# Clinical significance of asymptomatic cytomegalovirus viremia in lung transplant recipients receiving universal antiviral prophylaxis

Student: Nikta Jaamei

Tutor: PD Dr Oriol Manuel Prof Manuel Pascual

Expert: Prof John-David Aubert

December 2015

UNIL | Université de Lausanne



Centre hospitalier universitaire vaudois

#### ABSTRACT

**Background:** Cytomegalovirus (CMV) disease has been associated with the development of chronic lung allograft dysfunction (CLAD) after lung transplantation. However, the relevance of asymptomatic CMV viremia occurring after the discontinuation of antiviral prophylaxis (late-onset CMV replication) on the development of CLAD is not fully understood. We aimed to assess the long-term clinical impact of asymptomatic CMV replication in a cohort of lung transplant recipients receiving universal antiviral prophylaxis.

**Methods:** We performed a single-center study including all patients who underwent lung transplantation between 2004 and 2014. Patients received valganciclovir prophylaxis for 3 to 6 months (according to CMV serostatus risk of the donor [D] and recipient [R]), followed by monitoring of CMV replication by PCR during the first year post transplant. CLAD was defined according to ISHLT definitions. Risk factors for the development of CLAD and for mortality were assessed by univariate and multivariate Cox models. A lineal regression model was used to evaluate the influence of CMV replication in the evolution of FEV1.

**Results:** Overall, 69 patients were included. CMV serostatus was D-/R- in 13 (19%) patients, D-/R+ in 17 (25%) patients, D+/R+ in 27 (39%) patients and D+/R- in 12 (17%) patients. Overall, 34/69 (49%) patients developed at least one episode of asymptomatic CMV replication and 8/69 (11.5%) patients developed CMV disease. Median duration of CMV replication in viremic patients was 57.5 days. After a median follow-up of 3.69 years, 25/69 (36%) patients developed CLAD and 14/69 (20%) patients died. In the univariate cox analysis, bacterial pneumonia was associated with a higher incidence of CLAD (HR 2.58, p=0.06), but asymptomatic CMV replication (HR 1.36, p=0.45), CMV disease (HR 1.00, p=0.99), and duration of CMV replication (HR 1.00, p=0.76) were not. In the multivariate model, bacterial pneumonia remained associated with CLAD (HR 2.53, p=0.06). In the mixed model of linear regression, we did not observe a correlation between CMV replication and a significant decline of FEV1 (estimate -0.162, CI 95% [-0.498 to 0.170], p=0.35). CMV replication was not associated with a higher mortality (HR 0.75, p=0.62).

**Conclusion:** In this cohort of lung transplant recipients receiving antiviral universal prophylaxis, asymptomatic CMV replication did not influence long-term allograft lung function and patient survival. These results suggest that the use of universal prophylaxis is protective against the indirect effects of CMV, irrespective of the development of late-onset CMV replication.

Key words: viral infection, chronic lung allograft dysfunction, lung transplantation, outcomes

#### **1. INTRODUCTION**

#### **1.1 Lung transplantation**

The first lung transplantation ever was performed in 1963 and the patient survived only 18 days. Much has changed over these last decades, as lung transplantation is today the preferred treatment for many end-stage lung diseases, such as chronic obstructive pulmonary disease (COPD), interstitial lung diseases and cystic fibrosis. Worldwide, the amount of lung transplantation procedures achieved has dramatically increased since 1990. In 2014, 56 lung transplantations were performed in Switzerland, which of 24 were done at the Lausanne University Hospital (CHUV) (1). Median waiting time for lung transplantation was 465 days, with a stable waiting list since 2010 (Figure 1). In 2014, 8 patients died waiting for a lung.

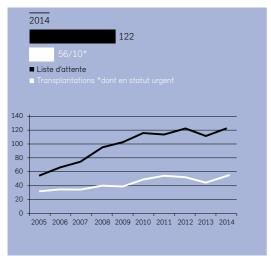


Figure 1: Lung transplantation in Switzerland and waiting list, year 2014 (Swisstransplant annual report 2014)

## **1.2 Outcome after lung transplantation**

The survival after lung transplantation has been increasing worldwide since 1990. In the era between 1990 and 1997, the median survival post-transplantation was 4.3 years. It has now reached 6.3 years for the 2005-2012 era (2). The median survival for single and double-lung transplantation according to the data from the ISHLT registry is shown in figure 2. However, survival after transplant depends on many different factors. Better survival is correlated with younger and female recipients, as well as the underlying lung disease of the recipient. When considering adjusted survival (patients surviving at least one year), median survival was the longest for CF patients (11 years) followed by idiopathic pulmonary arterial hypertension (IPAH), sarcoidosis and COPD associated with  $\alpha$ 1-antitrypsin deficiency. Transplant

recipients receiving a double-lung transplantation have a better survival (7 years vs 4.5 years for single transplant) (2).

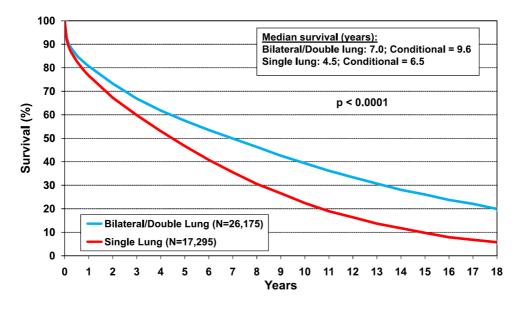


Figure 2: Median survival for single and double lung transplant recipients (ISHLT registry)

## 1.2.1 Causes of death

The most common causes of death within the first 30 days post-transplantation are graft failure and technical or cardiovascular causes. Infections are a major cause of death at any time. After the first year, another main cause of death is chronic lung allograft dysfunction (CLAD), which includes processes like bronchiolitis obliterans syndrome (BOS) that will be discussed later. Main causes of death are listed in table 1.

| First 30 days              | - Graft failure                      |
|----------------------------|--------------------------------------|
|                            | - Infectious complications           |
|                            | - Surgical complications             |
|                            | - Cardiovascular causes              |
| First year                 | - Infectious complications           |
| After 1 <sup>st</sup> year | - Chronic lung allograft dysfunction |
|                            | - Infectious complications           |

Table 1: main causes of death following lung transplantation (ISHLT registry)

# 1.2.2 Risk factors for mortality

Patients with BOS or acute rejection in the first-year post-transplant have a higher risk of mortality at 5 years (2). In a study involving 23'704 lung transplant recipients, the mortality risk model at 5 years demonstrated a negative impact from advanced recipient age and lower transplant center volume. The impact of age seems to be restricted to the recipient, as age appears not to be relevant when considering donor age. In fact, it was observed that allograft from older donors (>65 years old) did not affect the survival, although there seems to be a negative effect when given to a younger recipient (3). With the aging of the population, this question must be further studied to assess the impact of transplantation including older (>60 years old) donors or recipients. Other risk factors that may influence patient survival are listed in table 2.

| 5-year post-transplant | - Underlying lung disease of recipient                   |
|------------------------|--|
|                        | - Retransplantation                                      |
|                        | - Earlier era of transplant                              |
|                        | - Increased severity of recipient illness at the time of |
|                        | transplantation (intensive care unit)                    |
|                        | - Donor history of diabetes                              |
|                        | - CMV mismatch (D+/R-)                                   |
|                        | - Lower transplant center volume                         |
|                        | - Older recipient at age of transplant                   |
|                        | - Higher pretransplant supplemental oxygen required      |
|                        | - Lower cardiac output                                   |

Table 2: Risk factors of death following lung transplantation (ISHLT registry)

## 1.2.3 Quality of life

Lung transplantation has a positive impact on the quality of life (QoL) (4). There is an improvement in many dimensions of health - as defined by the Nottingham health profile - except for the aspects of pain and mental health. However there is a clear improvement in dimensions such as social life or physical mobility.

## 1.3 Immunosuppressive regimen

## 1.3.1 Induction therapy

At the time of transplantation, induction therapy is used to prevent the immune system of the recipient, especially T-cells, to attack the allograft. Induction agents are either monoclonal agents (i.e basiliximab, daclizumab) or polyclonal agents (anti-lymphocytes, anti-thymocytes). There is some conflict in the literature about the efficacy or superiority of one agent for induction therapy. There may be some benefit using monoclonal agents to prevent from onset of chronic rejection (5), but other studies do not confirm this hypothesis (6). Of note, 50% of centers do not generally use induction therapy, and this without a clear negative impact on allograft and patient survival.

# 1.3.2 Maintenance therapy

The aim of maintenance therapy is to keep a balance between over-immunosuppression that will lead to infection and under-immunosuppression that will cause allograft rejection. Different agents are used low-dose, to minimize the potential toxic adverse effects. Usual treatment combines three agents:

- 1. Glucocorticoids, i.e prednisone: always used for immunosuppression. It inhibits both the adaptative and innate immune system and down-regulates the transcription of inflammatory genes (7)
- Calcineurin inhibitors: main agents are cyclosporine and tacrolimus. Both agents are inhibiting T-cell activity, but tacrolimus is now more widely used, especially among lung transplant recipients (8). Tacrolimus has less nephrotoxic adverse effects and may reduce the onset of BOS (9), but is more diabetogenic.
- Nucleotide blocking agents: usually azathioprine (AZA) or mycophenolate mofetil (MMF). While some studies have shown superiority of MMF over AZA treatment (10), this has not been confirmed in a randomized clinical trial (11).

# 1.4 Allograft rejection and dysfunction

## 1.4.1 Acute rejection

They are three types of acute rejection:

- 1. Cellular rejection: mediated by T-cells directed towards HLA antigens. It is the predominant type of acute rejection
- 2. Humoral rejection: mediated by antibodies directed towards donor HLA

3. Hyperacute rejection: occurs within first 24 hours post-transplantation and is mediated by pre-formed antibodies from the recipient

Acute rejection occurs in approximately one-third of lung transplant recipients (12). Acute rejection should be confirmed by a histological diagnosis, as an infectious process needs to be excluded. The difference between a rejection or an infectious process cannot be always done on clinical findings, as the presentation may be similar in some instances (13). Symptoms may occur mostly from A2 grade rejection and more.

The actual nomenclature of allograft rejection has been established by the ISHLT in 2007 (14).

- A. Acute rejection: the diagnosis of acute rejection is made only on the presence of perivascular and interstitial mononuclear cell infiltrate. The grade can be between 0 (no acute rejection) to grade 4 (severe) and reflects the most advanced pattern (and not the predominant one).
- B. Airway inflammation: lymphocytic bronchiolitis. This designation applies to small airways (bronchioles) and the grading is B0 (absent), B1 (low grade), B2 (high grade) or Bx (ungradable).
- C. Chronic airway rejection obliterative bronchiolitis. This definition occurs when eosinophilic hyaline fibrosis in the sub-mucosa of membranous and respiratory bronchioles is seen, resulting in a partial or complete luminal occlusion. They may be associated with destruction of the smooth muscle and elastica of the airway. Mucostasis or foamy histiocytes are commonly associated with this phenomenon even in the absence of bronchiolar occlusion.
- D. Chronic vascular rejection accelerated graft vascular sclerosis. This process is similar to coronary artery disease in the heart. It describes a fibrointimal thickening of arteries and veins.

#### 1.4.2 Chronic lung allograft dysfunction (CLAD)

Surgical techniques and immunosuppressive regimens have greatly improved since 1963, however lung transplantation is still the solid organ transplantation (SOT) with least success, since 5 years survival is only 53% (15). The major factors that explain this disparity compared to other SOT is the highest incidence of chronic allograft rejection in lung transplant recipients, which is now defined as chronic lung allograft dysfunction (CLAD) (16).

CLAD is not an etiological definition, but indicates that the lung is not achieving its full function. It is not irreversible per se, although it is usually a permanent condition. Depending on the spirometrical parameters, CLAD may present as a restrictive or an obstructive pattern.

- Bronchiolitis Obliterans Syndrome (BOS): decreased FEV1 (FEV1) <80% from baseline for > 3 weeks without any confounding condition and evidence of airflow obstruction (17). FEV1 baseline is defined as "the average of the two highest values for each measurement that were obtained at least three weeks apart post-transplant without the administration of a bronchodilatator" (17).
- *Restrictive Allograft Syndrome (RAS)*: FVC or TLC <80% baseline FVC or TLC for > 3 weeks.

We will discuss the pathogenesis and management of BOS in the next section.

## 1.5 Bronchiolitis Obliterans Syndrome

BOS is the clinical correlate of obliterative bronchiolitis (OB). OB is a histological diagnosis presenting as progressive obliteration of small airways. Because it was difficult to make this diagnosis via transbronchial lung biopsy, the actual consensus is to use BOS as the clinical marker of chronic allograft dysfunction. BOS is a major issue in lung transplantation as it represent the main cause of death adjusted for conditional survival of one year (18). The incidence reaches 50% within 5 years after transplantation (19) and the median survival after diagnosis is 3 years.

#### 1.5.1 Pathogenesis of BOS

The first step of BOS is onset of lymphocytic infiltrates in the submucosa, with processes like lymphocytic bronchiolitis (LB) leading to epithelial injury. This injury will thus create the following sequence: necrosis  $\rightarrow$  ulceration  $\rightarrow$  inflammation that will end up by fibroblastic proliferation. Finally, the intraluminal polypoid granulation will eventually occlude totally or subtotally airways lumen (20). This is a complex process including immune and non-immune mechanisms. The molecular pathways are not fully understood yet, but we will shortly summarize the role of some cytokines in figure 3.

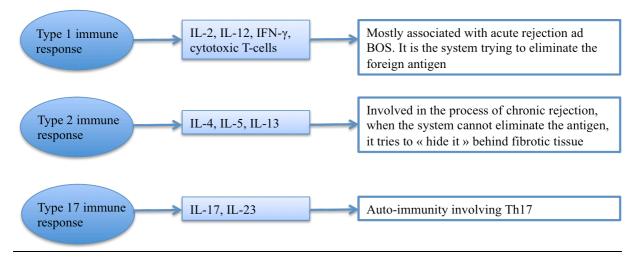


Figure 3: cytokines involved in BOS pathogenesis (Adapted from Weigt and al, 2010)

## 1.5.2 Risk factors of BOS

Here are listed some risk factors for onset of BOS.

- Alloimune mechanisms: BOS may be the clinical consequence of repeated acute rejections episodes (21). There is an increased risk of BOS when there are more HLA mismatches (22). Acute rejection may present with peri-airway infiltration of activated lymphocytes, which is described as lymphocytic bronchiolitis (LB). LB is recognized as the most significant risk factor for developing BOS (23).
- Primary graft dysfunction (PGD): occurs typically within the first 24 hours after transplantation and is caused by events during the transplantation procedure, such as pulmonary ischemia or donor tissue preservation (20). There is evidence that not only it does affect the survival on a short term, but influences also the long-term survival and is hence considered a BOS risk factor.
- Respiratory infections: community-acquired respiratory virus (CARV) have been identified as risk factor for BOS (24), and this risk was tripled when the lower respiratory tract was implicated. In smaller studies, *Chlamydia pneumophilae* and HHV-6 were also recognized as risk factors. The airway colonization by *Pseudomonas aeruginosa* and *Aspergillus* may be an independent risk factor for BOS as well (25)
- Others: gastro-intestinal reflux, air pollution.

CMV role will be discussed later in a separate section

#### 1.5.3 Prevention and detection of BOS

BOS is a clinical diagnosis mainly based on FEV1 evolution from baseline. However it needs some expertise by the transplant physician. There is an overall good agreement regarding the presence or absence of BOS diagnosis (26). There may be some variability when performing the spirometry, due to patient's performance. One issue with BOS diagnosis is that it is retrospective by definition therefore the impact on the clinical management is limited. However there is some literature about how to predict or prevent onset of BOS. In a retrospective study including 121 patients, the measurement of CRP and neutrophilia in the broncho-alveolar lavage (BAL) at 90 days after transplantation was a predictive factor for determining the outcome of the allograft (27). Patients with BOS had a higher BAL CRP (cutoff >173 pg/ml) than the BOS-free group (BAL CRP baseline: 50 in stable group and 540,4 in BOS group, p=0.002). CRP stimulates alveolar macrophages and alters surfactant function in the lungs which indicates a role during acute respiratory distress syndrome and could be used as a marker of lung injury (28). The use of azithromycine, a macrolide antibiotic with immunomodulatory effect, as prophylaxis (29) or treatment (30) is useful by inhibiting the neutrophilic airways inflammation pathway. When used as prophylaxis, it increases the BOS-free interval of 2.3 years and the pulmonary functions were significantly better (p=0.051). As a treatment, azithromycine given during 12 weeks increases the FEV1 significantly (FEV1 difference between azithromycine and placebo group was 0.035 l and 39% of patients in azithromycine group had >10% gain in FEV1 compared to 0% in the placebo group).

#### 1.6. Infections in a lung transplant recipients

Infections are one of the most common causes of death after transplantation (2), usually presenting as pneumonia (31). The pathogens are mostly bacteria, with gram-negative rods such as *Pseudomonas* and *Burkholderia* being the most frequent causes of pneumonia in lung transplant recipients (31).

#### 1.7. Human cytomegalovirus

Cytomegalovirus (CMV) is a double-stranded DNA virus in the family of Human herpes virus (HHV) and is also called HHV-5. CMV is transmitted by contact, blood transfusion or organ transplantation. In the immunocompetent host, the primary infection is usually

asymptomatic and CMV remains latent. However the virus may reactivate in case of stress or immunocompromise.

Cytomegalovirus disease is the most important viral infection developing after SOT (32). CMV disease usually presents as a syndrome with fever, fatigue and body ache. Regardless of the type of transplant, CMV tissue-invasive disease usually involves the gastro-intestinal tract, with complications such as hemorrhage or perforation. It can however also affect lungs, kidney, liver or any other organ (33). These effects are called the "direct effects" of CMV. Regarding lung transplantation, the main concern is about CMV pneumonitis, that causes alterations of the allograft.

Moreover, CMV infection leads to many "indirect effects" - which cannot directly be related to a viral invasion of tissue - like an increased rate of bacterial and fungal infections or acute rejection (34). A relationship between kidney transplant, late-onset CMV infection and the concurrent post-transplant diabetes mellitus has been observed as well (35), suggesting a broad spectrum of indirect CMV effects. Direct and indirect effects are summarized in table 3.

| Direct |                    | Indirect |   |  |
|--------|--------------------|----------|---|--|
| -      | CMV syndrome       | -        | ↑ Bacterial, fungal infections                            |  |
| -      | Tissue-invasive    | -        | Acute and chronic rejection                               |  |
|        | disease (mostly GI | -        | Diabetes  |  |
|        | tract)             | -        | More aggressive HCV infection after liver transplantation |  |
|        |                    | -        | Post transplant lymphoproliferative disorder              |  |
|        |                    | -        | Death   |  |

Table 3: Direct and indirect effects of CMV

## 1.7.1. Risk factors for CMV infection

The incidence of CMV infection depends on different factors. The most important one is the serostatus at transplantation. Naïve recipients receiving organs from seropositive donors (D+/R-) are considered at high-risk. Patients with either D-/R+ or D+/R+ seroconstellation are considered intermediate risk and finally D-/R- low risk (32). The state of immunosuppression also influences the risk of CMV disease (36). The risk also depends on the organ transplanted: lung, small intestine and pancreas have the highest risk of CMV, whereas liver and kidney have the lowest (36). Risk factors are summarized in table 4.

#### **Risk factors for CMV disease**

- Serostatus
- State of immunosuppression
- Lung, small intestine or pancreas transplantation

Table 4: Risk factors for CMV disease

# 1.7.2. Immunomodulation and CMV

To be able to maintain latency in the immunocompetent individual, CMV had to find ways to hide from the host immune system. For example, it interferes in the antigen presentation mechanism by producing HLA class I homologue that will block recognition by natural killers and other T cells (37) and also downregulates expression of antigen-presenting cells (APCs) (38). CMV also modulate the humoral immune mechanisms by producing its own FC receptor homolog (39). These receptors will bind circulating IgG and thus allow the infected cell to escape other immunoglobulin directed towards viral proteins. By altering cellular and humoral immune pathways, CMV inhibits the immune system in a non-specific manner, which would be likely to increase infection susceptibility in the host.

# 1.7.3. CMV and allograft rejection

There is a question whether CMV reactivation would increase the risk of allograft rejection or the allograft rejection would lead to CMV reactivation. The answer is that the mechanism is bidirectional, as summarized in Figure 8. Allograft rejection produces a pro-inflammatory cytokine (TNF) that will induce the reactivation of CMV (40). When CMV reactivates, it stimulates infected macrophages and AECs to produce molecules molecules that will increase the number of inflammatory cells in the lung. These molecules are endothelial adhesion molecules such as VCAM, ICAM, LFA-1 and VLA-4. It does also activates MHC class I antigen by molecular mimicry (41). Figure 4 shows this bidirectional interaction. There are some genetic factors affecting the susceptibility to CMV recurrence. Polymorphism in recipient's genes coding for IFN- $\lambda$ 3 and IFN- $\lambda$ 4 influence the susceptibility to CMV replication in SOT recipients (42). This correlates with previous studies finding that these polymorphisms were associated with a reduced hepatitis C virus clearance. There is also an association between SNP polymorphisms of IFN- $\gamma$  and CMV disease and high CMV viremia (43).

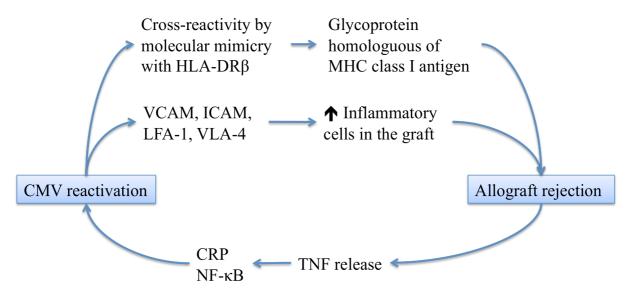


Figure 4: Bidirectional interaction between CMV reactivation and allograft rejection (Adapted from Freeman and al, 2009)

#### 1.7.4. Management and prevention of CMV replication/disease in lung transplant recipients

Strategies have been set to minimize the impact of CMV replication/disease on lung transplant recipients. The actual guidelines recommend a prophylactic approach over a preemptive one for high-risk (D+/R-) group with a 6 months therapy (44). Intermediate risk group (D-/R+, D+/R+) should be receiving 3 months of prophylaxis and there is no indication for prophylactic treatment regarding low-risk group (D-/R-). The use of universal prophylaxis with valganciclovir or ganciclovir has decreased overall the incidence of CMV infection and acute rejection in the current era (45).

#### 1.7.5. CMV infection and BOS

CMV pneumonitis is a well-known risk factor for developing a BOS. With the development of antiviral prophylaxis for high-risk (D+/R-) group, the incidence has consequently diminished. However, CMV replication remains associated with graft rejection and graft loss (46) and D+/R- recipients have still a higher mortality than the other groups despite the prophylaxis, which means they should be followed more carefully and proactively (47). Nowadays, we have more technical tools that allow us to identify CMV DNAemia in the blood and it has been therefore noticed that there may be a subclinical CMV replication in the bloodstream. There is still a debate about the clinical relevance of these subclinical DNAemia. Some studies showed an correlation between CMV DNAemia episodes and onset of BOS (48), which was not confirmed by other studies (49).

## **1.8. Filling a literature gap**

The impact of CMV disease on survival and allograft function is nowadays well studied. CMV disease, especially CMV pneumonitis, occurring at an early or late stage is known as a risk for graft dysfunction and mortality (50). But the impact of asymptomatic CMV replication is not well defined yet and how to manage it in clinical practice. The aim of this study is to examine the potential clinical consequences of asymptomatic CMV replication in blood on allograft outcomes in a cohort of lung transplant recipients.

#### 2. MATERIAL AND METHODS

#### 2.1 Study design and patient population

We performed a single-center retrospective cohort study including all patients who underwent lung transplantation between 2004 and 2014 in the University Hospital of Lausanne (CHUV). Patients that were transplanted in Lausanne but had a follow-up in another transplant center (Basel, Bern of Geneva) were not included in the analysis.

We reviewed patient's records to collect demographic parameters, immunosuppressive regimens, FEV1 values, as well as infectious complications and episodes of acute rejection. The "Comission d'éthique de la recherche du canton de Vaud" approved the study protocol.

#### 2.2 Definitions in the study

CMV infection was defined according to standard guidelines (44) by the evidence of CMV replication regardless of symptoms. CMV disease was the evidence of CMV infection with attribuable symptoms. CMV disease was further categorized as viral syndrome (fever, fatigue, myalgia) or end-organ disease (pneumonitis, colitis, gastro-enteritis or other). CLAD, BOS and RAS were classified according to ISHLT definitions. The term BOS is used when a persistent decline of FEV1 with no potential reversible cause found (30). Acute rejection was also defined following ISHLT definitions (14) which means the diagnosis of acute rejection is made only on the presence of perivascular and interstitial mononuclear cell infiltrate.

#### 2.3 Immunosuppressive regimen and antiviral prophylaxis

The induction treatment was done with basiliximab. Maintenance therapy regimen used was tacrolimus, prednisone and mycophenolate mofetil (MMF). Patients at an intermediate risk for CMV serostatus (D+/R+ and D-/R+) received valganciclovir prophylaxis during 3 months and patients at high-risk (D+/R-) during 6 months. Patients with D-/R- serostatus received no anti-CMV prophylaxis, but anti-herpes prophylaxis with valaciclovir for 3 months. After discontinuation of prophylaxis, patients were monitored for CMV replication by PCR every 2 weeks. The detection of CMV viral load in blood was done by PCR according to standard method (51).

Lung function tests, including FEV1, FVC and 6 minutes walking test were measured at 3, 6, 12 months post-transplant and then each year and they were used to calculate the development of BOS and RAS.

## 2.4 Statistical analysis

We aimed to assess the long-term clinical impact of asymptomatic CMV replication in a cohort of lung transplant recipients receiving universal antiviral prophylaxis.

Descriptive statistics were used to describe the demographics and clinical characteristics of the population. Risk factors for the development of CLAD and for mortality were assessed by univariate and multivariate Cox models, including variables such as CMV replication, CMV diseasem CMV serostatus, other infectious complications (bacterial pneumonia, invasive fungal infection) and acute rejection. A lineal regression model was used to evaluate the influence of CMV replication in the evolution of FEV1.

# **3. RESULTS**

## **3.1.** Characteristics of the study population

Overall, 69 patients were included in the study. The mean age was 52 years old and 61% of patients were female. Most patients received double-lung transplantation. The main indications for transplantation were COPD (40%) and cystic fibrosis (26%). The median duration of follow-up was 3.67 years. Overall, 17% of patients had a high-risk CMV serostatus (D+/R-). 81% of patients received antiviral prophylaxis with valganciclovir for a mean duration of 3.29 months and 14% received valaciclovir. Baseline characteristics and main outcomes are listed in tables 5 and 6.

| Demographic variables             | All recipients |
|-----------------------------------|----------------|
| Number of patients                | 69             |
| Time of follow-up in days, median | 1341 (34-3806) |
| (range)                           |                |
| Age, years mean (SD)              | 52.25 (14)     |
| Male sex (%)                      | 27 (39%)       |
| Type of lung transplant (%)       |                |
| - single                          | 7 (11%)        |
| - double                          | 62 (89%)       |
| Underlying lung disease (%)       |                |
| - COPD                            | 28 (41%)       |
| - Cystic fibrosis                 | 18 (26%)       |
| - Other (IPF, alpha1AT            | 23 (33%)       |
| deficiency, sarcoidosis)          |                |

| Immunosuppressive regimen    |            |  |  |  |
|------------------------------|------------|--|--|--|
| - Tacrolimus                 | 68 (98%)   |  |  |  |
| - MMF                        | 64 (93%)   |  |  |  |
| - Steroids                   | 69 (100%)  |  |  |  |
| CMV serostatus (%)           |            |  |  |  |
| - D-/R-                      | 13 (29%)   |  |  |  |
| - D-/R+                      | 17 (25%)   |  |  |  |
| - D+/R+                      | 27 (39%)   |  |  |  |
| - D+/R-                      | 12 (17%)   |  |  |  |
| Antiviral prophylaxis (%)    |            |  |  |  |
| - valganciclovir             | 56 (81%)   |  |  |  |
| - valaciclovir               | 10 (14%)   |  |  |  |
| Median duration of antiviral | 91 (0-1464 |  |  |  |
| prophylaxis in days (range)  |            |  |  |  |

Table 5: Baseline characteristics of the study population

| Outcomes                              | All recipients |
|---------------------------------------|----------------|
| CMV asymptomatic replication n (%)    | 34 (49%)       |
| Duration of CMV replication, days,    | 57.5 (1-506)   |
| median (range)                        |                |
| CMV disease, n (%)                    | 8 (11.5%)      |
| Pneumonia episodes (%)                | 41 (59%)       |
| Acute rejections (A1-A3) episodes (%) | 36 (52%)       |
| CLAD (%)                              | 25 (36%)       |
| CLAD free interval, median (range)    | 957 (237-2182) |
| Death (%)                             | 14 (20.3%)     |

Table 6: Main outcomes of patients included in the study

## **3.2.** CMV infection and disease

Overall, 34/69 (49%) patients developed at least one episode of asymptomatic CMV replication and 8/69 (11.5%) developed CMV disease (3 patients presented with colitis, 2 with pneumopathy, 1 with gastro-enteritis and 2 with other presentation). Median duration of CMV replication in viremic patients was 57.5 days.

## 3.3. Risk factors for the development of chronic lung allograft dysfunction

After a median follow-up of 3.67 years, 25/69 (36%) patients developed CLAD. Of the 25 cases of CLAD, 22 were considered to be BOS and 3 were considered to be RAS.

Table 7 shows the univariate Cox analysis for risk factors associated with CLAD. Asymptomatic CMV replication (HR 1.36, p=0.45) and CMV disease (HR 1.00, p=0.76) were not associated with a higher risk for the development of CLAD. We observed a lower proportion of CLAD in the D-/R+ group (HR 0.289, p=0.123), but this was not statistical significant. The duration of valganciclovir prophylaxis did not have an influence on the development of CLAD. Bacterial pneumonia was associated with a higher incidence of CLAD (HR 2.58, p=0.06). Kaplan-Meier curves of CLAD-free survival according to CMV replication, CMV serostatus, acute rejection and bacterial pneumonia are shown in Figure 5.

| Variable                    | Hazard<br>ratio | IC 95% |       | P-value |
|-----------------------------|-----------------|--------|-------|---------|
| Sex (male)                  | 0.775           | 0.345  | 1.741 | 0.538   |
| Age at Tx                   | 0.998           | 0.970  | 1.027 | 0.897   |
| CMV replication             | 1.368           | 0.601  | 3.118 | 0.455   |
| Duration of CMV replication | 1.004           | 0.979  | 1.029 | 0.758   |
| CMV disease                 | 1.001           | 0.297  | 3.374 | 0.998   |
| Duration of                 | 0.895           | 0.737  | 1.086 | 0.259   |
| valganciclovir              |                 |        |       |         |
| CMV serostatus              |                 |        |       |         |
| D-/R- (ref.)                | 1               |        |       |         |
| D-/R+                       | 0.289           | 0.060  | 1.402 | 0.123   |
| D+/R-                       | 0.556           | 0.176  | 1.756 | 0.317   |
| D+/R+                       | 0.818           | 0.313  | 2.137 | 0.681   |
| Acute rejection             | 1.561           | 0.668  | 3.645 | 0.304   |
| Bacterial pneumonia         | 2.583           | 0.959  | 6.958 | 0.060   |
| Fungal infection            | 2.020           | 0.756  | 5.396 | 0.161   |

Table 7: Univariate Cox regression model of risk factor for the development of CLAD

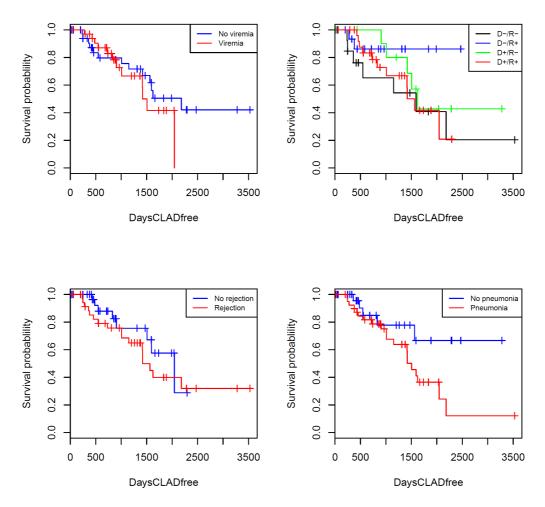


Figure 5: Kaplan-Meier curves on CLAD-free survival according to CMV replication (above, left), CMV serostatus (above, right), acute rejection (below, left), and bacterial pneumonia (below, right)

In the multivariate model (table 8), bacterial pneumonia remained associated with CLAD (HR 2.53, p=0.06) but CMV replication did not (HR 1.16, p=0.71).

| Variable            | Hazard<br>ratio | IC 95% |       | P-value |
|---------------------|-----------------|--------|-------|---------|
| CMV replication     | 1.162           | 0.511  | 2.639 | 0.719   |
| Bacterial pneumonia | 2.531           | 0.934  | 6.862 | 0.067   |

Table 8: Multivariate Cox regression model of risk factors for the development of CLAD

In the mixed model of linear regression (Table 9), we did not observe a correlation between CMV replication and a significant decline of FEV1 (estimate -0,162, CI 95% [-0.498 to 0.170], p=0.35).

| Coefficient          | Estimate | CI 95% |       | p-value |
|----------------------|----------|--------|-------|---------|
| (Intercept)          | 2.370    | 2.112  | 2.629 | 0.000   |
| viremia              | -0.162   | -0.498 | 0.175 | 0.346   |
| period               | 0.052    | -0.134 | 0.239 | 0.583   |
| I(period^2)          | -0.021   | -0.075 | 0.033 | 0.446   |
| I(period^3)          | 0.001    | -0.004 | 0.005 | 0.808   |
| viremia :period      | 0.216    | -0.169 | 0.601 | 0.271   |
| viremia :I(period^2) | -0.080   | -0.207 | 0.047 | 0.214   |
| viremia :I(period^3) | 0.007    | -0.004 | 0.018 | 0.197   |

Table 9: Linear regression on the evolution of FEV1 over time

#### 3.4. Risk factors for mortality

During the study period 14 out of the 69 patients died. In the univariate Cox model (Table 10), CMV replication was not associated with a higher mortality (HR 0.75, p=0.62). The presence of CLAD was not significantly associated with a higher mortality (HR 2.814, p=0.085). Kaplan-Meier curve of patient survival according to the presence of CLAD is shown in Figure 6.

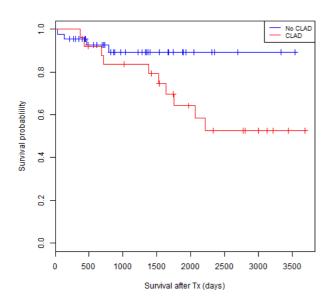


Figure 6: Survival after lung transplantation according to the presence of CLAD

Univariate Cox model of the risk factors for mortality are shown in Table 10.

| Exposition      | HR    | HR C  | p-value |       |
|-----------------|-------|-------|---------|-------|
| CLAD            | 2.814 | 0.866 | 9.148   | 0.085 |
| CMV replication | 0.758 | 0.250 | 2.299   | 0.625 |
| Duration of CMV | 0.991 | 0.953 | 1.031   | 0.667 |
| replication     |       |       |         |       |
| CMV disease     | 1.219 | 0.272 | 5.471   | 0.796 |
| Valganciclovir  | 1.087 | 0.964 | 1.225   | 0.174 |

| Sex (male)          | 0.433 | 0.135 | 1.385 | 0.158 |  |
|---------------------|-------|-------|-------|-------|--|
| Age at Tx           | 1.010 | 0.980 | 1.060 | 0.346 |  |
| CMV serostatus      |       |       |       |       |  |
| D-/R- (ref.)        |       |       |       |       |  |
| D-/R+               | 1.302 | 0.254 | 6.663 | 0.751 |  |
| D+/R-               | 1.733 | 0.413 | 7.271 | 0.452 |  |
| D+/R+               | 0.874 | 0.191 | 3.994 | 0.863 |  |
| Acute rejection     | 0.958 | 0.337 | 2.728 | 0.937 |  |
| Bacterial pneumonia | 1.443 | 0.458 | 4.545 | 0.531 |  |
| Fungal infection    | 0.436 | 0.057 | 3.336 | 0.424 |  |
|                     |       |       |       |       |  |

Table 10: univariate Cox model of the risk factors for mortality

#### 4. DISCUSSION

In our study, we aimed to assess the impact of late-onset CMV replication on allograft and patient outcomes in a single-center cohort of lung transplant recipients. We found that asymptomatic CMV replication occurring after the discontinuation of antiviral prophylaxis (i.e late-onset CMV replication) was not associated with a significant decrease in lung function and survival. The incidence of CLAD was similar in patients that developed CMV replication and there was no correlation between duration of CMV replication and the onset of CLAD. Moreover the decline of FEV1 was not affected by the development of CMV replication and there was no significant decrease of patient survival in patients with CMV replication. Finally, our results showed a decrease of patient survival in the CLAD group, which has already been widely accepted in the literature as the main cause of death in lung transplant recipients (17).

These data suggest that asymptomatic CMV replication occurring after the discontinuation of antiviral prophylaxis does not have a major clinical impact in our population, as lung function and survival were not altered in patients with or without replication. However, the influence of CMV replication on allograft outcomes is still debated. It appears that the administration of antiviral drugs may have an impact on reducing the indirect effects of CMV, as most of the studies that found a relationship between CMV and allograft dysfunction were performed before the introduction of universal antiviral prophylaxis protocols. Another issue is whether detection of CMV in blood. Some studies observed an association between CMV replication in the lung (CMV detected in BAL samples) and the occurrence of CLAD (48), which was not confirmed by another study (49). Because the CMV surveillance protocol was done more intensively in blood than in the BAL in our study, we decided to analyze only the presence of CMV in blood as the primary

endpoint. Our results showing a rather benign effect of CMV replication in lung transplant recipients may be useful for the management of these patients in the routine practice, as it might be not necessary to systematically monitor for CMV replication in asymptomatic patients.

In addition to CMV disease, bacterial pneumonia and respiratory virus infections (52) have been associated with development of CLAD in the literature. In our results, bacterial pneumonia episodes were the only risk factor identified by our analysis that was associated with a higher incidence of CLAD, which suggests an association between infectious episodes and the development of allograft dysfunction. Although this has previously been reported, a clear mechanism of the association between bacterial infection and allograft dysfunction is not fully understood yet (53). Overall, these data suggest an important interaction between mechanisms of host defense against infection and rejection. The balance between on one hand too high immunosuppressive state and development of rejection is a subtle adjustment, which need to be more explored to improve lung transplant recipients care.

Our study has some limitations. First, this is a single-center study with the inherent biases of a retrospective study. Moreover, the modest sample size limited the strength of the analysis, especially regarding the multivariate analysis. However, we did not observe any trend towards a deleterious effect of CMV replication or CMV disease in terms of allograft or patient survival; so that we do not think that a larger sample size would have changed the results of our study. The main strength of our study is the relatively long-term follow-up, with a median follow-up duration of 3.67 years. Moreover, the immunosuppressive regimens, antiviral prophylaxis protocols and the management of infection were homogenous during all the study period, which have potentially diminished the bias associated with a different transplant care over time. CMV detection by PCR was hence performed in a single laboratory, which is relevant since CMV detection has a poor inter-center agreement (54).

#### **5. CONCLUSION**

Asymptomatic CMV replication in the blood did not affect the lung function or survival of lung transplant recipients receiving anti-CMV universal prophylaxis. These results suggest that universal prophylaxis given to patients according to their serostatus is protective regarding CMV as a risk factor for the onset of CLAD.

# 6. REFERENCES

1. SWT\_de\_Jahresbericht\_2014.pdf [Internet]. [cité 10 oct 2015]. Disponible sur: http://www.swisstransplant.org/fileadmin/user\_upload/Infos\_und\_Material/Statistiken/Jahresb ericht/SWT\_de\_Jahresbericht\_2014.pdf

2. The Registry of the International Society for Heart and Lung Transplantation: Thirtyfirst Adult Lung and Heart–Lung Transplant Report—2014; Focus Theme: Retransplantation [Internet]. [cité 17 oct 2015]. Disponible sur:

https://crypto.unil.ch/science/article/pii/,DanaInfo=www.sciencedirect.com+S1053249814012 613

3. Hayes D, Black SM, Tobias JD, Higgins RS, Whitson BA. Influence of donor and recipient age in lung transplantation. J Heart Lung Transplant. 1 janv 2015;34(1):43-9.

4. Gross CR, Savik K, Bolman I R. Morton, Hertz MI. LOng-term health status and quality of life outcomes of lung transplant recipients. Chest. 1 déc 1995;108(6):1587-93.

5. Garrity ER, Villanueva J, Bhorade SM, Husain AN, Vigneswaran WT. Low rate of acute lung allograft rejection after the use of daclizumab, an interleukin 2 receptor antibody. Transplantation. 27 mars 2001;71(6):773-7.

6. Mullen JC, Oreopoulos A, Lien DC, Bentley MJ, Modry DL, Stewart K, et al. A randomized, controlled trial of daclizumab vs anti-thymocyte globulin induction for lung transplantation. J Heart Lung Transplant Off Publ Int Soc Heart Transplant. mai 2007;26(5):504-10.

7. Rhen T, Cidlowski JA. Antiinflammatory action of glucocorticoids--new mechanisms for old drugs. N Engl J Med. 20 oct 2005;353(16):1711-23.

8. Valapour M, Skeans MA, Heubner BM, Smith JM, Hertz MI, Edwards LB, et al. OPTN/SRTR 2013 Annual Data Report: lung. Am J Transplant Off J Am Soc Transplant Am Soc Transpl Surg. janv 2015;15 Suppl 2:1-28.

9. Keenan RJ, Konishi H, Kawai A, Paradis IL, Nunley DR, Iacono AT, et al. Clinical trial of tacrolimus versus cyclosporine in lung transplantation. Ann Thorac Surg. sept 1995;60(3):580-4; discussion 584-5.

10. Speich R, Schneider S, Hofer M, Irani S, Vogt P, Weder W, et al. Mycophenolate mofetil reduces alveolar inflammation, acute rejection and graft loss due to bronchiolitis obliterans syndrome after lung transplantation. Pulm Pharmacol Ther. oct 2010;23(5):445-9.

11. McNeil K, Glanville AR, Wahlers T, Knoop C, Speich R, Mamelok RD, et al. Comparison of mycophenolate mofetil and azathioprine for prevention of bronchiolitis obliterans syndrome in de novo lung transplant recipients. Transplantation. 15 avr 2006;81(7):998-1003.

12. Martinu T, Pavlisko EN, Chen D-F, Palmer SM. Acute allograft rejection: cellular and humoral processes. Clin Chest Med. juin 2011;32(2):295-310.

13. De Vito Dabbs A, Hoffman LA, Iacono AT, Zullo TG, McCurry KR, Dauber JH. Are symptom reports useful for differentiating between acute rejection and pulmonary infection after lung transplantation? Heart Lung J Crit Care. déc 2004;33(6):372-80.

14. Stewart S, Fishbein MC, Snell GI, Berry GJ, Boehler A, Burke MM, et al. Revision of the 1996 working formulation for the standardization of nomenclature in the diagnosis of lung rejection. J Heart Lung Transplant Off Publ Int Soc Heart Transplant. déc 2007;26(12):1229-42.

15. Budding K, van de Graaf EA, Otten HG. Humoral immunity and complement effector mechanisms after lung transplantation. Transpl Immunol [Internet]. [cité 26 oct 2014]; Disponible sur: http://www.sciencedirect.com/science/article/pii/S096632741400077X

16. Verleden GM, Raghu G, Meyer KC, Glanville AR, Corris P. A new classification system for chronic lung allograft dysfunction. J Heart Lung Transplant. 1 févr 2014;33(2):127-33.

17. Meyer KC, Raghu G, Verleden GM, Corris PA, Aurora P, Wilson KC, et al. An international ISHLT/ATS/ERS clinical practice guideline: diagnosis and management of bronchiolitis obliterans syndrome. Eur Respir J. déc 2014;44(6):1479-503.

18. Christie JD, Edwards LB, Aurora P, Dobbels F, Kirk R, Rahmel AO, et al. The Registry of the International Society for Heart and Lung Transplantation: Twenty-sixth Official Adult Lung and Heart-Lung Transplantation Report-2009. J Heart Lung Transplant Off Publ Int Soc Heart Transplant. oct 2009;28(10):1031-49.

 Nakajima T, Palchevsky V, Perkins DL, Belperio JA, Finn PW. Lung transplantation: infection, inflammation, and the microbiome. Semin Immunopathol. mars 2011;33(2):135-56.
 Weigt S, Wallace W, Derhovanessian A, Saggar R, Saggar R, Lynch J, et al. Chronic Allograft Rejection: Epidemiology, Diagnosis, Pathogenesis, and Treatment. Semin Respir Crit Care Med. avr 2010;31(02):189-207.

21. Burton CM, Iversen M, Carlsen J, Mortensen J, Andersen CB, Steinbrüchel D, et al. Acute cellular rejection is a risk factor for bronchiolitis obliterans syndrome independent of post-transplant baseline FEV1. J Heart Lung Transplant Off Publ Int Soc Heart Transplant. sept 2009;28(9):888-93.

22. Chalermskulrat W, Neuringer IP, Schmitz JL, Catellier DJ, Gurka MJ, Randell SH, et al. Human leukocyte antigen mismatches predispose to the severity of bronchiolitis obliterans syndrome after lung transplantation. Chest. juin 2003;123(6):1825-31.

23. Glanville AR, Aboyoun CL, Havryk A, Plit M, Rainer S, Malouf MA. Severity of lymphocytic bronchiolitis predicts long-term outcome after lung transplantation. Am J Respir Crit Care Med. 1 mai 2008;177(9):1033-40.

24. Khalifah AP, Hachem RR, Chakinala MM, Schechtman KB, Patterson GA, Schuster DP, et al. Respiratory Viral Infections Are a Distinct Risk for Bronchiolitis Obliterans Syndrome and Death. Am J Respir Crit Care Med. 15 juill 2004;170(2):181-7.

25. Vos R, Vanaudenaerde BM, Geudens N, Dupont LJ, Van Raemdonck DE, Verleden GM. Pseudomonal airway colonisation: risk factor for bronchiolitis obliterans syndrome after lung transplantation? Eur Respir J. mai 2008;31(5):1037-45.

26. Kapila A, Baz MA, Valentine VG, Bhorade SM. Reliability of diagnostic criteria for bronchiolitis obliterans syndrome after lung transplantation: A survey. J Heart Lung Transplant. 1 janv 2015;34(1):65-74.

27. Vos R, Vanaudenaerde BM, De Vleeschauwer SI, Willems-Widyastuti A, Scheers H, Van Raemdonck DE, et al. Circulating and intrapulmonary C-reactive protein: a predictor of bronchiolitis obliterans syndrome and pulmonary allograft outcome. J Heart Lung Transplant Off Publ Int Soc Heart Transplant. août 2009;28(8):799-807.

28. Li JJ, Sanders RL, McAdam KP, Hales CA, Thompson BT, Gelfand JA, et al. Impact of C-reactive protein (CRP) on surfactant function. J Trauma. déc 1989;29(12):1690-7.

29. Ruttens D, Verleden SE, Vandermeulen E, Bellon H, Vanaudenaerde BM, Somers J, et al. Prophylactic Azithromycin Therapy After Lung Transplantation: Post hoc Analysis of a Randomized Controlled Trial. Am J Transplant Off J Am Soc Transplant Am Soc Transpl Surg. 4 août 2015;

30. Corris PA, Ryan VA, Small T, Lordan J, Fisher AJ, Meachery G, et al. A randomised controlled trial of azithromycin therapy in bronchiolitis obliterans syndrome (BOS) post lung transplantation. Thorax. mai 2015;70(5):442-50.

31. Horvath J, Dummer S, Loyd J, Walker B, Merrill WH, Frist WH. Infection in the transplanted and native lung after single lung transplantation. Chest. sept 1993;104(3):681-5.
32. Impact of Antiviral Preventive Strategies on the Incidence and Outcomes of

Cytomegalovirus Disease in Solid Organ Transplant Recipients - Manuel - 2013 - American Journal of Transplantation - Wiley Online Library [Internet]. [cité 8 oct 2015]. Disponible sur: http://onlinelibrary.wiley.com/doi/10.1111/ajt.12388/full

33. New Developments in the Management of Cytomegalovirus Infection after Solid Organ Transplantation - Springer [Internet]. [cité 11 oct 2015]. Disponible sur: https://crypto.unil.ch/article/,DanaInfo=link.springer.com+10.2165%2F10898540-000000000-00000

34. Freeman J R. B. The 'Indirect' Effects of Cytomegalovirus Infection. Am J Transplant. 1 nov 2009;9(11):2453-8.

35. Leung Ki E-L, Venetz J-P, Meylan P, Lamoth F, Ruiz J, Pascual M. Cytomegalovirus infection and new-onset post-transplant diabetes mellitus. Clin Transplant. avr 2008;22(2):245-9.

36. Humar A, Snydman D, AST Infectious Diseases Community of Practice. Cytomegalovirus in solid organ transplant recipients. Am J Transplant Off J Am Soc Transplant Am Soc Transpl Surg. déc 2009;9 Suppl 4:S78-86.

37. Reyburn HT, Mandelboim O, Valés-Gómez M, Davis DM, Pazmany L, Strominger JL. The class I MHC homologue of human cytomegalovirus inhibits attack by natural killer cells. Nature. 3 avr 1997;386(6624):514-7.

38. Fletcher JM, Prentice HG, Grundy JE. Natural killer cell lysis of cytomegalovirus (CMV)-infected cells correlates with virally induced changes in cell surface lymphocyte function-associated antigen-3 (LFA-3) expression and not with the CMV-induced down-regulation of cell surface class I HLA. J Immunol Baltim Md 1950. 1 sept 1998;161(5):2365-74.

39. Atalay R, Zimmermann A, Wagner M, Borst E, Benz C, Messerle M, et al. Identification and Expression of Human Cytomegalovirus Transcription Units Coding for Two Distinct Fcγ Receptor Homologs. J Virol. sept 2002;76(17):8596-608.

40. Zamora MR. Cytomegalovirus and Lung Transplantation. Am J Transplant. 1 août 2004;4(8):1219-26.

41. Beck S, Barrell BG. Human cytomegalovirus encodes a glycoprotein homologous to MHC class-I antigens. Nature. 21 janv 1988;331(6153):269-72.

42. Manuel O, Wójtowicz A, Bibert S, Mueller NJ, van Delden C, Hirsch HH, et al. Influence of IFNL3/4 polymorphisms on the incidence of cytomegalovirus infection after solid-organ transplantation. J Infect Dis. 9 oct 2014;

43. Mitsani D, Nguyen MH, Girnita DM, Spichty K, Kwak EJ, Silveira FP, et al. A polymorphism linked to elevated levels of interferon- $\gamma$  is associated with an increased risk of cytomegalovirus disease among Caucasian lung transplant recipients at a single center. J Heart Lung Transplant Off Publ Int Soc Heart Transplant. mai 2011;30(5):523-9.

44. Kotton CN, Kumar D, Caliendo AM, Asberg A, Chou S, Danziger-Isakov L, et al. Updated international consensus guidelines on the management of cytomegalovirus in solid-organ transplantation. Transplantation. 27 août 2013;96(4):333-60.

45. Johansson I, Mårtensson G, Nyström U, Nasic S, Andersson R. Lower incidence of CMV infection and acute rejections with valganciclovir prophylaxis in lung transplant recipients. BMC Infect Dis. 2013;13:582.

46. Stern M, Hirsch H, Cusini A, van Delden C, Manuel O, Meylan P, et al. Cytomegalovirus serology and replication remain associated with solid organ graft rejection and graft loss in the era of prophylactic treatment. Transplantation. 15 nov 2014;98(9):1013-8.

47. Harvala H, Stewart C, Muller K, Burns S, Marson L, MacGilchrist A, et al. High risk of cytomegalovirus infection following solid organ transplantation despite prophylactic therapy. J Med Virol. mai 2013;85(5):893-8.

48. Paraskeva M, Bailey M, Levvey BJ, Griffiths AP, Kotsimbos TC, Williams TP, et al. Cytomegalovirus replication within the lung allograft is associated with bronchiolitis obliterans syndrome. Am J Transplant Off J Am Soc Transplant Am Soc Transpl Surg. oct

2011;11(10):2190-6.

49. Manuel O, Kumar D, Moussa G, Chen MH, Pilewski J, McCurry KR, et al. Lack of association between beta-herpesvirus infection and bronchiolitis obliterans syndrome in lung transplant recipients in the era of antiviral prophylaxis. Transplantation. 15 mars 2009;87(5):719-25.

50. Santos CAQ, Brennan DC, Yusen RD, Olsen MA. Incidence, Risk Factors and Outcomes of Delayed-onset Cytomegalovirus Disease in a Large Retrospective Cohort of Lung Transplant Recipients. Transplantation. août 2015;99(8):1658-66.

51. Muheim C, Vogel G, Seydoux C, Gillet M, Mosimann F, Von Segesser L, et al. Determinants of protracted cytomegalovirus infection in solid-organ transplant patients. Transplantation. 27 juill 2002;74(2):226-36.

52. Fisher CE, Preiksaitis CM, Lease ED, Edelman J, Kirby KA, Leisenring WM, et al. Symptomatic Respiratory Virus Infection and Chronic Lung Allograft Dysfunction. Clin Infect Dis [Internet]. 12 nov 2015 [cité 29 nov 2015]; Disponible sur: http://cid.oxfordjournals.org/lookup/doi/10.1093/cid/civ871

53. Martin-Gandul C, Mueller NJ, Pascual M, Manuel O. The Impact of Infection on Chronic Allograft Dysfunction and Allograft Survival After Solid Organ Transplantation. Am J Transplant Off J Am Soc Transplant Am Soc Transpl Surg. déc 2015;15(12):3024-40.

54. Hayden RT, Preiksaitis J, Tong Y, Pang X, Sun Y, Tang L, et al. Commutability of the First World Health Organization International Standard for Human Cytomegalovirus. J Clin Microbiol. oct 2015;53(10):3325-33.