

1 **Taphonomic bias in exceptionally preserved biotas**

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14 **ABSTRACT**

15 Exceptionally preserved fossil biotas provide crucial data on early animal evolution.
16 Fossil anatomy allows for reconstruction of the animal stem lineages, informing the stepwise
17 **process of crown group character acquisition**. However, a confounding factor to these
18 evolutionary analyses is information loss during fossil formation. Here we identify that the
19 Ordovician Fezouata Shale has a clear taphonomic difference when compared to the
20 Cambrian Burgess Shale and Chengjiang Biota. In the Fezouata **Shale**, soft cellular structures
21 are most commonly associated with partially mineralized and sclerotized tissues, which may
22 be protecting the soft tissue. **Also, entirely soft non-cuticularized organisms are absent from**
23 **the Fezouata Shale**. Conversely, the Cambrian sites commonly preserve entirely soft cellular
24 bodies and a higher diversity of tissue types per genus. **The Burgess and Chengjiang biotas**
25 **are remarkably similar, preserving near identical proportions of average tissue types per**
26 **genus**. However, the Burgess shale has almost double the proportion of genera that are
27 entirely soft **as compared to** the Chengjiang Biota, indicating that the classic Burgess Shale
28 **was** the acme for soft tissue preservation. Constraining these biases aids the differentiation of
29 evolutionary and taphonomic absences, which is vital to incorporating anatomical data into a
30 coherent framework of character acquisition during the earliest evolution of animals.

31

32 **1. INTRODUCTION**

33 Exceptionally preserved biotas have revolutionized our understanding of animal
34 origins and evolution owing to the preservation in these deposits of soft-bodied and lightly
35 sclerotized organisms, which under normal circumstances have little to no fossilization
36 potential (Butterfield, 1995). Burgess Shale-type (BST) preservation deposits including the
37 Burgess Shale (Wuliuan, Miaolingian; ~505 Ma, Canada) and the Chengjiang Biota (Stage 3,
38 Cambrian Series 2; ~530 Ma, China) are particularly famous *Lagerstätten*, yielding hundreds

39 of exceptionally preserved Cambrian taxa (Fig. 1a-c) critical to our understanding of the
40 earliest metazoan-dominated communities and evolutionary events such as the Cambrian
41 Explosion (Daley et al., 2018). The youngest of these deposits, the Fezouata **Shale**, is the only
42 Ordovician (Tremadocian; ~479-478 Ma, Morocco) *Lagerstätte* to yield a diverse
43 exceptionally preserved fauna (Fig. 1d-f). With over 185 taxa of marine invertebrates (Van
44 Roy et al., 2015a) recovered from specific intervals in the Zagora area (Lefebvre et al., 2018;
45 Saleh et al., 2018, 2019), this formation offers new insights into the diversification of
46 metazoans, at a key interval between the Cambrian Explosion and the Ordovician Radiation
47 (Van Roy et al., 2010, 2015b; Lefebvre et al., 2019). **Despite being anatomically and**
48 **biologically informative, even these spectacular fossil localities inevitably have taphonomic**
49 **biases, because no fossil site can ever be a perfect replication of all the anatomical and**
50 **ecological information of a living community (Butterfield, 2003; Brasier et al., 2010; Landing**
51 **et al., 2018). Gathering “complete” data is impossible even in studies on modern living**
52 **communities. It is therefore essential to understand what factors may be affecting the fossil**
53 **preservation at a community level in order to properly reconstruct ancient ecosystems and**
54 **biodiversity fluctuations over geological time.**

55 The aim of this study is to examine the taphonomic signal of these deposits, allowing a solid
56 understanding of the preservation bias at play in each locality. For this reason, a taphonomic
57 classification of all eumetazoan genera from the Fezouata **Shale** (N= 178) was established,
58 and compared with the preservation of genera from the Burgess Shale (N=103) and the
59 Chengjiang Biota (N=133) based on the presence / absence of different types of anatomical
60 structures: (A) biomineralized skeletons, (B) sclerotized parts (**i.e. possessing an organically**
61 **strengthened part or organ**) (C) soft with an unsclerotized cuticle (**i.e. a non-cellular outer**
62 **body surface that is either collagenous or formed by polymerized polysaccharides**), (D) soft

63 cellular outer layer defining at least a part of the body (e.g. tentacles of hyoliths), and (E) soft
64 internal cellular organ/tissue (e.g. digestive or nervous systems) (Fig.1).

65

66 2. MATERIAL AND METHODS

67 In order to define the preservation pattern in all three exceptionally preserved biotas,
68 the various possible co-occurrences of characters A (biomineralized), B (sclerotized), C
69 (unsclerotized, cuticularized), D (cellular body walls), and E (internal tissues) were tallied
70 (e.g. AB, AC, CDE, and ABCDE) (Tab. 1). To avoid any overlap between categories, the data
71 were analyzed on a five-fold Venn diagram per site. In order to see if there is any difference
72 between sites, the total number of genera having just one character regardless of its nature
73 (e.g. A, or B, or C, or D, or E) was plotted against the number of genera that have pairs (e.g.
74 AB), threes (e.g. ABC) or fours (e.g. ABCD) for all exceptionally preserved biotas (Fig. 2).
75 Afterward, the average number of tissue types per genus, as derived from the dataset, was
76 calculated by adding the probability of the occurrence of all classes of structures A, B, C, D,
77 and E (Tab. 2). In order to constrain the categories causing the biggest variations in
78 preservation between sites, plots were made to show the proportion of paired and triple
79 categories in localities (Fig. 3).

80 The association of soft internal organs (E) with other structures, in all three localities
81 was also investigated. For this, the probabilities of discovering two classes of structures
82 together having already found one of them were calculated (Tab. 3). For example, $p(E|A)$ is
83 the probability of E occurring if A has occurred. The reverse conditional approach was also
84 made and the probability of finding A given that E has been found $p(A|E)$ was also calculated
85 (Tab. 3). Then, the likelihood of producing the distribution of combinations of structures
86 found in the Burgess Shale and the Chengjiang Biota assuming that the Fezouata Shale has

87 the “true” preservation regime was investigated using the following parametrized binomial
88 $P(x \geq n) | Bi(n, p)$:

$$P(x) = \frac{\binom{n}{x} p^x q^{n-x}}{\sum_{k=x}^n \binom{n}{k} p^k q^{n-k}}$$

89 In this equation, $p = p(E|A)$ for the Fezouata Shale, $q = 1-p$, n is the number of genera
90 preserving an A in the Burgess Shale or the Chengjinag Biota, and x is the number of desired
91 success which is, in this case, at least the actual number n of genera preserving both A and E
92 in the Burgess Shale/Chengjiang Biota. All calculated probabilities are added up and the
93 probability $P(x \geq n) | Bi(n, p)$, of producing the actual Burgess Shale/Chengjinag Biota AE
94 category, considering that the Fezouata Shale regime is “true”, is then obtained (Tab. 4). This
95 was then performed for other tissues combinations (i.e. BE , CE , and DE) (Tab. 4). This
96 approach was then extended to the assumption that the Burgess Shale preservation
97 distribution is “true” and finally assuming that the Chengjiang Biota preservation distribution
98 is the “true” preservation model (Tab. 5).

99 Finally, the probability of finding organisms with only soft cellular tissues (both
100 internal and external to the exclusion of everything else with A' for instance indicating the set
101 that is defined as not containing and members of A) $p(A' \cap B' \cap C' \cap D \cap E | E)$ for all three
102 *Lagerstätten* was calculated.

103

104 3. RESULTS

105 All three *Lagerstätten* preserve numerous biomineralized skeletons (A), sclerotized
106 parts (B), unsclerotized, soft cuticular parts (C), and internal soft parts (E) (Tab. 1). However,
107 genera having cellular body walls defining the entire body (i.e. D , DE), with or without
108 internal organs (E) are absent in the Fezouata Shale. In comparison the Chengjiang Biota (9
109 genera) and the Burgess Shale (13 genera) have a considerable number of entirely soft
110 organisms preserved (Tab. 1). Further, numerous biomineralized and sclerotized genera in the

111 Burgess Shale and the Chengjiang Biota preserve external soft tissues defining a part of the
112 body (i.e. AD, BD, BDE, ACDE) (Tab. 1). These genera are absent from the Fezouata **Shale**,
113 with the exception of two specimens of aculiferan molluscs (both, however, densely covered
114 by sclerites). The Burgess Shale and the Chengjiang Biota preserve almost twice as many
115 tissues per genus as the Fezouata **Shale** (Fig. 2), with the mean number of tissue types per
116 genus in the Cambrian sites being 2.2 (Burgess = 2.206; Chengjiang = 2.185) whilst it is
117 1.316 for the Fezouata Shale (Tab. 2). The overall distribution of tissue frequency by genus
118 are similar for the Burgess Shale and the Chengjiang Biota, with mean and variance
119 suggesting they are drawn from comparable if not identical populations (variance Burgess
120 Shale = 0.026; Chengjiang Biota = 0.030; $t = -0.45$, $p(\text{same mean}) = 0.6532$; $F = 1.154$,
121 $p(\text{same variance}) = 0.454$). However, the distribution for the Fezouata **Shale** is very different
122 (variance = 0.08034), with both t and F -tests reporting significance for the mean and variance
123 respectively when compared to Burgess Shale ($t = 29.53$, $p(\text{same mean}) = 1.035 \times 10^{-87}$; $F =$
124 3.0685 , $p(\text{same variance}) = 3.195 \times 10^{-9}$) and the Chengjiang Biota ($t = 32.34$, $p(\text{same mean}) =$
125 3.414×10^{-101} ; $F = 2.5591$, $p(\text{same variance}) = 1.718 \times 10^{-8}$).

126 The three studied localities show a dominance of both BCE and ACE categories (Fig.
127 3). This is at least partly linked to the high number of arthropods found at all localities, with
128 their external anatomy often consisting of ventral unsclerotized cuticle (C) and a reinforced
129 dorsal area consisting of a biomineralized exoskeleton (A) or sclerotized cuticle (B), found in
130 conjunction with internal soft parts (E). However, when the preservation of two tissue types
131 occurs in the Fezouata **Shale**, it consists mostly of the association of biomineralized skeletons
132 and internal soft tissues (AE is 9 of the 21 pairs that consist of the possible sets AB, AC, AD,
133 AE, BC, BD, BE, CD, CE, DE), sclerotized tissue and internal soft tissue (7 of the 21 pairs),
134 and biominerals and sclerotized tissue (3 of 21 pairs). All other tissue associations are rare or
135 absent. In the Burgess Shale, the dominant association is between cellular soft bodied tissues

136 and internal organs (13 of 36 pairs), with sclerotized and cuticularized tissues also commonly
137 associated (7 of 36 pairs). In the Chengjiang Biota, the dominant association is between
138 sclerotized and cuticularized tissues (16 of 57 pairs), with additional common associations
139 between cuticularized tissues and internal organs (12 of 57 pairs), cellular soft bodied tissues
140 and internal organs (9 of 57 pairs), and biominerals and sclerotized tissues (8 of 57 pairs)
141 (Fig. 3). The probabilities of finding internal soft tissues in a given fossil genus, in co-
142 occurrence with any of the other types of structures, show that the distribution of tissues in the
143 Burgess Shale and the Chengjiang Biota are much more similar to each other (Tab. 3) and are
144 significantly different from the Fezouata Shale (Tab. 4). In the Fezouata Shale, only a small
145 proportion of all biomineralized genera also preserve internal organs ($p(E|A) = 0.162$) (Tab.
146 3), but of the genera that do have internal organs the majority are associated with biominerals
147 ($(A|E) = 0.667$) (Tab. 3). This means that although a biomineral does not guarantee the
148 preservation of internal anatomies, it could still be seen as a very helpful pre-requisite in the
149 Fezouata Shale. Conversely, biominerals in paleoenvironments such as the Burgess Shale and
150 the Chengjiang Biota do not seem to have any role in soft tissue preservation ($p(A|E) = 0.183$
151 and $p(A|E) = 0.273$ for the Burgess Shale and the Chengjiang Biota respectively, which are
152 not significantly different to chance association (Tab. 3). The result of probabilistic modelling
153 (Tab. 4) shows that the distributions of tissue associations found at the Fezouata Shale cannot
154 be generated by randomly sampling a biota with a similar composition to that of either the
155 Chengjiang Biota or the Burgess Shale, and in all possible soft tissue combinations the
156 Fezouata Shale is statistically significantly different to both of the Cambrian biotas studied
157 (Tab. 4). Finally, it is worth noting that the absence of entirely soft bodied organisms at the
158 Fezouata Shale is not just a striking observation, but it is also statistically significant from the
159 proportions found at the Cambrian sites. The absence of entirely soft bodied organisms at the
160 Fezouata Shale cannot be generated by randomly sampling a population like that found in the

161 Cambrian sites with any confidence (with p-values of 0.00137 and 0.03819 for Burgess Shale
162 and Chengjiang Biota models respectively). Therefore, the Burgess Shale ($p(D \cap E|E) =$
163 0.2167) and the Chengjiang Biota ($p(D \cap E|E) = 0.113$) both show significantly higher
164 probabilities of recovering entirely soft bodied genera. The preservation of entirely soft
165 bodied genera is also different between the Chengjiang Biota and the Burgess Shale (Tab. 3),
166 with the higher incidence being found in the Burgess Shale. This difference is significant and
167 could not be generated by chance or subsampling (Tab. 5).

168

169 4. DISCUSSION

170 Soft part preservation in the Fezouata Shale is strikingly different from the
171 preservation in the Chengjiang Biota and the Burgess Shale. **This difference in the**
172 **occurrences of soft tissues cannot result from a collection bias, because all three localities**
173 **were subjected to collecting efforts that actively focused on finding and sampling fossils with**
174 **labile soft part.** Instead, the observed pattern of preservation suggests that the presence of
175 non-cellular layers covering internal anatomies in the Fezouata Shale was essential for
176 exceptional preservation, unlike at the Burgess Shale and Chengjiang Biota. The near
177 complete absence of preserved external soft tissues is possibly related to them being less
178 decay-resistant than mineralized, sclerotized or even cuticularized structures. Under most
179 circumstances, even unsclerotized soft cuticle is more decay resistant than cellular tissue,
180 because cuticular structures are not subject to autolysis, and the composition of complex
181 polymerized polysaccharides means cuticle is more difficult to break down than cellular
182 tissues (Briggs and Kear, 1993). **The decay-resistance of complex biopolymers found in the**
183 **cuticle was also recently invoked to explain the rare but selective preservation of cuticularized**
184 **organisms in coarse clastic sediments (MacGabhann et al., 2019).**

185 In the Fezouata Shale, there was a pathway of preservation in place that systematically
186 failed to preserve (i) almost all soft-bodied organisms lacking a cuticular cover in particular,
187 and (ii) external soft cellular tissues in general. **In this deposit, dead individuals experienced**
188 **harsh decay prior to their preservation owing to a relative burial tardiness (Saleh et al., 2018)**
189 **in comparison with the Burgess Shale and the Chengjiang Biota in which fossils were killed**
190 **and preserved directly during an obrution event (Gaines, 2014). This decay may also have**
191 **been retarded by berthierine, a mineral that can slow down microbial activity through the**
192 **oxidative damage of bacterial cells (McMahon et al., 2016; Anderson et al., 2018; Saleh et al.,**
193 **2019). Therefore,** in contrast to the Burgess Shale and the Chengjiang Biota, the external
194 conditions at the Fezouata Shale were generally less permissive for the preservation of
195 external soft tissues. However, resistant skeletal parts and cuticular external surfaces created
196 isolated environments within the carcasses that maintained a chemical equilibrium conducive
197 to the preservation of internal organs.

198 The systematic taphonomic bias described here for the Fezouata Shale has
199 implications for understanding the original faunal community assemblage, specifically in
200 regard to the proportions of genera preserved in the fossil record. The systematic removal of
201 all soft-bodied organisms, lacking a non-cellular external envelope (cuticle), and external
202 cellular soft tissues leads to an underestimation of the original diversity at the Cambro-
203 Ordovician transition and distorts faunal composition to a greater extent than in the Burgess
204 Shale or the Chengjiang Biota. **Many animal groups could have lived in the Fezouata Shale**
205 **environment but left little to no trace behind, such as chordates (e.g. *Pikaia*, *Metaspriggina*).**
206 A corollary of this finding is that it is now possible to differentiate between ecological and
207 taphonomic absences of numerous genera. For example, the absence of priapulids such as
208 *Ottoia* in the Fezouata Shale (Van Roy et al., 2015a) is likely a real aspect of the fauna, since
209 these cuticle-bearing soft-bodied animals would not have been affected by the same

210 taphonomic bias responsible for the removal of the majority of soft-bodied genera lacking a
211 cuticle.

212 Now that a source of systematic taphonomic bias operating in the Fezouata Shale has
213 been identified (Fig. 4), and most importantly, compared to the biases in play in the Burgess
214 Shale and the Chengjiang Biota (Fig. 4), it can be accounted for in future paleoecological and
215 evolutionary analyses. This will facilitate more accurate comparisons of faunal community
216 compositions between these biotas in particular, and when comparing exceptionally preserved
217 faunas in general, as similar restrictive mechanisms are likely active to a varying extent at
218 other localities.

219

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284

285 **TABLES AND FIGURES**

286 Table 1. Number of genera in different categories in all exceptionally preserved biotas.

287 Table 2. Proportion of each type of tissue in all categories combined in the Fezouata Shale,
288 the Burgess Shale and the Chengjiang Biota. The probability of preserving cuticularized and
289 cellular tissues, in addition to the number of tissue per genus in the Fezouata Shale are lower
290 than in the Chengjiang Biota and the Burgess Shale.

291 Table 3. Probabilities of finding internal soft tissues in a fossil given that another tissue was
292 found and vice versa. The obtained numbers for the Burgess Shale and the Chengjiang Biota
293 are more similar to each other than to the Fezouata Shale.

294 Table 4. Probabilities of reproducing patterns of preservation of the Burgess Shale and the
295 Chengjiang Biota assuming that the Fezouata Shale preservation regime is true. All
296 probabilities are smaller than 0.05 showing that the preservation regime in the Fezouata Shale
297 is different from both the Chengjiang Biota and the Burgess Shale.

298 Table 5. A: Probabilities of reproducing patterns of preservation of the Burgess Shale
299 assuming that the Chengjiang biota preservation regime is true. B: Probabilities of
300 reproducing patterns of preservation of the Chengjiang Biota assuming that the Burgess Shale
301 preservation regime is true. **Some tissue associations are not reproducible in both models (i.e.**
302 **marked as “No” in the “Pass” column), showing that the pattern of preservation between the**
303 **Burgess Shale and the Chengjiang Biota is not exactly the same.**

304 **Figure 1. Fossils from the three studied exceptionally preserved biotas showing examples of**
305 **tissue associations. (a) Burgess Shale *Eldonia* USNM57540b preserving soft cellular body**
306 **walls and internal organs (i.e. DE). (b) *Branchiocaris pretiosa* from the Burgess Shale**
307 **USNM189028nc showing the association of sclerotized and cuticularized parts in addition to**
308 **internal organs (BCE). (c) *Anomalocaris saron* ELRC20001a from the Chengjiang Biota**
309 **belonging as well to the BCE category. (d) Marrellid arthropod from the Fezouata Shale AA-**

310 BIZ31-OI-39 preserving both sclerotized and cuticularized structures (BC). (e) Fezouata
311 Shale stylophoran echinoderm AA.BIZ.15.OI.259 showing the association of biominerals and
312 internal organs (AE). (f) Solutan echinoderm from the Fezouata Shale CASG72938
313 belonging also to the AE category.

314 Figure 2. Differences in proportions of genera (Y axis) between single, paired, triple and
315 quadruple character categories (marked as 1, 2, 3, and 4 on the X axis) between the Fezouata
316 Shale, the Burgess Shale and the Chengjiang Biota. The Fezouata Shale shows a dominance
317 of genera preserving only one tissue when compared to the Burgess Shale and Chengjiang
318 Biota.

319 Figure 3. Pie charts showing the differences in paired and triple character categories between
320 the Fezouata Shale, the Burgess Shale, and the Chengjiang Biota.

321 Figure 4. Preservation differences between exceptionally preserved biotas and one non-
322 *Lagerstätte* (i.e. preservation of only mineralized genera). The Chengjiang biota and the
323 Burgess Shale preserve more tissue-types than the Fezouata Shale in which soft tissues in
324 direct contact with sea water are not preserved.

Highlights :

- Exceptionally preserved biotas are not subject to the same fossilization bias
- Fossilization bias is constrained in three iconic Burgess Shale type biotas using a new statistical method
- Cambrian localities preserved with high fidelity snapshots of early animal life

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14 **ABSTRACT**

15 Exceptionally preserved fossil biotas provide crucial data on early animal evolution.
16 Fossil anatomy allows for reconstruction of the animal stem lineages, informing the stepwise
17 process of crown group character acquisition. However, a confounding factor to these
18 evolutionary analyses is information loss during fossil formation. Here we identify that the
19 Ordovician Fezouata Shale has a clear taphonomic difference when compared to the
20 Cambrian Burgess Shale and Chengjiang Biota. In the Fezouata Shale, soft cellular structures
21 are most commonly associated with partially mineralized and sclerotized tissues, which may
22 be protecting the soft tissue. Also, entirely soft non-cuticularized organisms are absent from
23 the Fezouata Shale. Conversely, the Cambrian sites commonly preserve entirely soft cellular
24 bodies and a higher diversity of tissue types per genus. The Burgess and Chengjiang biotas
25 are remarkably similar, preserving near identical proportions of average tissue types per
26 genus. However, the Burgess shale has almost double the proportion of genera that are
27 entirely soft as compared to the Chengjiang Biota, indicating that the classic Burgess Shale
28 was the acme for soft tissue preservation. Constraining these biases aids the differentiation of
29 evolutionary and taphonomic absences, which is vital to incorporating anatomical data into a
30 coherent framework of character acquisition during the earliest evolution of animals.

31

32 **1. INTRODUCTION**

33 Exceptionally preserved biotas have revolutionized our understanding of animal
34 origins and evolution owing to the preservation in these deposits of soft-bodied and lightly
35 sclerotized organisms, which under normal circumstances have little to no fossilization
36 potential (Butterfield, 1995). Burgess Shale-type (BST) preservation deposits including the
37 Burgess Shale (Wuliuan, Miaolingian; ~505 Ma, Canada) and the Chengjiang Biota (Stage 3,
38 Cambrian Series 2; ~530 Ma, China) are particularly famous *Lagerstätten*, yielding hundreds

39 of exceptionally preserved Cambrian taxa (Fig. 1a-c) critical to our understanding of the
40 earliest metazoan-dominated communities and evolutionary events such as the Cambrian
41 Explosion (Daley et al., 2018). The youngest of these deposits, the Fezouata Shale, is the only
42 Ordovician (Tremadocian; ~479-478 Ma, Morocco) *Lagerstätte* to yield a diverse
43 exceptionally preserved fauna (Fig. 1d-f). With over 185 taxa of marine invertebrates (Van
44 Roy et al., 2015a) recovered from specific intervals in the Zagora area (Lefebvre et al., 2018;
45 Saleh et al., 2018, 2019), this formation offers new insights into the diversification of
46 metazoans, at a key interval between the Cambrian Explosion and the Ordovician Radiation
47 (Van Roy et al., 2010, 2015b; Lefebvre et al., 2019). Despite being anatomically and
48 biologically informative, even these spectacular fossil localities inevitably have taphonomic
49 biases, because no fossil site can ever be a perfect replication of all the anatomical and
50 ecological information of a living community (Butterfield, 2003; Brasier et al., 2010; Landing
51 et al., 2018). Gathering “complete” data is impossible even in studies on modern living
52 communities. It is therefore essential to understand what factors may be affecting the fossil
53 preservation at a community level in order to properly reconstruct ancient ecosystems and
54 biodiversity fluctuations over geological time.

55 The aim of this study is to examine the taphonomic signal of these deposits, allowing a solid
56 understanding of the preservation bias at play in each locality. For this reason, a taphonomic
57 classification of all eumetazoan genera from the Fezouata Shale (N= 178) was established,
58 and compared with the preservation of genera from the Burgess Shale (N=103) and the
59 Chengjiang Biota (N=133) based on the presence / absence of different types of anatomical
60 structures: (A) biomineralized skeletons, (B) sclerotized parts (i.e. possessing an organically
61 strengthened part or organ) (C) soft with an unsclerotized cuticle (i.e. a non-cellular outer
62 body surface that is either collagenous or formed by polymerized polysaccharides), (D) soft

63 cellular outer layer defining at least a part of the body (e.g. tentacles of hyoliths), and (E) soft
64 internal cellular organ/tissue (e.g. digestive or nervous systems) (Fig.1).

65

66 **2. MATERIAL AND METHODS**

67 In order to define the preservation pattern in all three exceptionally preserved biotas,
68 the various possible co-occurrences of characters A (biomineralized), B (sclerotized), C
69 (unsclerotized, cuticularized), D (cellular body walls), and E (internal tissues) were tallied
70 (e.g. AB, AC, CDE, and ABCDE) (Tab. 1). To avoid any overlap between categories, the data
71 were analyzed on a five-fold Venn diagram per site. In order to see if there is any difference
72 between sites, the total number of genera having just one character regardless of its nature
73 (e.g. A, or B, or C, or D, or E) was plotted against the number of genera that have pairs (e.g.
74 AB), threes (e.g. ABC) or fours (e.g. ABCD) for all exceptionally preserved biotas (Fig. 2).
75 Afterward, the average number of tissue types per genus, as derived from the dataset, was
76 calculated by adding the probability of the occurrence of all classes of structures A, B, C, D,
77 and E (Tab. 2). In order to constrain the categories causing the biggest variations in
78 preservation between sites, plots were made to show the proportion of paired and triple
79 categories in localities (Fig. 3).

80 The association of soft internal organs (E) with other structures, in all three localities
81 was also investigated. For this, the probabilities of discovering two classes of structures
82 together having already found one of them were calculated (Tab. 3). For example, $p(E|A)$ is
83 the probability of E occurring if A has occurred. The reverse conditional approach was also
84 made and the probability of finding A given that E has been found $p(A|E)$ was also calculated
85 (Tab. 3). Then, the likelihood of producing the distribution of combinations of structures
86 found in the Burgess Shale and the Chengjiang Biota assuming that the Fezouata Shale has

87 the “true” preservation regime was investigated using the following parametrized binomial
88 $P(x \geq n) | \text{Bi}(n, p)$:

$$P(x) = \binom{n}{x} p^x q^{n-x} = \frac{n!}{(n-x)! x!} p^x q^{n-x}$$

89 In this equation, $p = p(E|A)$ for the Fezouata Shale, $q = 1-p$, n is the number of genera
90 preserving an A in the Burgess Shale or the Chengjinag Biota, and x is the number of desired
91 success which is, in this case, at least the actual number n of genera preserving both A and E
92 in the Burgess Shale/Chengjiang Biota. All calculated probabilities are added up and the
93 probability $P(x \geq n) | \text{Bi}(n, p)$, of producing the actual Burgess Shale/Chengjinag Biota AE
94 category, considering that the Fezouata Shale regime is “true”, is then obtained (Tab. 4). This
95 was then performed for other tissues combinations (i.e. BE , CE , and DE) (Tab. 4). This
96 approach was then extended to the assumption that the Burgess Shale preservation
97 distribution is “true” and finally assuming that the Chengjiang Biota preservation distribution
98 is the “true” preservation model (Tab. 5).

99 Finally, the probability of finding organisms with only soft cellular tissues (both
100 internal and external to the exclusion of everything else with A' for instance indicating the set
101 that is defined as not containing and members of A) $p(A' \cap B' \cap C' \cap D \cap E | E)$ for all three
102 *Lagerstätten* was calculated.

103

104 3. RESULTS

105 All three *Lagerstätten* preserve numerous biomineralized skeletons (A), sclerotized
106 parts (B), unsclerotized, soft cuticular parts (C), and internal soft parts (E) (Tab. 1). However,
107 genera having cellular body walls defining the entire body (i.e. D , DE), with or without
108 internal organs (E) are absent in the Fezouata Shale. In comparison the Chengjiang Biota (9
109 genera) and the Burgess Shale (13 genera) have a considerable number of entirely soft
110 organisms preserved (Tab. 1). Further, numerous biomineralized and sclerotized genera in the

111 Burgess Shale and the Chengjiang Biota preserve external soft tissues defining a part of the
112 body (i.e. AD, BD, BDE, ACDE) (Tab. 1). These genera are absent from the Fezouata Shale,
113 with the exception of two specimens of aculiferan molluscs (both, however, densely covered
114 by sclerites). The Burgess Shale and the Chengjiang Biota preserve almost twice as many
115 tissues per genus as the Fezouata Shale (Fig. 2), with the mean number of tissue types per
116 genus in the Cambrian sites being 2.2 (Burgess = 2.206; Chengjiang = 2.185) whilst it is
117 1.316 for the Fezouata Shale (Tab. 2). The overall distribution of tissue frequency by genus
118 are similar for the Burgess Shale and the Chengjiang Biota, with mean and variance
119 suggesting they are drawn from comparable if not identical populations (variance Burgess
120 Shale = 0.026; Chengjiang Biota = 0.030; $t = -0.45$, $p(\text{same mean}) = 0.6532$; $F = 1.154$,
121 $p(\text{same variance}) = 0.454$). However, the distribution for the Fezouata Shale is very different
122 (variance = 0.08034), with both t and F -tests reporting significance for the mean and variance
123 respectively when compared to Burgess Shale ($t = 29.53$, $p(\text{same mean}) = 1.035 \times 10^{-87}$; $F =$
124 3.0685 , $p(\text{same variance}) = 3.195 \times 10^{-9}$) and the Chengjiang Biota ($t = 32.34$, $p(\text{same mean}) =$
125 3.414×10^{-101} ; $F = 2.5591$, $p(\text{same variance}) = 1.718 \times 10^{-8}$).

126 The three studied localities show a dominance of both BCE and ACE categories (Fig.
127 3). This is at least partly linked to the high number of arthropods found at all localities, with
128 their external anatomy often consisting of ventral unsclerotized cuticle (C) and a reinforced
129 dorsal area consisting of a biomineralized exoskeleton (A) or sclerotized cuticle (B), found in
130 conjunction with internal soft parts (E). However, when the preservation of two tissue types
131 occurs in the Fezouata Shale, it consists mostly of the association of biomineralized skeletons
132 and internal soft tissues (AE is 9 of the 21 pairs that consist of the possible sets AB, AC, AD,
133 AE, BC, BD, BE, CD, CE, DE), sclerotized tissue and internal soft tissue (7 of the 21 pairs),
134 and biominerals and sclerotized tissue (3 of 21 pairs). All other tissue associations are rare or
135 absent. In the Burgess Shale, the dominant association is between cellular soft bodied tissues

136 and internal organs (13 of 36 pairs), with sclerotized and cuticularized tissues also commonly
137 associated (7 of 36 pairs). In the Chengjiang Biota, the dominant association is between
138 sclerotized and cuticularized tissues (16 of 57 pairs), with additional common associations
139 between cuticularized tissues and internal organs (12 of 57 pairs), cellular soft bodied tissues
140 and internal organs (9 of 57 pairs), and biominerals and sclerotized tissues (8 of 57 pairs)
141 (Fig. 3). The probabilities of finding internal soft tissues in a given fossil genus, in co-
142 occurrence with any of the other types of structures, show that the distribution of tissues in the
143 Burgess Shale and the Chengjiang Biota are much more similar to each other (Tab. 3) and are
144 significantly different from the Fezouata Shale (Tab. 4). In the Fezouata Shale, only a small
145 proportion of all biomineralized genera also preserve internal organs ($p(E|A) = 0.162$) (Tab.
146 3), but of the genera that do have internal organs the majority are associated with biominerals
147 ($(A|E) = 0.667$) (Tab. 3). This means that although a biomineral does not guarantee the
148 preservation of internal anatomies, it could still be seen as a very helpful pre-requisite in the
149 Fezouata Shale. Conversely, biominerals in paleoenvironments such as the Burgess Shale and
150 the Chengjiang Biota do not seem to have any role in soft tissue preservation ($p(A|E) = 0.183$
151 and $p(A|E) = 0.273$ for the Burgess Shale and the Chengjiang Biota respectively, which are
152 not significantly different to chance association (Tab. 3). The result of probabilistic modelling
153 (Tab. 4) shows that the distributions of tissue associations found at the Fezouata Shale cannot
154 be generated by randomly sampling a biota with a similar composition to that of either the
155 Chengjiang Biota or the Burgess Shale, and in all possible soft tissue combinations the
156 Fezouata Shale is statistically significantly different to both of the Cambrian biotas studied
157 (Tab. 4). Finally, it is worth noting that the absence of entirely soft bodied organisms at the
158 Fezouata Shale is not just a striking observation, but it is also statistically significant from the
159 proportions found at the Cambrian sites. The absence of entirely soft bodied organisms at the
160 Fezouata Shale cannot be generated by randomly sampling a population like that found in the

161 Cambrian sites with any confidence (with p-values of 0.00137 and 0.03819 for Burgess Shale
162 and Chengjiang Biota models respectively). Therefore, the Burgess Shale ($p(D \cap E|E) =$
163 0.2167) and the Chengjiang Biota ($p(D \cap E|E) = 0.113$) both show significantly higher
164 probabilities of recovering entirely soft bodied genera. The preservation of entirely soft
165 bodied genera is also different between the Chengjiang Biota and the Burgess Shale (Tab. 3),
166 with the higher incidence being found in the Burgess Shale. This difference is significant and
167 could not be generated by chance or subsampling (Tab. 5).

168

169 **4. DISCUSSION**

170 Soft part preservation in the Fezouata Shale is strikingly different from the
171 preservation in the Chengjiang Biota and the Burgess Shale. This difference in the
172 occurrences of soft tissues cannot result from a collection bias, because all three localities
173 were subjected to collecting efforts that actively focused on finding and sampling fossils with
174 labile soft part. Instead, the observed pattern of preservation suggests that the presence of
175 non-cellular layers covering internal anatomies in the Fezouata Shale was essential for
176 exceptional preservation, unlike at the Burgess Shale and Chengjiang Biota. The near
177 complete absence of preserved external soft tissues is possibly related to them being less
178 decay-resistant than mineralized, sclerotized or even cuticularized structures. Under most
179 circumstances, even unsclerotized soft cuticle is more decay resistant than cellular tissue,
180 because cuticular structures are not subject to autolysis, and the composition of complex
181 polymerized polysaccharides means cuticle is more difficult to break down than cellular
182 tissues (Briggs and Kear, 1993). The decay-resistance of complex biopolymers found in the
183 cuticle was also recently invoked to explain the rare but selective preservation of cuticularized
184 organisms in coarse clastic sediments (MacGabhann et al., 2019).

185 In the Fezouata Shale, there was a pathway of preservation in place that systematically
186 failed to preserve (i) almost all soft-bodied organisms lacking a cuticular cover in particular,
187 and (ii) external soft cellular tissues in general. In this deposit, dead individuals experienced
188 harsh decay prior to their preservation owing to a relative burial tardiness (Saleh et al., 2018)
189 in comparison with the Burgess Shale and the Chengjiang Biota in which fossils were killed
190 and preserved directly during an obrution event (Gaines, 2014). This decay may also have
191 been retarded by berthierine, a mineral that can slow down microbial activity through the
192 oxidative damage of bacterial cells (McMahon et al., 2016; Anderson et al., 2018; Saleh et al.,
193 2019). Therefore, in contrast to the Burgess Shale and the Chengjiang Biota, the external
194 conditions at the Fezouata Shale were generally less permissive for the preservation of
195 external soft tissues. However, resistant skeletal parts and cuticular external surfaces created
196 isolated environments within the carcasses that maintained a chemical equilibrium conducive
197 to the preservation of internal organs.

198 The systematic taphonomic bias described here for the Fezouata Shale has
199 implications for understanding the original faunal community assemblage, specifically in
200 regard to the proportions of genera preserved in the fossil record. The systematic removal of
201 all soft-bodied organisms, lacking a non-cellular external envelope (cuticle), and external
202 cellular soft tissues leads to an underestimation of the original diversity at the Cambro-
203 Ordovician transition and distorts faunal composition to a greater extent than in the Burgess
204 Shale or the Chengjiang Biota. Many animal groups could have lived in the Fezouata Shale
205 environment but left little to no trace behind, such as chordates (e.g. *Pikaia*, *Metaspriggina*).
206 A corollary of this finding is that it is now possible to differentiate between ecological and
207 taphonomic absences of numerous genera. For example, the absence of priapulids such as
208 *Ottoia* in the Fezouata Shale (Van Roy et al., 2015a) is likely a real aspect of the fauna, since
209 these cuticle-bearing soft-bodied animals would not have been affected by the same

210 taphonomic bias responsible for the removal of the majority of soft-bodied genera lacking a
211 cuticle.

212 Now that a source of systematic taphonomic bias operating in the Fezouata Shale has
213 been identified (Fig. 4), and most importantly, compared to the biases in play in the Burgess
214 Shale and the Chengjiang Biota (Fig. 4), it can be accounted for in future paleoecological and
215 evolutionary analyses. This will facilitate more accurate comparisons of faunal community
216 compositions between these biotas in particular, and when comparing exceptionally preserved
217 faunas in general, as similar restrictive mechanisms are likely active to a varying extent at
218 other localities.

219

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272

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284

285 **TABLES AND FIGURES**

286 Table 1. Number of genera in different categories in all exceptionally preserved biotas.

287 Table 2. Proportion of each type of tissue in all categories combined in the Fezouata Shale,
288 the Burgess Shale and the Chengjiang Biota. The probability of preserving cuticularized and
289 cellular tissues, in addition to the number of tissue per genus in the Fezouata Shale are lower
290 than in the Chengjiang Biota and the Burgess Shale.

291 Table 3. Probabilities of finding internal soft tissues in a fossil given that another tissue was
292 found and vice versa. The obtained numbers for the Burgess Shale and the Chengjiang Biota
293 are more similar to each other than to the Fezouata Shale.

294 Table 4. Probabilities of reproducing patterns of preservation of the Burgess Shale and the
295 Chengjiang Biota assuming that the Fezouata Shale preservation regime is true. All
296 probabilities are smaller than 0.05 showing that the preservation regime in the Fezouata Shale
297 is different from both the Chengjiang Biota and the Burgess Shale.

298 Table 5. A: Probabilities of reproducing patterns of preservation of the Burgess Shale
299 assuming that the Chengjiang biota preservation regime is true. B: Probabilities of
300 reproducing patterns of preservation of the Chengjiang Biota assuming that the Burgess Shale
301 preservation regime is true. Some tissue associations are not reproducible in both models (i.e.
302 marked as “No” in the “Pass” column), showing that the pattern of preservation between the
303 Burgess Shale and the Chengjiang Biota is not exactly the same.

304 Figure 1. Fossils from the three studied exceptionally preserved biotas showing examples of
305 tissue associations. (a) Burgess Shale *Eldonia* USNM57540b preserving soft cellular body
306 walls and internal organs (i.e. DE). (b) *Branchiocaris pretiosa* from the Burgess Shale
307 USNM189028nc showing the association of sclerotized and cuticularized parts in addition to
308 internal organs (BCE). (c) *Anomalocaris saron* ELRC20001a from the Chengjiang Biota
309 belonging as well to the BCE category. (d) Marrellid arthropod from the Fezouata Shale AA-

310 BIZ31-OI-39 preserving both sclerotized and cuticularized structures (BC). (e) Fezouata
311 Shale stylophoran echinoderm AA.BIZ.15.OI.259 showing the association of biominerals and
312 internal organs (AE). (f) Solutan echinoderm from the Fezouata Shale CASG72938
313 belonging also to the AE category.

314 Figure 2. Differences in proportions of genera (Y axis) between single, paired, triple and
315 quadruple character categories (marked as 1, 2, 3, and 4 on the X axis) between the Fezouata
316 Shale, the Burgess Shale and the Chengjiang Biota. The Fezouata Shale shows a dominance
317 of genera preserving only one tissue when compared to the Burgess Shale and Chengjiang
318 Biota.

319 Figure 3. Pie charts showing the differences in paired and triple character categories between
320 the Fezouata Shale, the Burgess Shale, and the Chengjiang Biota.

321 Figure 4. Preservation differences between exceptionally preserved biotas and one non-
322 *Lagerstätte* (i.e. preservation of only mineralized genera). The Chengjiang biota and the
323 Burgess Shale preserve more tissue-types than the Fezouata Shale in which soft tissues in
324 direct contact with sea water are not preserved.

Figure 1

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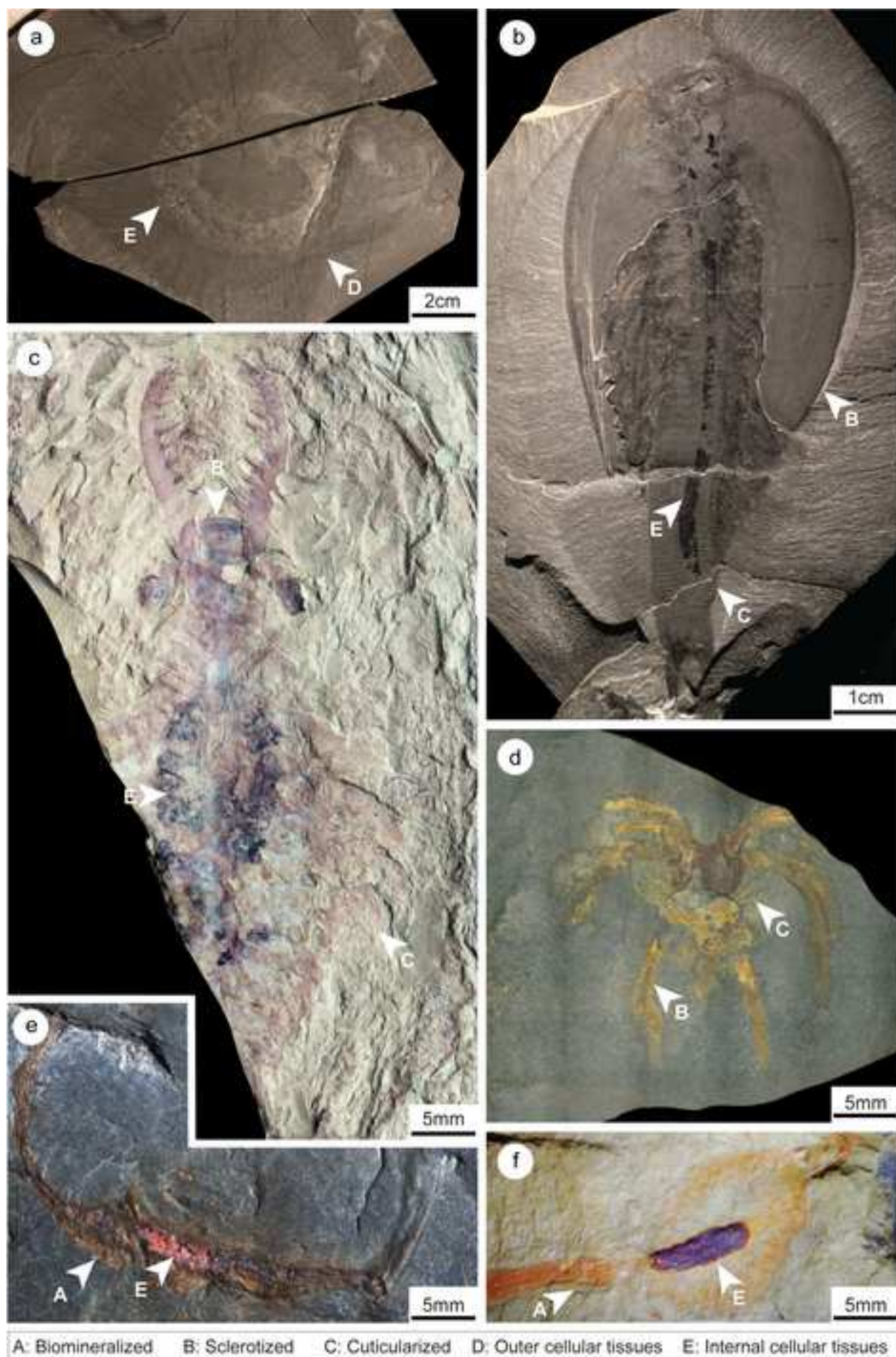


Figure 2

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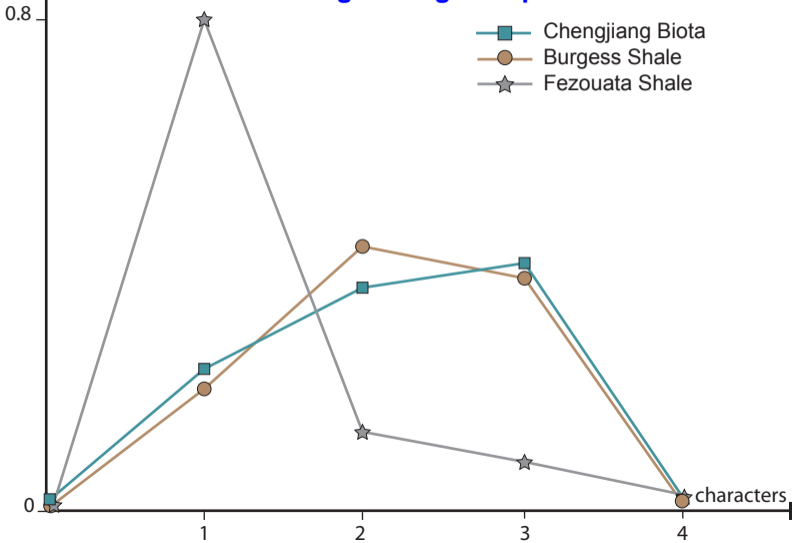
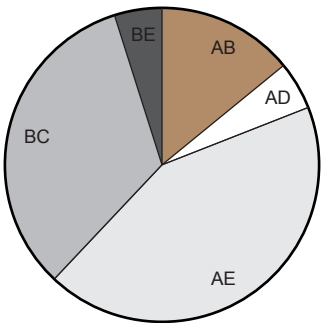
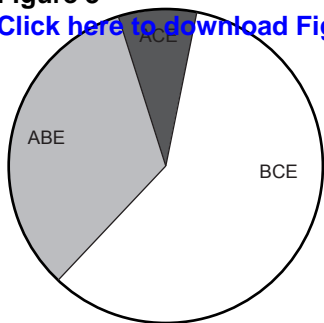
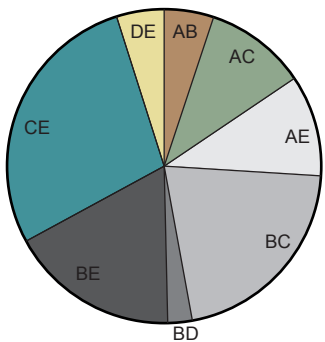
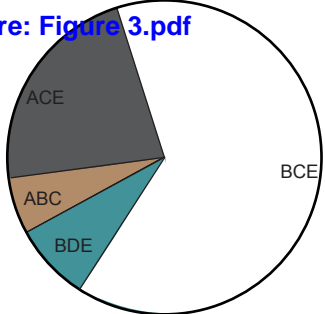


Figure 3

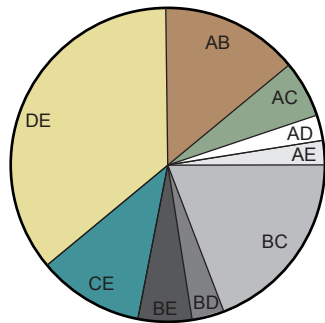
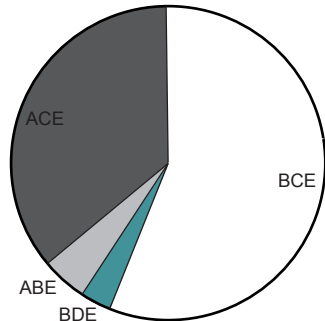
[Click here to download Figure: Figure 3.pdf](#)



Fezouata Shale



Chengjiang Biota

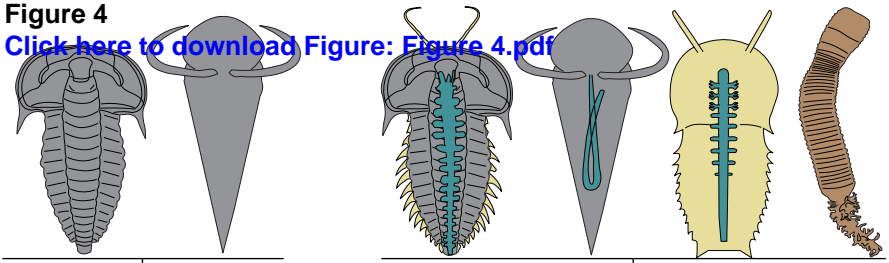


Burgess Shale

A: Biomineralized B: Sclerotized C: Cuticularized D: Outer cellular tissues E: Internal cellular tissues

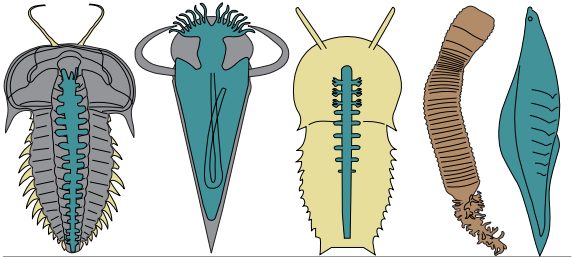
Figure 4

[Click here to download Figure: Figure 4.pdf](#)



Non-Lagerstätte

Fezouata Shale



Burgess Shale & Chengjiang biota

- Cellular tissues
- Cuticularized
- Sclerotized
- Biomineralized

Table 1

[Click here to download Table: Table 1.docx](#)

	Fezouata Shale	Burgess Shale	Chengjiang Biota
A	90	15	4
B	41	7	9
C	3	0	6
D	0	1	4
E	1	0	0
AB	3	5	8
AC	0	2	2
AD	1	1	0
AE	9	1	0
BC	7	7	16
BD	0	1	4
BE	1	2	6
CD	0	0	0
CE	0	4	12
DE	0	13	9
ABC	0	2	0
ABD	0	0	0
ABE	5	0	2
ACD	0	0	0
ACE	1	8	19
ADE	0	0	0
BCD	0	0	0
BCE	7	28	28
BDE	0	2	1
CDE	0	0	0
ABCD	0	0	0
ABCE	3	1	1
ACDE	0	1	0
ACDE	0	0	0
BCDE	0	0	0
ABCDE	0	0	0

Table 1

Table 2[Click here to download Table: Table 2.docx](#)

	Fezouata Shale N(total)=173	Burgess Shale N(total)=101	Chengjiang Biota N(total)=133
A	N(A)=112 p(A)=0.647	N(A)= 36 p(A)=0.356	N(A)= 36 p(A) = 0.270
B	N(B)=67 p(B) = 0.387	N(B)=55 p(B)=0.544	N(B)=75 p(B)=0.563
C	N(C)=21 p(C) = 0.121	N(C)=53 p(C)=0.524	N(C)=84 p(C)=0.631
D	N(D)=1 p(D)=0.005	N(D)=19 p(D)=0.188	N(D)=18 p(D)=0.135
E	N(E)=27 p(E)=0.156	N(E)=60 p(E)=0.594	N(E)=78 p(E)=0.586
Total = tissue/genus	1.316	2.206	2.185

Table 2

Table 3[Click here to download Table: Table 3.docx](#)

	Fezouata Shale	Burgess Shale	Chengjiang Biota
$p(E A)$	0.162	0.306	0.611
$p(E B)$	0.239	0.607	0.507
$p(E C)$	0.524	0.789	0.714
$p(E D)$	0	0.842	0.556
$p(A E)$	0.667	0.183	0.278
$p(B E)$	0.593	0.567	0.481
$p(C E)$	0.407	0.683	0.759
$p(D E)$	0	0.267	0.127

Table 3

Table 4[Click here to download Table: Table 4.docx](#)

	Burgess Shale	Chengjiang Biota
$p(E A)$	$P(X \geq 11) \text{Bi}(36, 0.162)$ $= 0.0235$	$P(X \geq 22) \text{Bi}(36, 0.162)$ < 0.000001
$p(E B)$	$P(X \geq 34) \text{Bi}(56, 0.239)$ < 0.000001	$P(X \geq 38) \text{Bi}(75, 0.239)$ < 0.000001
$p(E C)$	$P(X \geq 41) \text{Bi}(52, 0.524)$ $= 0.0000738$	$P(X \geq 60) \text{Bi}(84, 0.524)$ $= 0.000291$
$p(E D)$	0	0

Table 4

Table 5[Click here to download Table: Table 5.docx](#)

	A: Burgess given a Chengjiang Biota model	B: Chengjiang given the Burgess Shale model	Pass?
$p(E A)$	$P(X \leq 11) \text{Bi}(36, 0.611) = 0.000201$	$P(X \geq 22) \text{Bi}(36, 0.306) = 0.000149$	No
$p(E B)$	$P(X \geq 34) \text{Bi}(56, 0.507) = 0.0857$	$P(X \leq 38) \text{Bi}(75, 0.607) = 0.292$	Yes
$p(E C)$	$P(X \geq 41) \text{Bi}(52, 0.714) = 0.150$	$P(X \leq 60) \text{Bi}(84, 0.789) = 0.0649$	Yes
$p(E D)$	$P(X \geq 16) \text{Bi}(19, 0.556) = 0.00887$	$P(X \leq 10) \text{Bi}(18, 0.842) = 0.000758$	No

Table 5