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## Genome-wide meta-analysis identifies six novel loci associated with habitual coffee consumption

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# These authors contributed equally to this work.

Coffee, a major dietary source of caffeine, is amongst the most widely consumed beverages in the world and has received considerable attention regarding health risks and benefits. We conducted a genome-wide(GW) meta-analysis of predominately regular-type coffee consumption (cups/day) among up to 91,462 coffee consumers of European ancestry with top single-nucleotide polymorphisms (SNPs) followed-up in ~30,062 and 7,964, coffee consumers of European and African American ancestry, respectively. Studies from both stages were combined in a trans-ethnic meta-analysis. Confirmed loci were examined for putative functional and biological relevance. Eight loci, including six novel loci, met GW-significance ( $\log_{10}$ Bayes-factor $>5.64$ ) with per allele effect sizes of 0.03-0.14 cups/day. Six are located in or near genes potentially involved in pharmacokinetics (*ABCG2*, *AHR*, *POR*, *CYP1A2*) and pharmacodynamics (*BDNF*, *SLC6A4*) of caffeine. Two map to *GCKR* and *MLXIPL*, genes related to metabolic traits but lacking known roles in coffee consumption. Enhancer and promoter histone marks populate the regions of many confirmed loci and several potential regulatory SNPs are highly correlated with the lead SNP of each. SNP alleles near *GCKR*, *MLXIPL*, *BDNF* and *CYP1A2* that were associated with higher coffee consumption have previously been associated with smoking initiation, higher adiposity and fasting insulin and glucose but lower blood pressure and favorable lipid, inflammatory and liver enzyme profiles ( $P<5\times 10^{-8}$ ). Our genetic findings among European and African American adults reinforce the role of caffeine in mediating habitual coffee consumption and may point to molecular mechanisms underlying inter-individual variability in pharmacological and health effects of coffee.

### INTRODUCTION

Coffee is amongst the most widely consumed beverages in the world<sup>1</sup>. North American coffee drinkers typically consume ~2 cups per day while the norm is at least 4 cups in many European countries<sup>1</sup>. In prospective cohort studies, coffee consumption is consistently associated with lower risk of Parkinson's disease, liver disease and type 2 diabetes<sup>2</sup>. However, the effects of coffee on cancer development, cardiovascular and birth outcomes and other health conditions remain controversial<sup>2</sup>. For most populations, coffee is the

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#### CONFLICTS OF INTEREST

The authors declare no conflict of interest.

Supplementary information is available at *Molecular Psychiatry's* website

primary source of caffeine, a stimulant also present in other beverages, foods and medications<sup>1,3</sup>. The fifth edition of the Diagnostic and Statistical Manual of Mental Disorders does not include a diagnosis of caffeine dependence or abuse due to a paucity of evidence but lists caffeine intoxication and withdrawal as Disorders<sup>4</sup>. Knowledge of factors contributing to coffee's consumption and physiological effects may greatly advance the design and interpretation of population and clinical research on coffee and caffeine<sup>5</sup>. Genetic factors could be especially valuable as they offer ways to study the potential health effects of coffee via instrumental variables or gene-environment interactions<sup>5</sup>. Heritability estimates for coffee and caffeine use range between 36 and 58%<sup>6</sup>. Genome-wide association studies (GWAS) of habitual caffeine and coffee intake have identified variants near *CYP1A2* and *AHR*<sup>7-9</sup>. Cytochrome P450 (CYP)1A2 is responsible for ~95% of caffeine metabolism in humans and aryl hydrocarbon receptor (AHR) plays a regulatory role in basal and substrate-induced expression of target genes, including *CYP1A1* and *CYP1A2*<sup>10,11</sup>.

To identify additional loci, we conducted a staged GW meta-analysis of coffee consumption including over 120,000 coffee consumers sourced from population-based studies of European and African American ancestry.

## MATERIALS AND METHODS

### Study design and populations

Supplementary Figure S1 depicts an overview of the current study. We performed a meta-analysis of GWAS summary statistics from 28 population-based studies of European ancestry to detect single-nucleotide polymorphisms (SNPs) that are associated with coffee consumption. Top loci were followed-up in studies of European (13 studies) and African American (7 studies) ancestry and confirmed loci were explored in a single Pakistani population. Detailed information on study design, participant characteristics, genotyping and imputation for all contributing studies are provided in the Supplementary Information and Supplementary Tables S1-S6.

### Phenotype

All phenotype data were previously collected via interviewer- or self-administered questionnaires (Supplementary Table S1). Our primary phenotype ('phenotype 1') was cups of predominately regular-type coffee consumed per day among coffee consumers. Coffee data collected categorically (e.g. 2-3 cups/day) was converted to cups/day by taking the median value of each category (e.g. 2.5 cups/day). A secondary analysis was performed comparing high to infrequent/non- coffee consumers ('phenotype 2'). A subset of stage 1 studies collected information on decaffeinated coffee consumption; which was examined in follow-up analysis of the confirmed loci.

### Statistical analysis

Each stage 1 (discovery) study performed GWA-testing for each phenotype across ~2.5 million genotyped or imputed autosomal SNPs (HapMap II, Centre d'Etude du Polymorphisme Humain [CEU] reference), based on linear (cups/day, phenotype-1) or logistic (high vs. none/low, phenotype-2) regression under an additive genetic model.

Analyses were adjusted for age, smoking status and, when applicable, sex, case-control status, study-site, family structure and/or study specific principal components of population substructure (Supplementary Table S7). SNPs with minor allele frequency (MAF) <0.02 or with low imputation quality scores were removed prior to meta-analysis (Supplementary Table S5). The GWAtoolbox (see Supplementary Information for URLs) was used for initial quality control. MAFs and a plot comparing (1/median standard error of effect size) vs. (square root of sample size) for each study were also reviewed for outliers and these were addressed prior to the final meta-analysis.

For both phenotypes, GW meta-analysis was conducted using a fixed effects model and inverse-variance weighting with a single genomic control (GC) correction as implemented in METAL<sup>12</sup> and GWAMA<sup>13</sup> ( $r>0.99$  for correlation between METAL and GWAMA results). The phenotypic variance explained by additive SNP effects was estimated in the Women's Genome Health Study (WGHS,  $n=15,987$  with identity-by-state <0.025) using GCTA<sup>14</sup>. Stage 1 summary statistics were also subject to pathway analysis using MAGENTA<sup>15</sup> (Supplementary Information).

For regions achieving association  $P$  values  $<5\times 10^{-8}$  (7p21, 7q23.11, 11p13, 15q24) we performed conditional analysis using the summary statistics from the meta-analysis to test for the association of each SNP while conditioning on the top SNPs, with correlations between SNPs due to linkage disequilibrium (LD) estimated from the imputed genotype data from the Atherosclerosis Risk in Communities cohort<sup>16</sup>, a large and representative cohort of men and women of European ancestry.

Our approach to selecting SNPs for replication (stage 2) is described in Supplementary Information. Stage 2 meta-analyses were performed separately for European and African American populations, using the same statistical models and methods as described for stage 1, but without GC (Supplementary Information).

Studies from all stages were included in an overall meta-analysis using Meta-ANalysis of TRans-ethnic Association studies (MANTRA)<sup>17</sup>; which adopts a Bayesian framework to combine results from different ethnic groups by taking advantage of the expected similarity in allelic effects between the most closely related populations. MANTRA was limited to SNPs selected for replication thus no GC was applied. A random effects analysis using GWAMA was performed in parallel to obtain effect estimates, which are not generated by MANTRA. The GW-significance threshold of  $\log_{10} BF >5.64$  approximates a traditional GW P-value threshold of  $5\times 10^{-8}$  under general assumptions<sup>18, 19</sup>. Subgroup analysis and meta-regression were performed to investigate possible sources of between-study heterogeneity (Supplementary Information). *Fine-mapping*: To assess the improvement in fine-mapping resolution due to trans-ethnic meta-analysis we applied the methods of Franceschini *et al*<sup>17</sup> to stage 1 and stage 2 (African Americans only) GW-summary level data (Supplementary Information).

### Potential SNP-function and biological and clinical inferences

Details pertaining to follow-up of confirmed loci are provided in the Supplementary Information. Briefly, all confirmed index SNPs and their correlated proxies were examined

for putative function using publicly available resources. Bioinformatics and computational tools were used to systematically mine available knowledge and experimental databases to inform biological hypotheses underlying the link between loci and coffee consumption as well as connections between loci. For these analyses all genes mapping to the confirmed regions were considered potential candidates. Finally, we searched the National Human Genome Research Institute GWAS catalogue<sup>20</sup> and Metabolomics GWAS server<sup>21</sup> for all GW-significant associations with our confirmed coffee-SNPs. Complete GWAS summary data for coffee-implicated diseases or traits were additionally queried.

## RESULTS

### SNPs associated with coffee consumption

**Discovery stage**—Results from the discovery stage are summarized in Supplementary Figs S2-S5. Little evidence for genomic inflation ( $<1.07$ ) was observed for either phenotype. The two analyses yielded similarly ranked loci and significant enrichment of ‘xenobiotic’ genes (MAGENTA’s  $FDR < 0.006$ ), suggesting no major difference in the genetic influence on coffee drinking initiation compared with the level of coffee consumption among coffee consumers at these loci. Overall, approximately 7.1% (standard error: 2%) of the variance in coffee cups consumed per day (phenotype-1) could be explained by additive and common SNP effects in the WGHS.

Conditioning on the index SNPs of each region achieving association  $P$  values  $< 5 \times 10^{-8}$  (7p21, 7q23.11, 11p13, 15q24) in the discovery stage provided little evidence for multiple independent variants (Supplementary Figure S6). Only four of the SNPs on chromosome 7 were potentially independent and carried forward with other promising SNPs.

**Replication and trans-ethnic meta-analysis**—Forty-four SNPs spanning 33 genomic regions met significance criteria for candidate associations and were followed-up in stage 2 (Supplementary Tables S8-S13). Eight loci, including six novel, met our criteria for GW-significance ( $\log_{10} BF > 5.64$ ) in a trans-ethnic meta-analysis of all discovery and replication studies (Table 1, Supplementary Tables S14-S16, Supplementary Figs S7 and S8). Confirmed loci have effect sizes of 0.03-0.14 cups/day per allele and together explain ~1.3% of the phenotypic variance of coffee intake. We were underpowered to replicate these associations in a Pakistani population (Supplementary Information).

### Functional and biological inferences

Enhancer (H3K4me1) and promoter (H3K4me3) histone marks densely populate many of these regions and several nonsynonymous and potential regulatory SNPs are highly correlated ( $r^2 > 0.8$ ) with the lead SNP and thus strong candidates for being a causal variant (Table 2, Supplementary Information, Supplementary Tables S17-S19). Candidate genes form a highly connected network of interactions, featuring discernible clusters of genes around *BDNF* and *AHR* (Figure 1, Supplementary Information, Supplementary Tables S20 and S21). At least one gene in each of the eight regions i) is highly expressed in brain, liver and/or taste buds, ii) results in phenotype abnormalities relevant to coffee consumption behavior when modified in mice, and iii) is differentially expressed in human hepatocytes

when treated with high (7500 M) but not low (1500 M) doses of caffeine (Table 2, Supplementary Tables S22-S24).

Additional genomic characterization of the top loci allows further biological inference as follows:

**i.** Previously identified loci near *AHR* (7p21) and *CYP1A2* (15q24)

Consistent with previous reports in smaller samples<sup>7-9</sup>, the intergenic 7p21 and 15q24 loci near *AHR* and *CYP1A1/CYP1A2* respectively remained the most prominent and highly heterogeneous loci associated with coffee consumption. The same index SNPs were identified in European and African Americans suggesting they are robust HapMap proxies for causal variants in these two populations. Cohort-wide mean coffee consumption explained part of the heterogeneity in study results for both loci (Supplementary Table S25, Supplementary Information). The rs2472297 T and rs4410790 C alleles associated with increased coffee consumption have recently been associated with lower plasma caffeine levels<sup>21</sup> and shown to increase *CYP1A2*-mediated metabolism of olanzapine<sup>22</sup>. The C allele of rs4410790 is also positively correlated with cerebellum *AHR* methylation, suggesting a novel role of *Ahr* in motor or learning pathways that may trigger coffee consumption. The most significant variants at 15q24 reside in the *CYP1A1-CYP1A2* bidirectional promoter where *AHR* response elements have been identified and shown to be important for transcriptional activation of both *CYP1A1* and *CYP1A2*<sup>23</sup>. The rs2472297 T variant putatively weakens the binding of SP1, a co-activator in the *Ahr*-Arnt complex regulating *CYP1* locus transcription<sup>24</sup> and is also implicated in the expression of several neighboring genes. The latter observation, together with this region's high LD and long range chromatin interactions (Supplementary Figure S9), suggests a regulatory network among these genes.

**ii.** Novel loci at 7q11.23 (*POR*) and 4q22 (*ABCG2*) likely function in caffeine metabolism

Variants at 7q11.23 (rs17685) and 4q22 (rs1481012) map to novel yet biologically plausible candidate genes involved in xenobiotic metabolism. rs17685 maps to the 3'UTR of *POR*, encoding P450 oxidoreductase which transfers electrons to all microsomal *CYP450s* enzymes<sup>25</sup>. The rs17685 A variant associated with higher coffee consumption is linked to increased *POR* expression and potentially weakens the DNA binding of several transcriptional regulatory proteins including BHLHE40, which inhibits *POR* expression<sup>26</sup>. The same SNP is in LD (CEU:  $r^2=0.93$ ) with *POR*\*28 (rs1057868, Ala503Val), which is associated with differential *CYP* activity depending on the *CYP* isoform, substrate and experimental model used<sup>27</sup>. rs1481012 at 4q22 maps to *ABCG2*, encoding a xenobiotic efflux transporter. rs1481012 is in LD (CEU:  $r^2=0.92$ ) with rs2231142 (Gln141Lys), a functional variant at an evolutionarily constrained residue<sup>28</sup>. However, fine-mapping of this region on the basis of reduced LD in the African American sample limited an initial 189102 kb region to a credible span of 6249 kb (Supplementary Table S16), that excluded rs2231142.

- iii. Novel loci at 11p13 (*BDNF*) and 17q11.2 ('*SLC6A4*') likely mediate the positive reinforcing properties of coffee constituents

The index SNP at 11p13 is the widely investigated missense mutation (rs6265, Val66Met) in *BDNF* (Supplementary Table S26). Brain-derived neurotrophin factor (*BDNF*) modulates the activity of serotonin, dopamine and glutamate, neurotransmitters involved in mood-related circuits and plays a key role in memory and learning<sup>29</sup>. The Met66 allele impairs neuronal activity-dependent *BDNF* secretion<sup>30</sup> and thus may attenuate the rewarding effects of coffee and, in turn, motivation to consume coffee. The increasingly recognized roles of *BDNF* in the chemosensory system and conditioned taste preferences may also be relevant<sup>31</sup>. The index SNP (rs9902453) at 17q11.2 maps to the *EFCAB5* gene and is in LD (CEU:  $r^2 > 0.8$ ) with SNPs that alter regulatory motifs for AhR<sup>32</sup> in the neighboring gene *NSRP1*, but neither gene is an obvious candidate for coffee consumption. Upstream of rs9902453 lies a possibly stronger candidate: *SLC6A4* encoding the serotonin transporter. Serotonergic neurotransmission affects a wide range of behaviors including sensory processing and food-intake<sup>33</sup>.

- iv. Novel loci at 2p24 (*GCKR*) and 7q11.2 (*MLXIPL*)

Variants at 2p24 (rs1260326) and 7q11.23 (rs7800944) map to *GCKR* and *MLXIPL*, respectively. The former has been associated with plasma glucose and multiple metabolic traits and the latter with plasma triglycerides (Table 3, Supplementary Table S27). Adjustment of regression models for plasma lipids in the Women's Genome Health Study (WGHS,  $n \sim 17,000$ ) and plasma glucose in TwinGene ( $n \sim 8,800$ ) did not significantly change the relationship between SNPs at these two loci and coffee consumption ( $P > 0.48$ , Supplementary Tables S28 and S29). The rs1260326 T allele encodes a nonsynonymous change in the encoded, glucokinase regulatory protein (GKRP) leading to increased hepatic glucokinase activity<sup>34</sup>. GKRP and glucokinase may also cooperatively function in the glucose-sensing process of the brain<sup>35</sup> that may, in turn, influence central pathways responding to coffee constituents. A direct link between *MLXIPL* and coffee consumption remains unclear, except for the interactions with other candidate genes (Figure 1). Experimental evidence and results from formal prioritization analyses also warrants consideration of other candidates in these regions (Table 2, Figure 1, Supplementary Tables S23, S26, S30, S31). For example, in the frontal cortex, the rs1260326 allele positively associated with coffee consumption correlates with lower methylation of *PPM1G*; a putative regulatory target for AhR and binding target for *PPP1R1B*, which mediates psychostimulant effects of caffeine<sup>36</sup>.

### Pleiotropy and clinical inferences

None of the eight loci was significantly associated with caffeine taste-intensity ( $P > 0.02$ ) or caffeine-induced insomnia ( $P > 0.08$ ), according to previously published GWAS of these traits<sup>37-39</sup>. SNPs near *AHR* associated with higher coffee consumption were also significantly associated with higher decaffeinated coffee consumption ( $\sim 0.05$  cups/day,

$P < 0.0004$ ,  $n = 24,426$ ); perhaps a result of Pavlovian conditioning among individuals moderating their intake of regular coffee or the small amounts of caffeine in decaffeinated coffee<sup>1</sup>.

Across phenotypes in the GWAS catalog<sup>20</sup>, the alleles leading to higher coffee consumption at 2p24, 4q22, 7q11.23, 11p13 and 15q24 have been associated with one or more of the following: smoking initiation, higher adiposity and fasting insulin and glucose but lower blood pressure and favorable lipid, inflammatory and liver enzyme profiles ( $P < 5 \times 10^{-8}$ , Table 3, Supplementary Table S27). Focused on metabolic, neurologic and psychiatric traits for which coffee has been implicated (Table 3, Supplementary Table S32), there were additional sub-GW significant associations in published GWAS. Variants associated with higher coffee consumption increased adiposity (rs1481012,  $P = 4.85 \times 10^{-3}$ ), birth weight (rs7800944,  $P = 2.10 \times 10^{-3}$ ), plasma high-density lipoprotein (HDL, rs7800944,  $P = 2.24 \times 10^{-3}$ ), risk of Parkinson's disease (rs1481012,  $P = 7.11 \times 10^{-3}$ ), reduced blood pressure (rs6265,  $P = 6.58 \times 10^{-4}$ ; rs2472297,  $P < 6.80 \times 10^{-5}$  and rs9902453,  $P = 6.05 \times 10^{-3}$ ), HDL (rs6968554,  $P = 1.18 \times 10^{-3}$ ), risk of major depressive disorder (rs17685,  $P = 6.98 \times 10^{-3}$ ) and bipolar disorder (rs1260326,  $P = 2.31 \times 10^{-3}$ ). Associations with adiposity, birth weight, blood pressure, HDL and bipolar disorder remain significant after correcting for the number of SNPs tested.

## DISCUSSION

Coffee's widespread popularity and availability has fostered public health concerns of the potential health consequences of regular coffee consumption. Findings from epidemiological studies of coffee consumption and certain health conditions remain controversial<sup>2</sup>. Knowledge of genetic factors contributing to coffee's consumption and physiological effects may inform the design and interpretation of population and clinical research on coffee<sup>5</sup>. In the current report, we present results of the largest GWAS of coffee intake to-date and the first to include populations of African American ancestry. In addition to confirming associations with *AHR* and *CYP1A2*, we have identified six new loci, not previously implicated in coffee drinking behavior.

Our findings highlight an important role of the pharmacokinetic and pharmacodynamic properties of the caffeine component of coffee underlying a genetic propensity to consume the beverage. Loci near *BDNF* and *SLC6A4* potentially impact consumption behavior by modulating the acute behavioral and reinforcing properties of caffeine. Others near *AHR*, *CYP1A2*, *POR*, and *ABCG2* act indirectly by altering the metabolism of caffeine and thus the physiological levels of this stimulant. The strength of these four associations with coffee intake, along with results from pathway analysis showing significant enrichment for 'xenobiotic' genes, emphasize an especially pronounced role of caffeine metabolism in coffee drinking behavior. The current study is the first to link *GCKR* and *MLXIPL* variation to a behavioral trait. The nonsynonymous rs1260326 SNP in *GCKR* has been a GW-signal for various metabolic traits particularly those reflecting glucose-homeostasis (Table 3). *GCKR* variation may impact the glucose-sensing process of the brain<sup>35</sup> that may, in turn, influence central pathways responding to coffee constituents. mQTL and binding motif analysis suggests *PPM1G* may be another candidate underlying the association between



rs1260326 and coffee consumption. Variants near *MLXIPL* have also topped the list of variants associated with plasma triglycerides (Table 3), but their link to coffee consumption remains unclear. Future studies on the potential pleiotropic effects of these two loci are clearly warranted. Interestingly, several candidate genes implicated in coffee consumption behavior, but not confirmed in our GWAS, interact with one or more of the eight confirmed loci (Figure 1). While these findings are encouraging for ongoing efforts they also emphasize the need to study sets or pathways of genes in the future.

Specific SNPs associated with higher coffee consumption have previously been associated with smoking initiation, higher adiposity and fasting insulin and glucose but lower blood pressure and favorable lipid, inflammatory and liver enzyme profiles. Whether these relationships reflect pleiotropy, confounding, or offer insight to the potential causal role coffee plays in these traits merits further investigation. Future research, particularly Mendelian Randomization and gene-coffee interaction studies, will need to consider the direct and indirect roles that each SNP has in altering coffee drinking behavior as well as the potential for interactions between loci (Figure 1). The heterogeneous effects specific to *AHR*- and *CYP1A2*-coffee associations point to SNP-specific interactions with the environment or population characteristics that might also warrant consideration (Supplementary Information).

The strong cultural influences on norms of coffee drinking may have reduced our power for loci discovery. This might, in part, underlie our lack of replication in a Pakistani population, wherein coffee consumption is extremely rare. Methodological limitations specific to our approach may also have reduced our power for loci discovery or precision in estimating effect sizes (Supplementary Information). For example, some studies collected coffee data in categories of cups/day (e.g. 2-3 cups/day) rendering a less precise record of intake as well as a non-Gaussian distributed trait for analysis. The precise chemical composition of different coffee preparations is also not captured by standard FFQs and is likely to vary within and between populations. Nevertheless, the eight loci together explain ~1.3% of the phenotypic variance, a value *at least* as great as that reported for smoking behavior and alcohol consumption which are subject to similar limitations in GWAS<sup>40, 41</sup>.

The additive genetic variance (or narrow-sense heritability) of coffee intake as estimated by GCTA in WGHS (7%) is considerably lower than estimates based on pedigrees (36 to 57%)<sup>6</sup>. The marked discrepancies between the GCTA and pedigree estimates of heritability may be due to one or more of the following: the potential contribution of rare variants to heritability (not captured by GCTA's 'chip-based heritability'), biases in pedigree analysis resulting in overestimates of heritability, differences in phenotype ascertainment or definition, and cultural differences in the populations studied<sup>42</sup>.

In conclusion, our results support the hypothesis that metabolic and neurological mechanisms of caffeine contribute to coffee consumption habits. Individuals adapt their coffee consumption habits to balance perceived negative and reinforcing symptoms that are affected by genetic variation. Genetic control of this potential 'titrating' behavior would incidentally govern exposure to other potentially 'bioactive' constituents of coffee that may be related to the health effects of coffee or other sources of caffeine. Thus, our findings may

point to molecular mechanisms underlying inter-individual variability in pharmacological and health effects of coffee and caffeine.

## Supplementary Material

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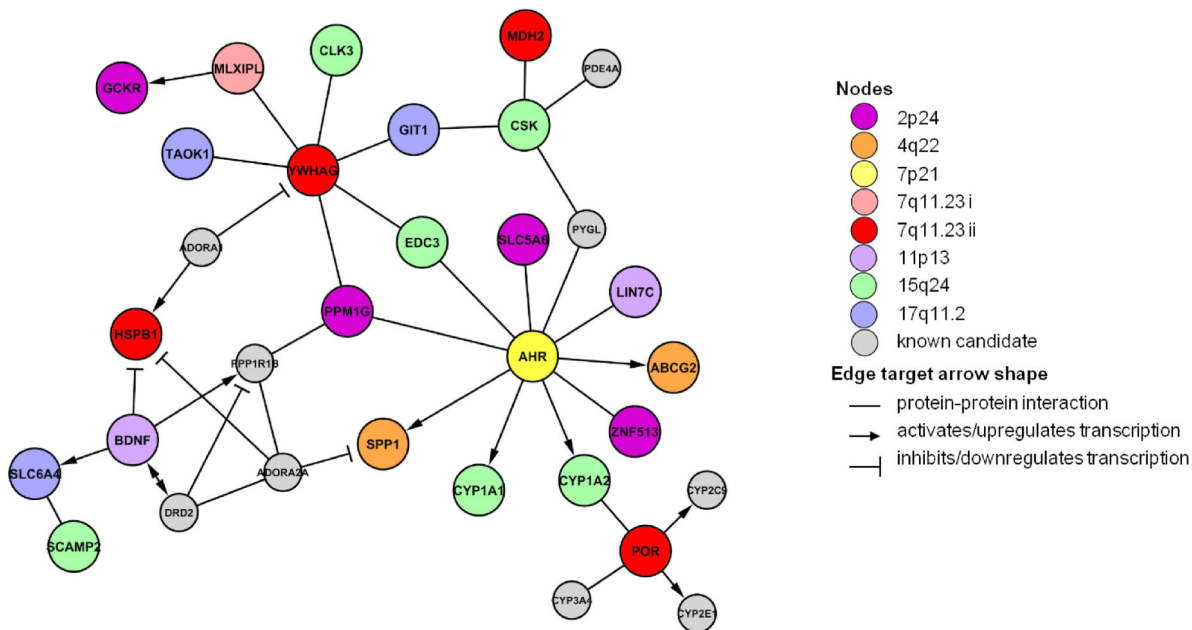
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## REFERENCES

1. Fredholm BB, Battig K, Holmen J, Nehlig A, Zvartau EE. Actions of caffeine in the brain with special reference to factors that contribute to its widespread use. *Pharmacol Rev.* 1999; 51(1):83–133. [PubMed: 10049999]
2. Cornelis M. Gene-coffee interactions and health. *Curr Nutr Rep.* 2014 in press.
3. Spiller, MA. The chemical components of coffee. In: Spiller, GA., editor. *Caffeine.* CRC; Boca Raton: 1998. p. 97-161.
4. American Psychiatric Association. *Diagnostic and statistical manual of mental disorders.* 5 edn.. Arlington, VA; American Psychiatric Publishing: 2013.
5. Cornelis MC. Coffee intake. *Progress in molecular biology and translational science.* 2012; 108:293–322. [PubMed: 22656382]
6. Yang A, Palmer AA, de Wit H. Genetics of caffeine consumption and responses to caffeine. *Psychopharmacology.* 2010; 211(3):245–257. [PubMed: 20532872]
7. Cornelis MC, Monda KL, Yu K, Paynter N, Azzato EM, Bennett SN, et al. Genome-wide meta-analysis identifies regions on 7p21 (AHR) and 15q24 (CYP1A2) as determinants of habitual caffeine consumption. *PLoS Genet.* 2011; 7(4):e1002033. [PubMed: 21490707]
8. Sulem P, Gudbjartsson DF, Geller F, Prokopenko I, Feenstra B, Aben KK, et al. Sequence variants at CYP1A1-CYP1A2 and AHR associate with coffee consumption. *Hum Mol Genet.* 2011; 20(10):2071–2077. [PubMed: 21357676]
9. Amin N, Byrne E, Johnson J, Chenevix-Trench G, Walter S, Nolte IM, et al. Genome-wide association analysis of coffee drinking suggests association with CYP1A1/CYP1A2 and NRCAM. *Molecular psychiatry.* 2011; 17(11):1116–1129. [PubMed: 21876539]
10. Kot M, Daniel WA. The relative contribution of human cytochrome P450 isoforms to the four caffeine oxidation pathways: an in vitro comparative study with cDNA-expressed P450s including CYP2C isoforms. *Biochemical pharmacology.* 2008; 76(4):543–551. [PubMed: 18619574]
11. Le Vee M, Jouan E, Fardel O. Involvement of aryl hydrocarbon receptor in basal and 2,3,7,8-tetrachlorodibenzo-p-dioxin-induced expression of target genes in primary human hepatocytes. *Toxicol In Vitro.* 2010; 24(6):1775–1781. [PubMed: 20619336]
12. Willer CJ, Li Y, Abecasis GR. METAL: fast and efficient meta-analysis of genomewide association scans. *Bioinformatics.* 2010; 26(17):2190–2191. [PubMed: 20616382]
13. Mägi R, Morris AP. GWAMA: software for genome-wide association meta-analysis. *BMC bioinformatics.* 2010; 11:288. [PubMed: 20509871]
14. Yang J, Lee SH, Goddard ME, Visscher PM. GCTA: a tool for genome-wide complex trait analysis. *Am J Hum Genet.* 2011; 88(1):76–82. [PubMed: 21167468]
15. Segre AV, Groop L, Mootha VK, Daly MJ, Altshuler D. Common inherited variation in mitochondrial genes is not enriched for associations with type 2 diabetes or related glycemic traits. *PLoS Genet.* 2010; 6(8):e1001058. [PubMed: 20714348]
16. Yang J, Ferreira T, Morris AP, Medland SE, Madden PA, Heath AC, et al. Conditional and joint multiple-SNP analysis of GWAS summary statistics identifies additional variants influencing complex traits. *Nat Genet.* 2012; 44(4):369–375. S361-363. [PubMed: 22426310]
17. Franceschini N, van Rooij FJ, Prins BP, Feitosa MF, Karakas M, Eckfeldt JH, et al. Discovery and fine mapping of serum protein loci through transethnic meta-analysis. *Am J Hum Genet.* 2012; 91(4):744–753. [PubMed: 23022100]
18. Stephens M, Balding DJ. Bayesian statistical methods for genetic association studies. *Nat Rev Genet.* 2009; 10(10):681–690. [PubMed: 19763151]
19. Sellke T, Bayarri M, Berger J. Calibration of p values for testing precise null hypotheses. *Am Stat.* 2001; 55(1):62–71.

20. Hindorf L, MacArthur J, Morales J, Junkins H, Hall P, Klemm A, et al. Catalogue of Published Genome-Wide Association Studies.
21. Shin SY, Fauman EB, Petersen AK, Krumsiek J, Santos R, Huang J, et al. An atlas of genetic influences on human blood metabolites. *Nat Genet.* 2014; 46(6):543–550. [PubMed: 24816252]
22. Soderberg MM, Haslemo T, Molden E, Dahl ML. Influence of CYP1A1/CYP1A2 and AHR polymorphisms on systemic olanzapine exposure. *Pharmacogenet Genomics.* 2013; 23(5):279–285. [PubMed: 23492908]
23. Jorge-Nebert LF, Jiang Z, Chakraborty R, Watson J, Jin L, McGarvey ST, et al. Analysis of human CYP1A1 and CYP1A2 genes and their shared bidirectional promoter in eight world populations. *Hum Mutat.* 2010; 31(1):27–40. [PubMed: 19802894]
24. Swanson HI. DNA binding and protein interactions of the AHR/ARNT heterodimer that facilitate gene activation. *Chemico-biological interactions.* 2002; 141(1-2):63–76. [PubMed: 12213385]
25. Hu L, Zhuo W, He YJ, Zhou HH, Fan L. Pharmacogenetics of P450 oxidoreductase: implications in drug metabolism and therapy. *Pharmacogenet Genomics.* 2012; 22(11):812–819. [PubMed: 23047293]
26. Rome S, Meugnier E, Lecomte V, Berbe V, Besson J, Cerutti C, et al. Microarray analysis of genes with impaired insulin regulation in the skeletal muscle of type 2 diabetic patients indicates the involvement of basic helix-loop-helix domain-containing, class B, 2 protein (BHLHB2). *Diabetologia.* 2009; 52(9):1899–1912. [PubMed: 19590847]
27. Pandey AV, Fluck CE. NADPH P450 oxidoreductase: structure, function, and pathology of diseases. *Pharmacol Ther.* 2013; 138(2):229–254. [PubMed: 23353702]
28. Woodward OM, Tukaye DN, Cui J, Greenwell P, Constantoulakis LM, Parker BS, et al. Gout-causing Q141K mutation in ABCG2 leads to instability of the nucleotide-binding domain and can be corrected with small molecules. *Proc Natl Acad Sci U S A.* 2013; 110(13):5223–5228. [PubMed: 23493553]
29. Numakawa T, Suzuki S, Kumamaru E, Adachi N, Richards M, Kunugi H. BDNF function and intracellular signaling in neurons. *Histology and histopathology.* 2010; 25(2):237–258. [PubMed: 20017110]
30. Egan MF, Kojima M, Callicott JH, Goldberg TE, Kolachana BS, Bertolino A, et al. The BDNF val66met polymorphism affects activity-dependent secretion of BDNF and human memory and hippocampal function. *Cell.* 2003; 112(2):257–269. [PubMed: 12553913]
31. Nosrat IV, Margolskee RF, Nosrat CA. Targeted taste cell-specific overexpression of brain-derived neurotrophic factor in adult taste buds elevates phosphorylated TrkB protein levels in taste cells, increases taste bud size, and promotes gustatory innervation. *J Biol Chem.* 2012; 287(20):16791–16800. [PubMed: 22442142]
32. Ernst J, Kheradpour P, Mikkelsen TS, Shores N, Ward LD, Epstein CB, et al. Mapping and analysis of chromatin state dynamics in nine human cell types. *Nature.* 2011; 473(7345):43–49. [PubMed: 21441907]
33. Canli T, Lesch KP. Long story short: the serotonin transporter in emotion regulation and social cognition. *Nature neuroscience.* 2007; 10(9):1103–1109.
34. Beer NL, Tribble ND, McCulloch LJ, Roos C, Johnson PR, Orho-Melander M, et al. The P446L variant in GCKR associated with fasting plasma glucose and triglyceride levels exerts its effect through increased glucokinase activity in liver. *Hum Mol Genet.* 2009; 18(21):4081–4088. [PubMed: 19643913]
35. Alvarez E, Roncero I, Chowen JA, Vazquez P, Blazquez E. Evidence that glucokinase regulatory protein is expressed and interacts with glucokinase in rat brain. *Journal of neurochemistry.* 2002; 80(1):45–53. [PubMed: 11796742]
36. Lindskog M, Svenningsson P, Pozzi L, Kim Y, Fienberg AA, Bibb JA, et al. Involvement of DARPP-32 phosphorylation in the stimulant action of caffeine. *Nature.* 2002; 418:774–778. [PubMed: 12181566]
37. Reed DR, Zhu G, Breslin PA, Duke FF, Henders AK, Campbell MJ, et al. The perception of quinine taste intensity is associated with common genetic variants in a bitter receptor cluster on chromosome 12. *Hum Mol Genet.* 2010; 19(21):4278–4285. [PubMed: 20675712]

38. Ledda M, Kotalik Z, Souza Destito MC, Souza MM, Cirillo CA, Zamboni A, et al. GWAS of Human Bitter Taste Perception Identifies New Loci and Reveals Additional Complexity of Bitter Taste Genetics. *Hum Mol Genet.* 2013; 23(1):259–267. [PubMed: 23966204]
39. Byrne EM, Johnson J, McRae AF, Nyholt DR, Medland SE, Gehrman PR, et al. A genome-wide association study of caffeine-related sleep disturbance: confirmation of a role for a common variant in the adenosine receptor. *Sleep.* 2012; 35(7):967–975. [PubMed: 22754043]
40. Tobacco and Genetics Consortium. Genome-wide meta-analyses identify multiple loci associated with smoking behavior. *Nat Genet.* 2010; 42(5):441–447. [PubMed: 20418890]
41. Schumann G, Coin LJ, Lourdasamy A, Charoen P, Berger KH, Stacey D, et al. Genome-wide association and genetic functional studies identify autism susceptibility candidate 2 gene (AUTS2) in the regulation of alcohol consumption. *Proc Natl Acad Sci U S A.* 2011; 108(17):7119–7124. [PubMed: 21471458]
42. Vinkhuyzen AA, Wray NR, Yang J, Goddard ME, Visscher PM. Estimation and partition of heritability in human populations using whole-genome analysis methods. *Annu Rev Genet.* 2013; 47:75–95. [PubMed: 23988118]



**Figure 1.** Network describing direct interactions between candidate genes of confirmed loci. Relationships were retrieved from databases of transcription regulation and protein-protein interaction experiments (Supplementary Table S21). Genes are represented as nodes that are colored according to locus. Candidate genes for loci identified in the current study were supplemented with known candidate genes related to caffeine pharmacology (grey nodes). Edges indicate known interactions.

**Table 1**

SNPs associated with cups of coffee consumed per day among coffee consumers

Locus	Index SNP <sup>d</sup>	Closest Gene	EA/ NEA	EAF EUR/AA	Stage 1 <sup>b</sup> EUR n 91,462		Stage 2 <sup>b</sup> EUR n 30,062		AA n 7,964		Trans-Ethnic Meta-Analysis <sup>c</sup>			
					β (SE)	P	β (SE)	P	β (SE)	P	β (SE)	Log <sub>10</sub> BF	Post Prob	n
2p24	<b>rs1260326</b>	GCKR	T/C	.41/.17	-0.04 (0.01)	1.06 × 10 <sup>-07</sup>	-0.03 (0.01)	0.02	-0.01 (0.03)	0.77	-0.04 (0.01)	6.48	0.07	129,417
4q22	<b>rs1481012</b>	ABCG2	A/G	.89/.95	0.06 (0.01)	1.13 × 10 <sup>-06</sup>	0.03 (0.02)	0.11	0.16 (0.05)	1.27 × 10 <sup>-03</sup>	0.06 (0.01)	6.08	0.23	126,019
7p21	rs4410790 rs6968554	AHR	T/C A/G	.37/.52 .39/.33	-0.14 (0.01) -0.13 (0.01)	1.48 × 10 <sup>-57</sup> 2.54 × 10 <sup>-57</sup>	-0.05 (0.01) -0.07 (0.01)	1.66 × 10 <sup>-04</sup> 2.78 × 10 <sup>-10</sup>	-0.09 (0.02) -0.05 (0.02)	2.37 × 10 <sup>-06</sup> 0.02	-0.10 (0.01) -0.10 (0.01)	58.87 69.69	0.96 1.00	116,674 124,849
7q11.23	<b>rs7800944</b>	MLXIPL	T/C	.72/.67	-0.05 (0.01)	7.82 × 10 <sup>-09</sup>	-0.06 (0.02)	4.20 × 10 <sup>-04</sup>	-0.02 (0.02)	0.37	-0.05 (0.01)	8.83	0.09	116,417
7q11.23	<b>rs17685</b>	POR	A/G	.29/.19	0.07 (0.01)	9.06 × 10 <sup>-14</sup>	0.05 (0.01)	1.01 × 10 <sup>-03</sup>	0.07 (0.03)	7.55 × 10 <sup>-03</sup>	0.07 (0.01)	15.12	0.08	115,465
11p13	<b>rs6265</b>	BDNF	T/C	.19/.07	-0.05 (0.01)	3.40 × 10 <sup>-07</sup>	-0.03 (0.01)	0.07	-0.05 (0.04)	0.25	-0.04 (0.01)	5.76	0.10	127,828
15q24	rs2470893 rs2472297	CYP1A1, CYP1A2	T/C T/C	.31/.06 .24/.06	0.12 (0.01) 0.15 (0.01)	6.89 × 10 <sup>-44</sup> 6.45 × 10 <sup>-47</sup>	0.09 (0.01) 0.11 (0.01)	9.92 × 10 <sup>-11</sup> 3.26 × 10 <sup>-16</sup>	0.20 (0.07) 0.19 (0.05)	4.23 × 10 <sup>-03</sup> 8.62 × 10 <sup>-05</sup>	0.12 (0.01) 0.14 (0.01)	57.79 62.77	1.00 0.97	113,273 116,272
17q11.2	<b>rs9902453</b>	EFCAB5	A/G	.54/.80	-0.04 (0.01)	2.26 × 10 <sup>-06</sup>	-0.03 (0.01)	9.13 × 10 <sup>-03</sup>	-0.04 (0.03)	0.17	-0.03 (0.01)	6.29	0.05	126,819

Abbreviations: EA, effect allele; NEA, non-effect allele; EAF, effect allele frequency; EUR, European ancestry; AA, African American ancestry; AA, African American ancestry; SE, standard error; BF, Bayes-factor; Post Prob, posterior probability

<sup>a</sup> Genic SNPs are in bold-face.

<sup>b</sup> Effect coefficients (SE), representing cups/day per effect allele, and corresponding P-values from stage 1 fixed-effects meta-analysis (columns 6, 7) and stage 2 fixed-effects meta-analyses (columns 8-11).

<sup>c</sup> Effect coefficients (SE), representing cups/day per effect allele, from random-effects meta-analysis of all stage 1 and stage 2 studies (column 12), Log<sub>10</sub>BF (column 13) and corresponding posterior probabilities (column 14) from trans-ethnic meta-analysis of all stage 1 and stage 2 studies. A posterior probability > 0.5 suggests heterogeneity in allelic effects.

**Table 2**

Potential function of loci associated with coffee consumption<sup>a</sup>

Locus	Gene expression response to caffeine <sup>b</sup>	Lead-SNP, allele ↑ coffee consumption <sup>c</sup>	Non-Syn SNPs in LD <sup>d</sup>	CR <sup>e</sup>	DNAse <sup>f</sup>	Proteins Bound <sup>g</sup>	Histone Marks <sup>h</sup>	Motifs changed <sup>i</sup>	eQTL <sup>j</sup>	mQTL <sup>k</sup>
2p24	GCCR, CCDC121, FNDC4, ZNF513, SNX17, PPM1G, GPN1, SUP17L, MPV17, SLC4A1AP, PREB, ATRAID, GTF3C2	rs1260326, C	Leu446Pro	✓	✓	✓	enhancer	NRSF	EIF2B4, SNX17, NRBP1	KRTCAP3, PPM1G
4q22	ABC G2, SPP1	rs1481012, A	✓	✓	✓	✓	enhancer	AIRE, Zfp105		
7p21	AHR	rs4410790, C rs6968554, G		✓	✓			Cdx2, DMRT3, E4BP4, Foxa, GR, Hoxa10, Hoxa9, Hoxb13, Hoxb9, Hoxc9, Hoxd10, Myc, p300, TR4		AHR
7q11.23	MLXIPL, BCL7B, DNAJC30, TBL2, WBSR22	rs7800944, C			✓	✓	promoter enhancer	AP-4, BHLHE40, GATA, GR, Inf, Pax-5	WBSR22, MLXIPL	FZD9
7q11.23	RHBDD2, POR, STYXLI, TMEM120A, MDH2, HSPB1	rs17685, A	✓	✓	✓	✓		Amt, BHLHE40, DEC, Eis, Mxi1, Myc, Pax-5, Sin3Ak-20, TFE	RHBDD2, POR, TMEM120A, STYXLI, MDH2	STYXLI
11p13	CCDC34, LIN7C, METTL15,	rs6265, C	Val66Met	✓	✓	✓	promoter enhancer	BHLHE40, Myc, SREBP		
15q24	PPCDC, ARID3B, ULK3, SEMA7A, EDC3, COX5A, CSK, RPP25, MPI	rs2470893, T rs2472297, T						SP1	MPI, SCAMP2, ULK3, ISLR, SNUPN, RPP25, CSK,	SCAMP2
17q11.2	TAOK1, SLC6A4, NSRP1, BLMH	rs9902453, G	✓	✓	✓	✓	promoter enhancer	STAT	GIT1, ATAD5, SLC6A4	NSRP1, ANKRD13B, CRLF3, CORO6

Abbreviations: CR, conserved region; eQTL, expression quantitative trait loci; mQTL, methylation quantitative trait loci

<sup>a</sup> See Supplementary Information for details and references to data resources.

<sup>b</sup> In vitro human hepatic gene expression in response to caffeine. Red and green font corresponds to increased and decreased expression, respectively.

<sup>c</sup> Lead SNP-allele associated with higher coffee consumption. Check marks (✓) denote presence of

<sup>d</sup> non-synonymous SNPs in LD (CEU; r<sup>2</sup> > 0.80) with lead SNP (details provided for lead SNP only).



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<sup>e</sup> a conserved region (spanning lead SNP and its correlated proxies, CEU:  $r^2$  0.8).

<sup>f</sup> DNase hypersensitivity sites at region spanning lead SNP and its correlated proxies, CEU:  $r^2$  0.8.

<sup>g</sup> proteins bound at region spanning lead SNP and its correlated proxies, CEU:  $r^2$  0.8.

<sup>h</sup> Enhancer (H3K4me1) or promoter (H3K4me3) histone marks (as defined by Ernst et al<sup>32</sup>) spanning lead SNP and its correlated proxies, CEU:  $r^2$  0.8.

<sup>i</sup> Regulatory motifs altered by lead SNP.

<sup>j</sup> Expression QTLs for lead SNP or perfect proxy (CEU:  $r^2=1$ ) derived from lymphoblastoid cell lines, blood, or liver, adipose and brain tissues. Red and green font corresponds to increased and decreased expression, respectively, relative to allele associated with *higher* coffee consumption. Direction of *GIT1* expression is not available.

<sup>k</sup> Methylation QTLs for lead SNP derived from cerebellum and frontal cortex. Red and green font corresponds to increased and decreased expression, respectively, relative to allele associated with *higher* coffee consumption.

**Table 3**

Associations between coffee consumption loci and other traits

Lead SNP, allele ↑ coffee consumption <sup>a</sup> closest gene	Other Traits <sup>b</sup>	
	higher levels/risk <sup>c</sup>	lower levels/risk <sup>c</sup>
rs1260326, C <i>GCKR</i>	non-albumin protein fasting glucose HOMA-IR fasting insulin mannose	serum albumin 2-hr glucose challenge metabolic syndrome glucose/mannose ratio total cholesterol triglycerides hypertriglyceridemia chronic kidney disease uric acid SHBG Crohn's disease C-reactive protein platelet counts GGT docosapentaenoic acid alanine/glutamine ratio alanine
		LDL ( $P=2.33 \times 10^{-4}$ ) waist-to-hip-ratio ( $P=3.40 \times 10^{-4}$ ) bipolar disorder ( $P=2.31 \times 10^{-3}$ )
rs1481012, A <i>ABCG2</i>		LDL response to statins ('responders') uric acid
	body mass index ( $P=4.85 \times 10^{-3}$ )	
rs6968554, G <i>AHR</i>		caffeine
		HDL ( $P=1.18 \times 10^{-3}$ )
rs7800944, C <i>MLXIPL</i>		triglycerides
	HDL ( $P=2.24 \times 10^{-3}$ ) birth weight ( $P=2.10 \times 10^{-3}$ )	
rs6265, C <i>BDNF</i>	smoking initiation body mass index	
		DBP ( $P=6.58 \times 10^{-4}$ )
rs2472297 <sup>d</sup> , T <i>CYP11A1_CYP11A2</i>		caffeine <sup>e</sup>
		SBP ( $P=6.81 \times 10^{-5}$ ) DBP ( $P=6.75 \times 10^{-6}$ )
rs9902453, G <i>EFCAB5</i>		SBP ( $P=6.05 \times 10^{-3}$ )

Abbreviations: DBP, diastolic blood pressure; GGT, gamma-glutamyltransferase; HDL, high density lipoprotein; LDL, low density lipoprotein; SBP, systolic blood pressure; SHBG, sex hormone binding globulin

<sup>a</sup>Lead SNP-allele associated with higher coffee consumption.

<sup>b</sup>Traits associated with lead SNP (or close proxies:  $r^2 > 0.80$ ) according to previous GWAS<sup>20</sup>(Shin et al, 2014). Grey cells denote all GW-significant significant associations ( $P < 5.00 \times 10^{-8}$  20 or  $P < 1.03 \times 10^{-10}$  (Shin et al, 2014) and white cells denote coffee-relevant trait associations ( $P < 6.25 \times 10^{-3}$ ). See Supplementary Information for details and references to original GWAS.

<sup>c</sup>Relative to allele associated with higher coffee consumption.

<sup>d</sup>rs1378942 A, also associated with higher coffee consumption ( $P < 1.46 \times 10^{-17}$ ) in stage 1 of the current report but in low LD with rs2472297 (CEU:  $r^2=0.10$ ), was previously associated with lower DBP in GWAS ( $P < 5.00 \times 10^{-8}$ ).

<sup>e</sup>Borderline significant ( $P < 1.51 \times 10^{-10}$ ) according to Shin *et al*<sup>21</sup>.

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