results is precluded by methodological differences regarding the following aspects: (1) Gene transfer vector (hemagglutinating virus of Japan-liposomes vs. adenovirus); (2) MCP-1 inhibitor (7ND vs. 8ND); (3) Gene dosage (two doses vs. single dose); (4) Target vessel (aortic root vs. carotid artery); (5) Atherogenic stimuli (Western-type diet vs. surgical manipulation and perivascular collar); and (6) Route of gene administration (systemic vs. arterial gene transfer in the previous and the present study, respectively). In spite of divergent conclusions as to the efficacy of ND-MCP-1 gene therapy in the respective models, however results from the previous [25] and the present study actually are not contradictory. Indeed, mean values for inhibition of lesion surface areas were similar in the two studies ($\approx 30\%$ and 27% , respectively). However, the decrease in lesion surface area did not reach statistical significance in the present study due to greater data variation. Surgical manipulation of the target vessel and implantation of a perivascular collar presumably contributed to inter-individual variation which appears to be inherent to the model [44]. For comparison of ND-MCP-1 expression, we injected our Ad.CMV-8ND-MCP-1 vector into remote muscles in a separate group of mice according to the protocol used in the two previous studies [15, 25]. Human ND-MCP-1 protein levels determined by ELISA in injected muscles

at 1 and 4 weeks post-gene transfer (data not shown) were comparable to those reported in the previous study [15], suggesting 8ND-MCP-1 expression levels from our adenoviral vector were adequate. It is conceivable that systemic gene transfer of a secreted ND-MCP-1 peptide may be more effective than direct arterial gene transfer because it blocks the CCR2 receptor on circulating monocytes. From the point of view of potential applications, however, systemic anti-MCP-1 gene therapy raises safety concerns because monocyte/macrophage activities are important mechanisms against infection and cancer.

Limitations of the present study include the short follow-up time (5 weeks) and the use of a perivascular collar. Because of the limited duration (\approx 2–6 weeks) of adenovirus-mediated transgene expression in mouse arteries [18, 42], it was necessary to stimulate arterial lesion formation within this time frame in order to assess the biological effects of the delivered genes. To achieve this goal, we used a recently described model [44, 45] based on a perivascular collar that acts by inducing flow disturbances that accelerate lesion formation, while maintaining blood flow. This model seemed highly preferable compared to traditional balloon-injury models because it preserves endothelial integrity, as discussed above.

Conclusion

eNOS gene transfer improved endothelium-dependent relaxations and inhibited lesion formation in carotid arteries in ApoE^{-/-} mice. This is one of the first reports on decreased lesion formation as a result of arterial gene transfer of a protective factor in a primary arteriosclerosis model. A previous report [18] showed that adenovirus-mediated gene transfer of heme oxygenase-1 (HO-1), an enzyme involved in iron metabolism and cytoprotection, inhibits lesion formation in the aorta in ApoE^{-/-} mice (6-week follow-up). Of note, the same vector was ineffective when delivered systemically, which emphasizes the role of localized gene delivery depending on the therapeutic gene. Gene transfer vectors such as adeno-associated virus (AAV) vectors that provide a potential for prolonged transgene expression in the vessel wall [34] could eventually achieve sustained therapeutic effects in vascular gene therapy. On the other hand, NO donors with a very long half-life could offer an alternate approach to eNOS gene therapy, which may warrant further investigation in preclinical arteriosclerosis models.

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